



REPORT OF PROGRESS 2004 – 2005

**Science Division
August 2005**

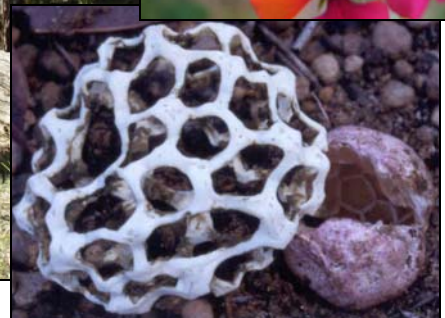


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EXECUTIVE SUMMARY

This document reports the results of FORESTCHECK monitoring in the eastern jarrah forest in Wellington District in 2004-05. Ten monitoring grids were installed in 2004, bringing the total number of FORESTCHECK grids to 37. Previously, grids were established in Donnelly, Wellington and Perth Hills Districts. At the time of writing a further 11 grids have been selected and are in the process of being installed in the Blackwood District and will be monitored in 2005-06. This report, and previous reports, can be viewed on and downloaded from CALM's Naturebase website at <http://www.calm.wa.gov.au/science/science.html>.

The integrated monitoring system used in the FORESTCHECK program has revealed the complex nature of the jarrah forest environment through the variation in species composition and community structure found in various forest types. Each year large numbers of species have been shown to be specific to particular locations and vegetation complexes within the jarrah forest range. The drier eastern jarrah forest monitored in 2004-05 included many species not found in previous years. Wildfires in late December 2004 and January 2005 resulted in a number of grids being burnt. The Perth Hills external control grid in Occidental block in particular was severely burnt, along with a complete set of treatment grids in Bell forest block in the eastern jarrah in Wellington District. The occurrence of these fires will provide the FORESTCHECK program an opportunity to monitor and compare post-fire recovery on the sites.

The FORESTCHECK project continues to collect valuable data and voucher collections for groups of organisms for which we have little knowledge. The invertebrate collection has expanded to include 1271 morphospecies. The list of macrofungi and cryptogams (lichens, mosses and liverworts) includes 482 and 235 species respectively. Many of these species are undescribed and have been collected and recorded for the first time.

During the year, all 37 FORESTCHECK grids were assessed to determine the number, size and age of cut stumps, which will permit the effects of past harvesting on forest structure to be quantified.

Some interesting points that have emerged from the 2004-05 results are:

- Shelterwood and gap release grids were well stocked except for the Stockyard shelterwood, which had poor regeneration
- The Wellington East soils appear to have higher concentrations of nutrients than soils in the Donnelly, Perth Hills and Wellington locations
- The network of snig tracks on the Stockyard shelterwood was extensive, and the possibility of re-using existing snig tracks to limit further ground disturbance should be investigated
- Litter and small wood and twig loads reflected the time since last burn
- Unique species of macrofungi were recorded on the burnt grids in Bell forest block
- Some species of macrofungi are common throughout the jarrah forest (recorded each year in the different FORESTCHECK locations) but many appear to have a restricted distribution
- Species richness of cryptogams was higher on the external control (unlogged) grids

- Fewer species of mosses and liverworts were recorded in the drier eastern jarrah forest compared to locations monitored in previous years
- The eastern jarrah appears to have a remarkably diverse invertebrate fauna (including 306 morphospecies not recorded previously)
- Vascular plant species richness and abundance appeared to be related to time since fire
- There were significantly more native mammals trapped in Godfrey forest block, which is fox baited, compared to Stockyard and Bell forest blocks which are unbaited
- The wet and cold autumn weather had an adverse effect on the success of small mammal and reptile trapping
- Although nocturnal birds were not surveyed, masked owls, southern boobook owls, tawny frogmouths and owl nightjars were all recorded from opportunistic sightings.

Each year, the data collected are entered and stored in a central database. The results in this report represent preliminary observations from basic analyses; however, it is the intention to subject the data collected in the first 5 years to a rigorous, integrated analysis, report and external review. The results will be important in supporting ecologically sustainable forest management in WA.

I thank the FORESTCHECK team for their professionalism, enthusiasm and commitment to the project, and Verna Tunsell and Richard Robinson for compiling and editing this report.

Dr Neil Burrows
Director, Science Division

September 2005

INTRODUCTION

Scope

This report has been compiled from chapters prepared by scientists and technical staff involved in the FORESTCHECK monitoring program. It represents a summary of monitoring activities completed in eastern jarrah forest in the Wellington District during the 2004/2005 financial year.

FORESTCHECK is an integrated monitoring system that has been developed to provide information to forest managers in the southwest of Western Australia about changes and trends in key elements of forest biodiversity associated with a variety of forest management activities. Although the initial focus of FORESTCHECK will be on timber harvesting and silvicultural treatments in jarrah (*Eucalyptus marginata*) forest, the protocol lends itself to monitoring other forest ecosystems, fire (prescribed and wildfire), mining, the effects of forest disturbance for utility corridors (e.g. roads, power transmission lines), and the impacts of recreation uses. (Note, however, that the Forest Products Commission will only fund the part of FORESTCHECK that is specific to its activities).

FORESTCHECK was developed to meet a range of compliance conditions placed on the Forest Management Plan 1994-2003 through Ministerial Conditions and the Codd Report of 1999 and is included as an operational program in the current Forest Management Plan 2004-2013¹. Integrated monitoring is a fundamental component of Ecologically Sustainable Forest Management (ESFM), and is necessary for reporting against the Montreal Process criteria and indicators for ESFM. In addition, monitoring forms the basis for adaptive management, which is recognized as an appropriate strategy for managing under conditions of uncertainty and change.

The Science Division of the Department of Conservation and Land Management has primary responsibility for the implementation of FORESTCHECK. The development of the program took place over 2 years and included input from scientists and managers within the Department of Conservation and Land Management, and from a number of external scientific agencies. The background to this process is described in the FORESTCHECK Concept Plan, and details of methods are provided in the FORESTCHECK Operations Plan. Annual Progress Reports, the Concept Plan and Operations Plan may be viewed on the Department's Naturebase website at <http://www.calm.wa.gov.au/science/science.html>.

Sampling strategy

Between 1995 and 2004 timber harvesting in jarrah forests was undertaken according to Silvicultural Guideline 1/95, which recognizes 3 silvicultural objectives:

- (1) Thinning: to promote growth on retained trees.
- (2) Release of regeneration by gap creation, where existing advance growth is encouraged to develop unimpeded by the removal of competing overstorey.
- (3) Regeneration establishment by shelterwood, where seedlings are encouraged to establish and develop into the lignotuberous ground coppice stage. This is achieved by reducing the competition from the overstorey, but retaining sufficient overstorey to provide a seed source and maintain other forest values until the ground coppice is developed and capable of responding to release.

¹ Conservation Commission of Western Australia (2004). Forest management plan 2004-2013. Conservation Commission of Western Australia. 144pp + maps.

Silvicultural guidelines were revised in conjunction with the preparation of the Forest Management Plan 2004-2013 and are now available as SFM Guideline No. 1(CALM 2004²).

Gap creation and shelterwood treatments are being given priority in the initial stages of FORESTCHECK as these are the most widespread operations and involve the greatest extent of disturbance to the forest. Thinning will also be monitored where the structure of the forest dictates that this treatment is appropriate on a significant scale.

FORESTCHECK sites are being established at a number of locations across the forest, stratified according to recognized ecological gradients of rainfall, evapo-transpiration and soil fertility. Forest ecosystem mapping (Mattiske and Havel 1998³, 2000⁴) provides a systematic basis for stratification of sampling. Allocation of sites also takes account of scheduled future harvesting within the jarrah forest, with priority given to those ecosystems likely to be subject to harvesting on an extensive scale in the next decade.

Each FORESTCHECK site consists of up to 4 sampling grids. Grids will be established in forest subject to the following treatments:

- (1) gap release,
- (2) shelterwood,
- (3) coupe buffer or internal reference forest i.e. temporary exclusion areas (TEAS) between adjacent gaps or shelterwood forest,
- (4) external reference or control forest i.e. not recently harvested, or has had minimal harvesting, and will not be subject to harvesting in the foreseeable future.

The intention is that grids be closely matched in terms of site characteristics (climate, geomorphology, soils, topography, altitude, aspect), pre-harvest forest structure and vegetation attributes in order that differences between grids reflect the effects of harvesting, rather than inherent site differences. Not all treatment types will be found in the one locality and it is expected that external reference forest may have to be located some distance from their harvested counterparts. It may not always be possible to find gap release and shelterwood treatments together, because underlying relationships between rainfall, soil fertility and jarrah lignotuber development influence the broad pattern of silvicultural treatment across the jarrah forest, as have previous silvicultural activities.

Methodology

A range of ecosystem attributes are monitored at each site including:

1. Forest structure and regeneration stocking
2. Foliar and soil nutrients
3. Soil disturbance
4. Coarse woody debris and leaf litter
5. Macrofungi
6. Cryptogams
7. Vascular flora
8. Invertebrate fauna
9. Vertebrate fauna (birds, herpetofauna, and mammals)

² CALM 2004. Silvicultural practice in the jarrah forest. Dept. CALM, SFM guideline No. 1.

³ Mattiske, E.M. and Havel, J.J. 1998. Regional Forest Agreement Vegetation Complexes, Collie, Western Australia [cartographic material – scale 1:250,000]. WA Department of Conservation and Land Management.

⁴ Mattiske, E.M. and Havel, J.J 2000. Vegetation Mapping in the South West of Western Australia. CALM, Perth.

Sampling methodologies for each set of ecosystem attributes are described in the FORESTCHECK Operations Plan, together with examples of protocols for data collection and storage. Monitoring of biodiversity is based on a sampling grid (see Fig. 15 on p.18). The main grid is 100 m x 100 m, with 30 m x 30 m vegetation sample plots at each corner.

Sampling in Wellington District (eastern jarrah) 2004-05.

Ten FORESTCHECK monitoring grids were established and sampled in the eastern jarrah forest of the Wellington District during 2004/05. The location encompassing the sites will be referred to from here on as Wellington East and grids are identified by alphanumeric codes FC28 to FC37. Four grids (FC28-31) were established in Nalyerin and Godfrey forest blocks (Figs 1 & 2), 3 (FC32-34) in Stockyard forest block (Fig. 3) and 3 (FC35-37) in Bell forest block (Fig. 4). All grids were located in areas of forest immediately east of the 800 mm annual rainfall isohyets and meet the definition of eastern jarrah forest outlined in SFM Guideline No 1 (CALM 2004).

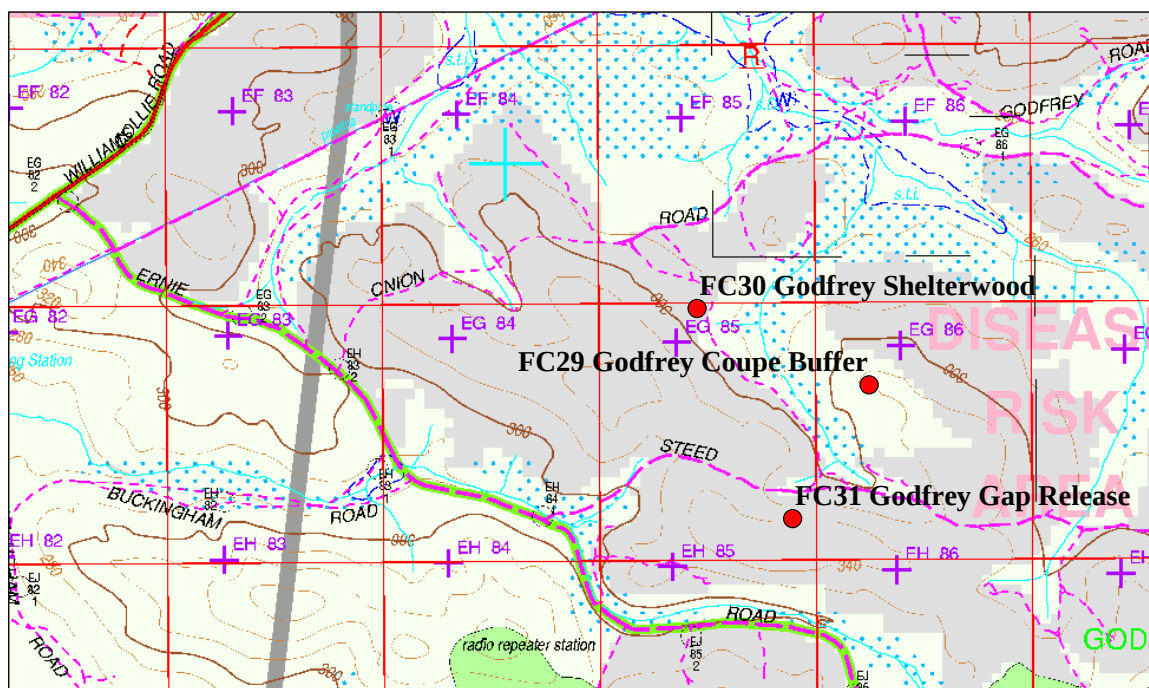


Figure 1. Location of FORESTCHECK sampling grids established in 2004 in Godfrey forest block.

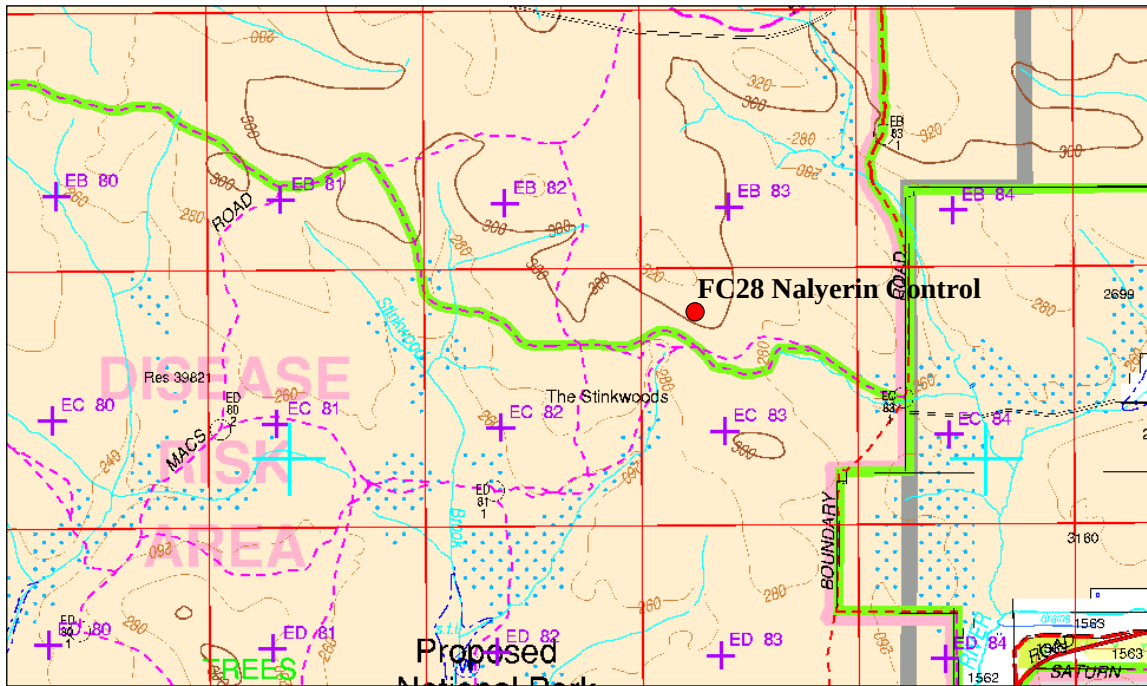


Figure 2. Location of FORESTCHECK sampling grid established in 2004 in Nalyerin forest block.

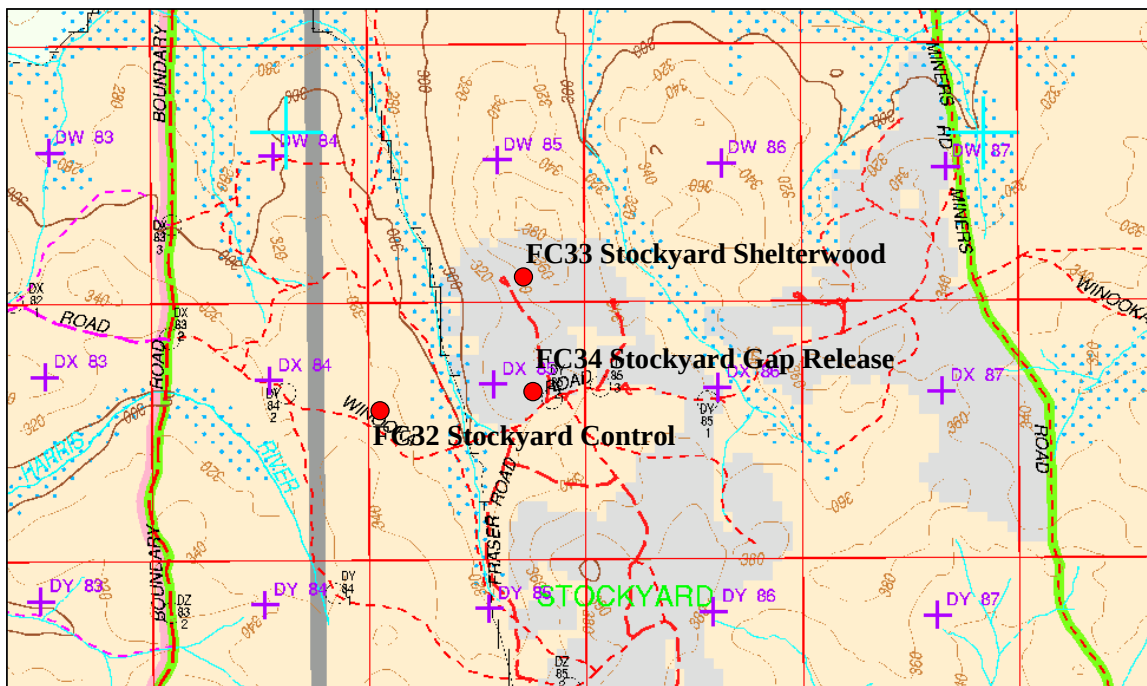


Figure 3. Location of FORESTCHECK sampling grids established in 2004 in Stockyard forest block.

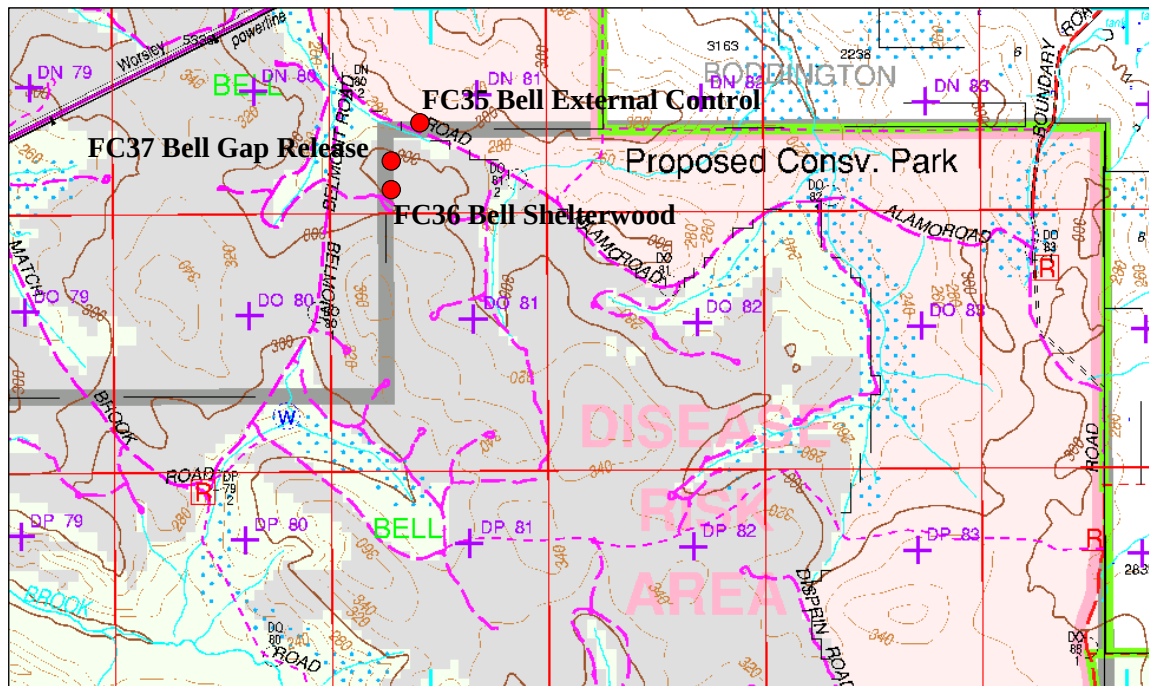


Figure 4. Location of FORESTCHECK sampling grids established in 2004 in Bell forest block.

All grids are located within the Dwellingup 4 vegetation complex of Matiske and Havel (1998). These complexes are found in open forests or woodland jarrah (*E. marginata* ssp. *thalassica*) and marri (*Corymbia calophylla*), on semi-arid lateritic uplands (Table 1).

Table 1 Locations and attributes for each FORESTCHECK grid in Wellington East.

Grid ID	Block	Vegetation Complex	Latitude (S)	Longitude (E)	Elevation (m)	Aspect	Slope (degrees)
FC28	Nalyerin	Dwell. 4	33° 12' 15"	116° 23' 21"	300	SW	-
FC29	Godfrey	Dwell. 4	33° 16' 24"	116° 26' 16"	320	W	2
FC30	Godfrey	Dwell. 4	33° 16' 02"	116° 25' 25"	300	E	1
FC31	Godfrey	Dwell. 4	33° 16' 52"	116° 25' 51"	340	N	3-4
FC32	Stockyard	Dwell. 4	33° 08' 21"	116° 24' 55"	320	NE	-
FC33	Stockyard	Dwell. 4	33° 07' 53"	116° 25' 44"	340	W	3
FC34	Stockyard	Dwell. 4	33° 08' 21"	116° 25' 44"	320	S	3
FC35	Bell	Dwell. 4	32° 59' 40"	116° 21' 35"	300	SW	-
FC36	Bell	Dwell. 4	32° 59' 53"	116° 21' 18"	300	SW	-
FC37	Bell	Dwell. 4	32° 59' 51"	116° 21' 32"	300	NE	-

The 3 external control grids are situated in unlogged forest (but there are a small number of stumps present in the Stockyard grid suggesting a couple of trees were selectively harvested in the past), and have a range of 6-17 years since the time of the last fire (Table 2). The Stockyard external control grid is situated in National Park, the Bell grid in Conservation Park and the Nalyerin grid in the Lane-Poole Reserve. The remaining grids are in forest that was harvested during the period of 1996-2000. All the treatment grids were thus established at sites in stands with comparable management histories.

Table 2. Management history for each FORESTCHECK grid in Wellington East.

Treatment/ Plot	Block	Logged			Burnt		Type of Burn
		Most Recent	Years since	Previous cutting	Most Recent	Years since	
Control							
FC28	Nalyerin	Unlogged			1987-88	17	Fuel reduction
FC32	Stockyard	Uncut		1950s	1998-99	6	Fuel reduction
FC35*	Bell	Unlogged			1997-98	7	Fuel reduction
TEAS							
FC29	Godfrey	Uncut		1940s	2002-03	2	Fuel reduction
Shelterwood							
FC30	Godfrey	2000	5	1930s, 1950s	2002-03	2	Establishment
FC33	Stockyard	1998	7	1950	1998-99	6	Establishment
FC36*	Bell	1996	9		1997-98	7	Establishment
Gap Release							
FC31	Godfrey	2000	5	1950s	2002-03	2	Regeneration release
FC34	Stockyard	1998	7	1950s	1998-99	6	Regeneration release
FC37*	Bell	1996	9		1997-98	7	Regeneration release

*NB. Bell grids were all burnt in a wildfire on 28 December 2004.

In December 2004, about 3 months following establishment, a fire initiated by a lightning strike in Bell forest block resulted in all 3 grids in Bell forest block being burnt (see Figs 12-14). External control and shelterwood treatments were burnt during mild evening conditions and fire intensities were sufficiently low to avoid crown defoliation and stem damage to saplings taller than about 6 m. The Bell gap release treatment was burnt at moderate intensity resulting in overstorey crown scorch and death of some saplings back to ground level. A portion of the monitoring had been completed prior to the fire, including vascular plant and cryptogam surveys, spring invertebrate survey and the spring animal trapping and spotlighting program, however, this meant that a significant proportion of the monitoring program was conducted following the fire. A post fire severity survey was also conducted on the burnt grids in January 2005.

On May 16 2005, a mini tornado hit the coast at Bunbury. The storm travelled inland and passed through the forest north of Collie. Storm damage was clearly evident in the Godfrey gap release grid where many fallen trees and broken branches had affected the stand structure and the amount of CWD on site (see Fig. 8b).

Reference photographs taken of each sampling grid are presented in Figs. 5-14. All photos were taken from peg W2.1 looking towards the centre peg (W2.3) (see Fig. 15), and will allow changes in vegetation structure and condition to be observed in each subsequent photograph. All photographs were taken during plot establishment in October 2004. The additional photos of grids in Bell forest block were taken in January 2005, 10 days following the fire, and the Godfrey gap release on July 7 2005, 6 weeks following the tornado/wind storm.

Godfrey/Nalyerin grids:



Figure 5. FC28 Nalyerin forest block, External Control



Figure 6. FC30 Godfrey forest block, Shelterwood



Figure 7. FC31 Godfrey forest block, Gap Release.
Before (above) and after (below) the wind storm in May 2005.



Figure 8. FC29 Godfrey forest block, Coupe Buffer

Stockyard grids:



Figure 9. FC32 Stockyard forest block, External Control;



Figure 10. FC33 Stockyard forest block, Shelterwood.



Figure 11. FC34 Stockyard forest block, Gap Release.

Bell grids:



Figure 12. FC35 Bell forest block, External Control.
Before (above) and 10 days after (below) being burnt.



Figure 13. FC36 Bell forest block, Shelterwood.
Before (above) and 10 days after (below) being burnt.



Figure 14. FC37 Bell forest block, Gap Release.
Before (above) and 10 days after (below) being burnt.

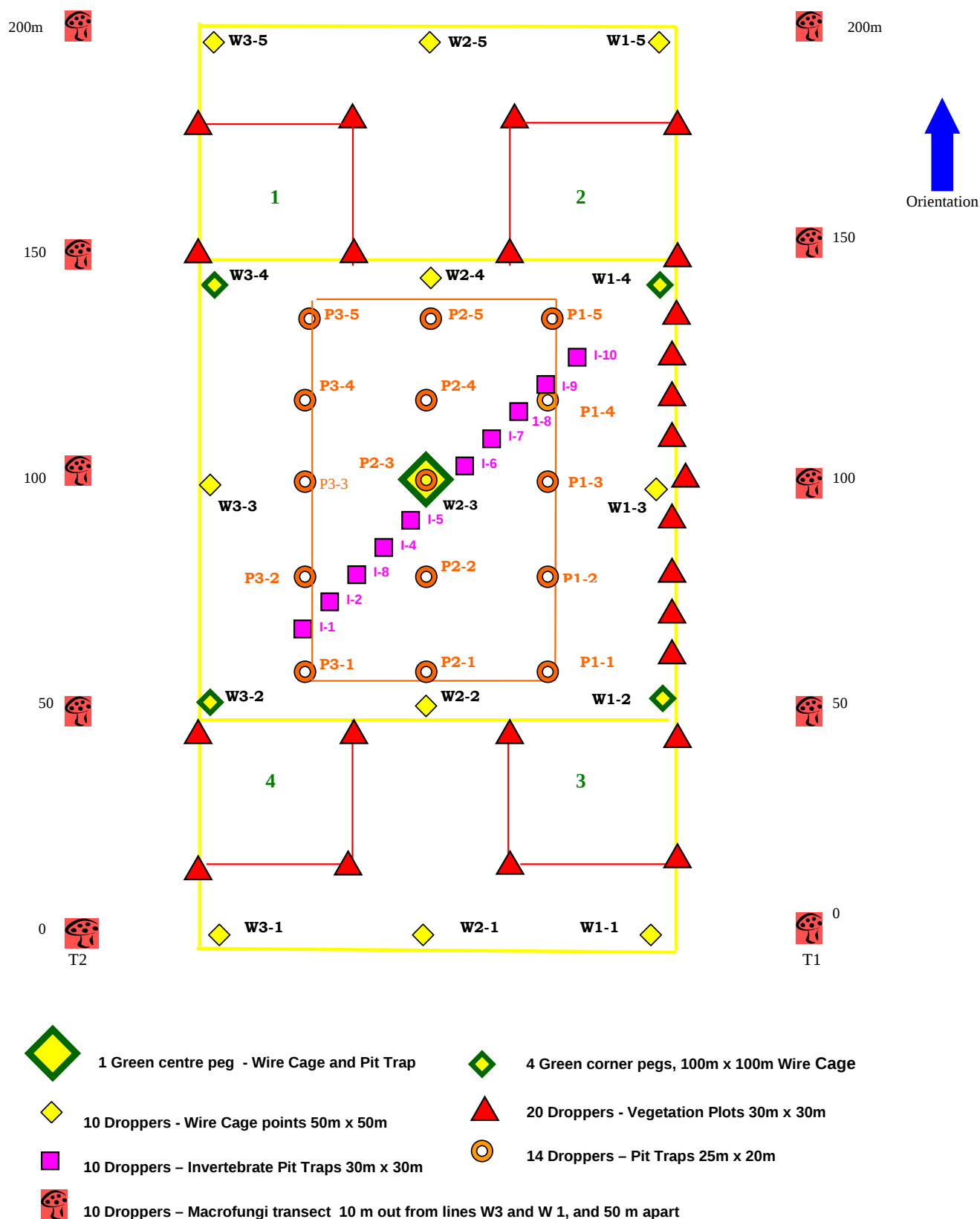


Figure 15. The layout of a FORESTCHECK sampling grid.

The budget and expenditure for the 2004-05 FORESTCHECK program is detailed in Table 4. Funding for the program is provided by the Forest Products Commission and CALM, of which the FPC contributed \$200, 000.

Table 4. Budget for establishment and monitoring of FORESTCHECK sites in Wellington East in 2004/05

Task/Activity	Officer-in-charge	Total Operating
OPERATIONAL		
Grid establishment	McCaw	10 000
Forest structure and regeneration	McCaw	6 000
Soil and foliar nutrients	McCaw	5 000
Soils disturbance	Whitford	10 000
Macrofungi / Litter & CWD	Robinson	7 000
Vascular flora	Ward	5 000
Cryptogams	Cranfield	5 000
Invertebrates	Farr	13 000
Birds (diurnal)	Liddelow	4 000
Birds (nocturnal)	Liddelow	5 000
Fauna (grid trapping)	Liddelow	4 000
Spotlight Road surveys (vertebrate.)	Liddelow	4 000
OTHER		
Administration and overheads	McCaw	63 882
Database management	McCaw	46 796
Directorate		11 321
SUB TOTAL		200 000
SALARY		173 437
TOTAL		373 437

FOREST STRUCTURE AND REGENERATION STOCKING

Lachlan McCaw, Bob Smith and John Neal

Introduction

The adequacy of regeneration following harvesting and silvicultural treatment is one of the core indicators of Ecologically Sustainable Forest Management (ESFM). The current framework of regional level indicators provides for assessment of the area and per cent of harvested area of native forest effectively regenerated (Indicator 2.1.g). This is recognized as a Category A indicator that can be reported upon immediately.

Regeneration outcomes have for a number of years been assessed as a matter of routine on at least a portion of the area of forest subject to harvesting. For uneven-aged forest stands, there is a need to consider the existing structure of the stand and whether sufficient sapling and advance growth is present to re-establish the stand following harvesting. Under the current silvicultural guidelines for jarrah-marri forest the decision as to whether the stand should be cut to gap release or shelterwood is determined by the density of existing lignotuberous advance growth.

Forest managers also require information about the rate of growth and species composition of stands so that future stand conditions can be projected over time. These attributes can affect the potential of forest stands to produce wood and other products, and to achieve ecological outcomes.

Following discussion at the 2004 annual project meeting the sampling methodology for stand structure was extended to include assessment of cut stumps and trees that have been either pushed over or blown down. Collection of this data makes possible a quantification of the basal area removed in each previous cutting cycle, and the effect of stand improvement silvicultural practices on forest structure. All 37 sampling sites established to date as part of the project were assessed for stumps and pushed trees during 2005.

The objectives of this component of FORESTCHECK monitoring are therefore:

- To describe the stand structure, species composition and developmental stage of tree species present at each sampling grid,
- To quantify the basal area removed in past harvesting events, and
- To measure the contribution of mid-storey species to stand structure, density and basal area.

Monitoring

All the grids were assessed in mid March 2005, but the burnt sites in Bell forest block were not assessed for regeneration stocking because seedlings and above ground parts of lignotuberous advance growth had been consumed by the fire.

Sampling techniques were the same as in previous years, apart from the additional assessment of cut stumps and pushed trees. The diameter, height and species of stumps >10 cm diameter on the cut face were recorded in the 800 m² transect used to assess stand structure. The age of cutting event that resulted in the stump was noted (1 = most recent harvest event, 2 = harvested prior to the most recent event and so on for earlier harvests). Trees that had been pushed or blown down were measured for diameter at 1.3 m above nominal ground level, and a judgement was made as to whether they had fallen as a result

of disturbance before during or after harvesting. Regeneration stocking was assessed using the triangular tessellation method at 50 points around the outer perimeter of each grid. Triangular tessellation sampling is the standard technique employed for regeneration surveys undertaken before and after timber harvesting operations. Stand structure was assessed by measurement of all trees taller than 2 m in a transect 200 m long by 4 m wide. In stands cut to gap release and shelterwood treatments, the height and species of regeneration was assessed at 4 locations on each grid to indicate the rate of regrowth. Canopy cover was measured as part of the vascular plant assessment.

Results

Stand structure and species composition

Eucalypt basal areas ranged from 13 to 57 m²/ha in the 3 external control grids, and were comprised predominantly of jarrah (Figure 1, Table 1). Intermediate trees of *Allocasuarina fraseriana* contributed a further 10 m²/ha at Stockyard (FC 32) and 17 m²/ha at Nalyerin (FC28). The external control at Stockyard has been subject to timber harvesting in the past with almost 20 m²/ha of basal area removed prior to the current cutting cycle, probably during the decade of the 1920-29. Previous harvesting had also removed 36 m²/ha of basal area from the coupe buffer at Godfrey forest block (FC29), although the standing basal area of 31 m²/ha at this site was still substantial.

Shelterwood stands had retained basal area of jarrah and marri ranging from 20 to 26 m²/ha. At Godfrey (FC30) and Bell (FC36) the most recent harvesting operation had removed 9-11 m²/ha of jarrah basal area, while at Stockyard (FC33) the reduction in basal area within the sample transects resulted from push down of *A. fraseriana* and *Banksia grandis* (Table 2). Marri basal area had only been reduced at Bell, where a large mature tree had fallen over after having been burnt through at the base of the stem. Cut stumps from earlier harvesting operations were not recorded in any of the shelterwood stands, but several mature jarrah trees at Godfrey had fallen over as a result of fire damage.

Gap release treatments had relatively high basal areas (18-32 m²/ha) comprised mostly of jarrah. At Godfrey (FC31) this was due to retention of a cohort of trees 35-50 cm dbh and occasional large habitat trees. At Stockyard (FC34) the high basal area resulted from dense sapling regeneration released by silvicultural treatment and from retention of trees of pole-size (15-45 dbh) and occasional mature trees. Sapling regeneration also contributed to the basal area at Bell (FC37) but to a lesser extent than at Stockyard. Basal areas of *A. fraseriana* in gap release treatments were low (<1 m²/ha) and there had been only limited pushing down of standing trees to achieve this result (Table 2).

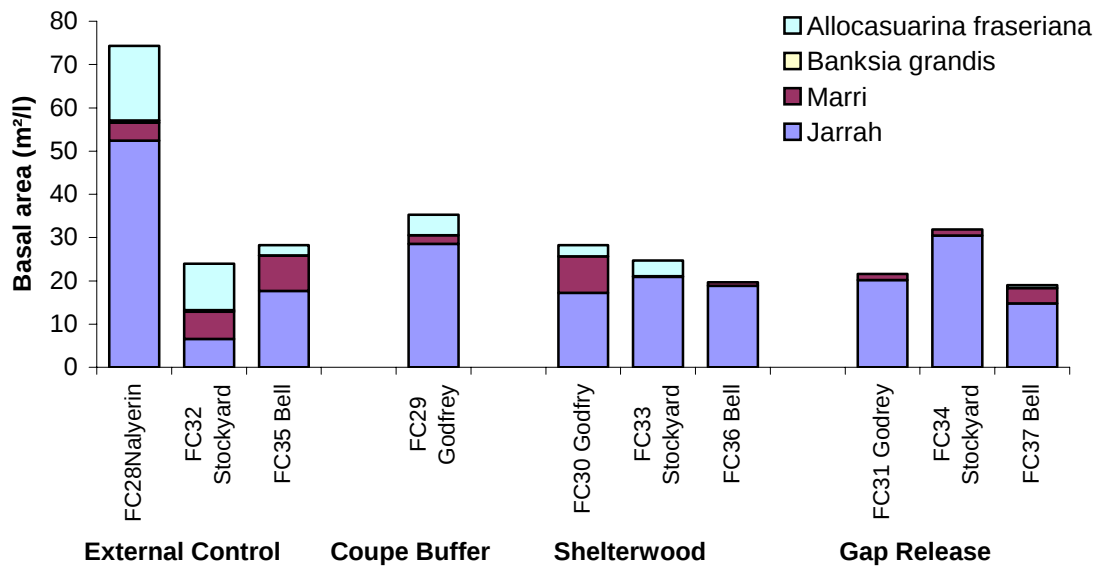


Figure 1. Basal area of jarrah, marri and intermediate trees.

Marri was present as a secondary species although its contribution to basal area varied considerably between stands. Marri made the largest relative contribution to basal area in the external control at Stockyard, although this may have been because the basal area of jarrah had been reduced by earlier timber harvesting. Marri contribution to basal area ranged from 7-31 % in the other external control grids that had no evidence of previous harvesting. Relative proportions of jarrah and marri were generally consistent between basal area, stems per ha, and the regeneration cohort of saplings and advance growth (Table 1). The current silvicultural guideline requires that on predominantly jarrah sites the species mix of the eucalypt regeneration should contain at least 20 % jarrah. This requirement was achieved in silviculturally treated stands at Godfrey and Stockyard, but could not be assessed at Bell because of the effects of recent fire.

Table 1. Analysis of eucalypt species composition determined according to basal area and stem density of live standing trees >2 m tall, and the species mix determined in regeneration surveys of FORESTCHECK grids in Wellington East. Regeneration stocking could not be assessed at Bell forest block because of the grids were burnt by an unplanned fire in December 2004.

Treatment Grid	Basal area (m ² /ha)			Stems/ha			Regeneration Species Composition	
	jarrah	marri	total	jarrah	marri	total	jarrah	marri
External Control								
FC28	52.45	4.11	56.56	238	63	301	-	-
	93%	7%		79%	21%		70%	30%
FC32	6.56	6.34	12.90	300	300	600	-	-
	51%	49%		80%	20%		37%	63%
FC35	17.70	8.10	25.80	163	50	213	-	-
	69%	31%		77%	23%		n/d	n/d
Coupe Buffer								
FC29	28.51	2.02	30.53	600	163	763	-	-
	93%	7%		79%	21%		33%	66%
Shelterwood								
FC30	17.21	8.45	25.66	1513	338	1851	-	-
	67%	33%		82%	18%		65%	35%
FC33	20.91	0	20.91	313	0	100	-	-
	100%	0%		100%	0%		73%	27%
FC36	18.86	0.80	19.66	350	150	500	-	-
	96%	4%		70%	30%		n/d	n/d
Gap Release								
FC31	20.21	1.35	21.56	125	63	188	-	-
	97%	3%		66%	33%		53%	47%
FC34	30.48	1.41	31.89	1650	350	200	-	-
	97%	3%		83%	17%		97%	2%
FC37	14.80	3.54	18.34	513	213	726	-	-
	81%	19%		70%	30%		n/d	n/d

Table 2. Basal area of cut stumps and trees pushed down in FORESTCHECK grids in Wellington East. Figures in italics show how the percentage reduction in basal area is distributed between 2 transects on left and right hand sides of the sampling grid. Other trees and shrubs were predominantly *A. fraseriana* and *B. grandis*.

Treatment Grid	Basal area reduction from most recent harvesting (m ² /ha)			Basal area reduction from earlier harvesting or disturbance (m ² /ha)		
	jarrah	marri	other	jarrah	marri	other
External Control						
FC28	-	-	-	-	-	2.45 <i>100/0</i>
FC32	-	-	-	19.8 <i>40/60</i>	-	-
FC35	-	-	-	-	-	0.10 <i>100/0</i>
Coupe Buffer						
FC29	-	-	-	35.54 <i>78/22</i>	-	-
Shelterwood						
FC30	8.98 <i>100/0</i>	-	-	16.69 <i>0/100</i>	-	-
FC33	-	-	10.37	-	-	-
FC36	10.73 <i>58/42</i>	28.37 <i>0/100</i>	-	-	-	-
Gap Release						
FC31	10.40 <i>71/29</i>	-	2.15 <i>0/100</i>	-	-	-
FC34	21.62 <i>100/0</i>	1.49 <i>0/100</i>	-	-	-	-
FC37	7.36 <i>77/33</i>	-	-	4.81 <i>100/0</i>	-	-

Regeneration stocking

Grids in uncut forest contained relatively few saplings but moderate stocking levels of ground coppice and advance growth (Table 3). The general absence of saplings can be attributed to the lack of recent disturbance and associated temporary reduction in competition from the overstorey canopy.

Satisfactory stocking in stands cut to shelterwood is defined as having 65 % of sample points with 500 or more stems/ha of saplings or stool coppice from stumps <30 cm diameter, or 1000 or more stems/ha of saplings, stool coppice and jarrah ground coppice or marri advance growth, and can also include lignotuberous seedlings at 5000 or more stems/ha. The Godfrey shelterwood (FC30) was marginally stocked (52 % of sample points) with saplings, advance growth and some seedlings established as a result of silvicultural treatment (Table 3). The level of retained basal area on the Godfrey shelterwood was higher than recommended in the current specification and resulted in 44 % of sample points being influenced by overwood. The Stockyard shelterwood (FC33) was poorly stocked with eucalypt regeneration, probably reflecting a low stocking of

lignotuberous advance growth at the time of harvest. Few seedlings had established as a result of silvicultural treatment.

The Godfrey gap release (FC31) was 40 % stocked, mostly with ground coppice, and did not meet the regeneration standard of 65 %. Only 2 years have elapsed since post-harvest burning and so the proportion stocked with saplings can be expected to increase as ground coppice develops. Much of the current ground coppice showed evidence of dynamic growth. The Stockyard gap release (FC34) was 78 % stocked and contained a stratum of dynamic well-formed saplings (Tables 1 and 3).

Table 3. Regeneration stocking and species composition for FORESTCHECK grids in Wellington East assessed during March 2005. Regeneration stocking was not assessed on 3 grids at Bell forest block burnt by unplanned fire in December 2004. Values for per cent stocking are based on 50 sample points per grid. Retained over-wood was not assessed in uncut stands or coupe buffers.

Treatment Grid	Height range of eucalypt regeneration (m)	Percent affected by overwood	Percent stocked with saplings	Percent stocked with saplings & ground coppice	Per cent stocked including seedlings	Per cent not stocked to standard
External Control						
FC28	Uncut	N/a	2	46	N/a	6
FC32	Uncut	N/a	16	36	N/a	6
FC35	Uncut	N/a	N/d	N/d	N/a	N/d
Coupe Buffer						
FC29	Uncut	N/a	6	38	N/a	2
Shelterwood						
FC30	3-4	44	22	22	8	4
FC33	2-4	28	-	14	2	56
FC36	N/d	N/a	N/d	N/d	N/a	N/d
Gap Release						
FC31	1-2	30	4	36	N/a	30
FC34	4-5	14	34	44	N/a	14
FC37	N/d	N/a	N/d	N/d	N/a	N/d

Regeneration originating from stool coppice on stems >30 cm diameter and from stump coppice is not considered to contribute to the effective stocking at a point. This is because it is prone to wind-throw and is readily damaged by fire if the parent stump burns. However on most sites there are a considerable number of additional stems originating from stump coppice that contribute to the initial re-establishment of crown cover and basal area.

The origin of all jarrah and marri stems >2 m tall measured in the 4 x 200 m transect on each grid is shown in Figure 2. Saplings on the external control and coupe buffer grids were mostly of lignotuberous origin. This reflects the fact that the Bell and Nalyerin sites had no evidence of previous harvesting, and that the harvesting in the Stockyard control and Godfrey coupe buffer

had removed only large trees thus creating a relatively small number of cut stumps. Trees of lignotuberosus origin and stool coppice from small stumps contributed to adequate stocking at the Godfrey shelterwood, Stockyard gap release and Bell gap release. Stump coppice made only a small contribution to stocking at any of the grids.

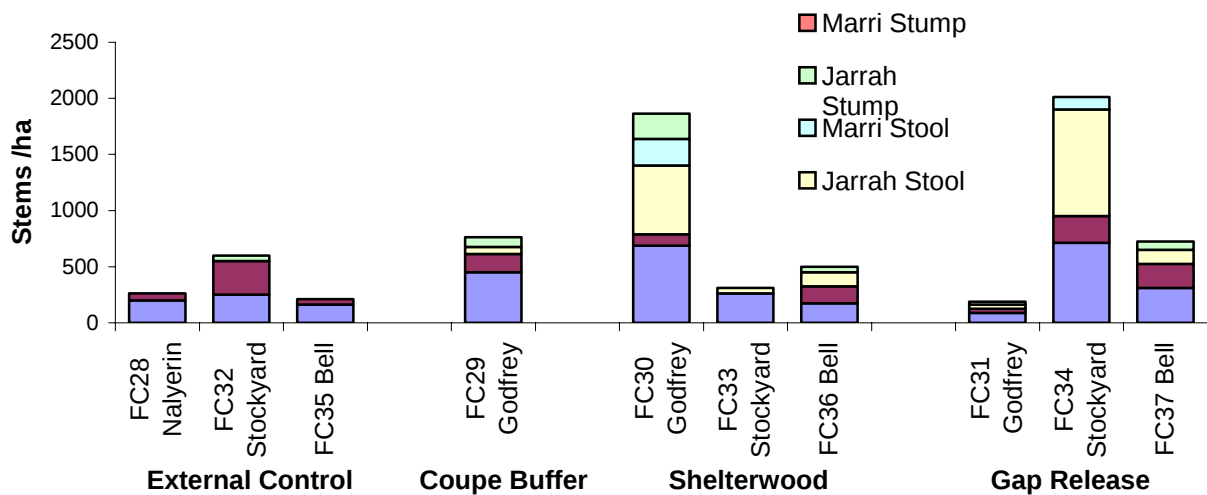


Figure 2. Origin of all jarrah and marri stems >2 m tall measured in the 4 x 200 m transect on ten grids in Wellington East.

Canopy cover

Overstorey canopy cover in external control and coupe buffer grids ranged from 47-62 % (Figure 3). Shelterwood grids had canopy cover ranging from 32 % in the most recently treated stand at Godfrey through to 50 % at Bell, even though Bell had experienced the largest reduction in basal area as a result of silvicultural treatment and had the lowest retained basal area (Table 2). Gap release grids had canopy cover ranging from 24 to 40 %, with a trend for cover to increase with increasing time since silvicultural treatment. This is expected because of the recruitment of saplings, and increased foliage density on retained trees.

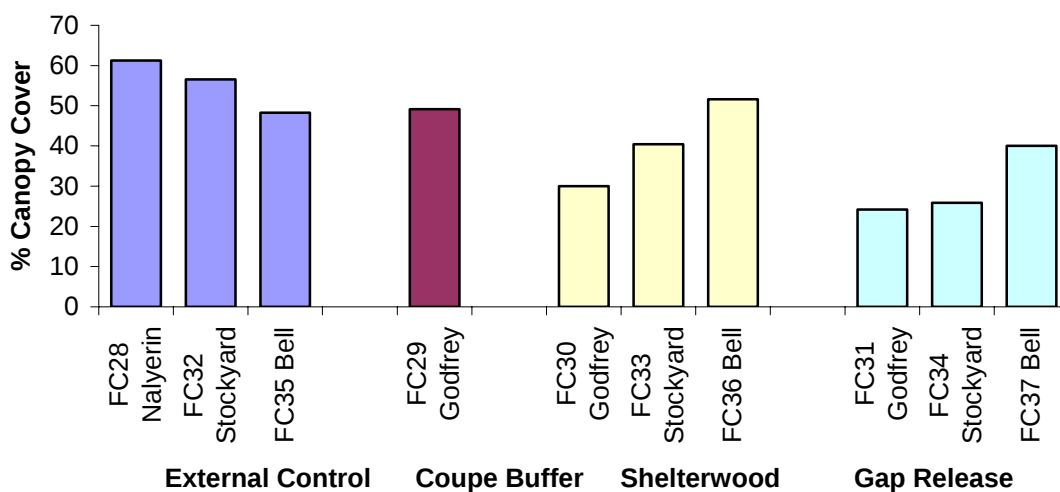


Figure 3. Canopy cover estimated using point intercept sampling.

Assessment of burnt grids

A standard procedure has been developed for assessment of grids burnt by prescribed fires wildfires. The 15 main pegs that define the layout of each grid are used as sample points, and at each peg an assessment is made to determine:

1. the proportion of area within 5m radius of the peg that has burnt,
2. the height of crown scorch to the overstorey canopy,
3. average height of the scrub layer
4. depth of any residual litter layer.

Post-fire assessment was undertaken at the 3 Bell forest block grids, and at the Occidental forest block external control grid (FC27) that was burnt at very high intensity during the large Perth Hills bushfire in mid-January 2005 (Table 4).

Table 4. Number of sample points (from a total of 15) allocated to various categories indicative of fire intensity.

Grid	Category				Total
	Defoliated	Full crown scorch	Partial crown scorch	Unburnt	
Occidental					
FC27	15	-	-	-	15
Bell					
FC35	-	6	9	-	15
FC36	-	3	8	4	15
FC37	-	3	12	-	15

Discussion

Eastern jarrah forest (CALM 2004) is a broad description for forest that occurs generally east of the 900 mm rainfall isohyet. Eastern forest is characterized for the most part by more widely spaced trees and a low herbaceous understorey. Lignotuberous advance growth is patchier and less vigorous than in higher rainfall forest closer to the Darling Scarp, and thickets of *A. fraseriana* and *B. grandis* are less extensive. All of the sampling grids for 2004-05 were located in vegetation complex Dwellingup 4 (Mattiske and Havel 1998) which is regarded as transitional between western and eastern forest. Stand growth and individual tree growth are typically slower in these drier eastern forests than in western forests (Abbott and Loneragan 1983).

External control and coupe buffer grids included in this year's program had either not previously been harvested for timber, or had not been harvested for at least 50 –70 years. The Nalyerin external control grid was notable for its high stand basal area (56 m²/ha), comprised predominantly of jarrah. Considering the current basal area of standing jarrah and of cut stumps we conclude that the stand density of the Godfrey coupe buffer grid would have been similarly large. Abbott and Loneragan reported stand basal area increments of 0.6 m²/ha/decade for eastern forest, and extrapolating from this figure the expected recovery of basal area following earlier partial cutting would be 3-4 m²/ha. Superior soil fertility and long absence of major disturbance to the overstorey are factors that could account for the large basal area of these stands. Mature trees had dbh up to 1 m, with most codominant trees having dbh of 70-90 cm. There were fewer very large trees in the eastern forest than recorded in the 2002 set of Wellington District sites that adjoined the 900 mm annual rainfall isohyet.

Shelterwood treatments had been applied conservatively, with retained basal areas on the 3 grids substantially exceeding the minimum level of 6 m²/ha recommended (CALM 2004). Jarrah poles (15-45 cm dbh) and small mature trees (45-70 cm dbh) made up the majority of retained basal area in shelterwoods. None of the shelterwood treatments had involved extensive push down of *A. fraseriana*, and retained basal areas of *A. fraseriana* were small. The regeneration result in the Stockyard shelterwood was poor, with little seedling establishment and few saplings exhibiting dynamic growth.

Gap cutting at Stockyard had released a dense cohort of saplings with a high proportion of these originating from lignotubers, and this treatment could be judged as very successful. The Godfrey gap was not as well stocked but still has a substantial overwood of established trees.

The Bell forest block sites will be assessed for regeneration stocking in autumn 2006 by which time lignotuberos advance growth burnt back by the December 2004 fire will have had an opportunity to re-sprout.

Recommended change to sampling procedure

The only change to field sampling procedure that should be considered at this stage relates to how established trees of pole size (15-45 cm dbh) are recorded during regeneration stocking assessment in areas cut to gap release. Currently the sampling protocol specifies that saplings may be included in regeneration assessment, and that sampling points with a tree of >50 cm dbh within 4 m should be recorded as stocked. However, pole sized trees are not directly accounted for in this protocol even though they may often be the preferred form of regeneration and should be retained where possible. From a silvicultural perspective the logical approach would be to consider a point as stocked if it included a pole sized tree of acceptable form within 4 m radius of the sample point. Cull trees having poor form or high levels of stem defect would not be counted as acceptable stocking.

Data management

Regeneration stocking data were summarized and entered into the FORESTCHECK database. Data were analysed to determine the following key information:

- whether the grid meets current stocking standards,
- species composition of overstorey trees and the regeneration cohort,
- proportion of the grid affected by retained overwood (gap and shelterwood grids only),
- average density of saplings and ground coppice at points that meet the stocking standard.

References

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FOLIAR AND SOIL NUTRIENTS

Lachlan McCaw, John Neal, Bob Smith

Introduction

Concentrations of nitrogen (N), phosphorus (P) and potassium (K) in the foliage of advance growth, saplings and overstorey trees, and in surface soils are measured at each FORESTCHECK monitoring grid to provide information about the nutritional status of the forest ecosystem. Data will be used to investigate correlations between macronutrient concentrations and measures of plant and animal abundance, and ecosystem health.

Monitoring

Samples of foliage and soil were collected from 7 of the Wellington East grids in mid March 2005. The 3 Bell forest block grids that were burnt during an unplanned fire in December 2004 were not sampled but will be in autumn 2006. The methodology for collection and analysis of samples is described in the Operations Plan. Samples were analysed at the WA Chemistry Centre.

Foliar nutrients

Foliage from mature marri trees tended to have slightly higher concentrations of N, P and K than measured in advance growth or saplings. Growth stage appeared to have little or no effect on concentrations of N, P or K in jarrah foliage (Table 1). Mature marri foliage had higher concentrations of N, P and K than mature jarrah foliage, with this difference being almost threefold in the case of K. These patterns of foliar nutrient concentration are generally consistent with those observed in the previous 3 years of sampling.

Table 1. Concentrations of N, P and K in the foliage of jarrah and marri advance growth, saplings and mature trees for 7 grids in Wellington East. Data are shown as minimum-**mean**-maximum.

Species	Foliage source	% Nitrogen (total)	% Phosphorus	% Potassium
Jarrah	Advance growth	0.63- 0.69 -0.76	0.020- 0.023 -0.027	0.24- 0.38 -0.56
	Sapling	0.59- 0.74 -0.95	0.020- 0.026 -0.034	0.30- 0.34 -0.37
	Mature	0.69- 0.77 -0.82	0.023- 0.027 -0.032	0.31- 0.37 -0.41
Marri	Advance growth	0.71- 0.83 -0.96	0.024- 0.031 -0.035	0.56- 0.78 -0.88
	Sapling	0.73- 0.82 -0.94	0.025- 0.031 -0.037	0.55- 0.81 -0.97
	Mature	0.73- 0.90 -0.99	0.030- 0.037 -0.043	0.74- 0.90 -1.14

Soil nutrients

Soil N concentrations for the Godfrey and Nalyerin grids were consistently twice as high as those measured in the 3 Stockyard grids (Table 2). Extractable P concentrations were also consistently greater for the Godfrey and Nalyerin grids, although total P concentrations were more comparable, except for the external control at Stockyard forest block (FC32).

Extractable K concentrations varied by a factor of about 2.5 with the lowest levels measured in the Stockyard grids. The Stockyard external control also had a notably low concentration of total K.

Table 2. Mean concentrations of N, extractable and total P, and extractable and total K determined from 5 surface soil samples. The overall mean (s.e.m.) for 7 grids in Wellington East is indicated.

Grid	N Per cent	P_extract ppm	P_total ppm	K_extract ppm	K_total ppm
FC28	0.28	3.25	166	114.7	123
FC29	0.18	2.75	123	65.6	102
FC30	0.23	3.80	133	128.4	94
FC31	0.25	3.50	146	132.0	146
FC32	0.11	<2	55	45.0	42
FC33	0.13	<2	112	49.4	148
FC34	0.14	<2	123	70.2	130
MEAN	0.18	2.76	122.3	86.47	112.14
(s.e.m)	(0.02)	(0.29)	(13.35)	(14.16)	(14.00)

Discussion

There was a clear distinction between the low nutrient concentrations in the soils at Stockyard forest block and the higher levels measured at Nalyerin and Godfrey. The Stockyard Control (FC32) stood out as having the lowest concentrations of all macronutrients. The poor nutrient status of this site was reflected in a low basal area of jarrah and marri. Soil and foliar nutrients will be sampled in the Bell forest block grids in autumn 2006 and this will allow a more complete comparison of nutritional status of eastern jarrah forest sites.

Comparing the overall mean concentrations of soil nutrients measured this year in Wellington East and previous years in Donnelly, Perth Hills and Wellington (Table 3), shows that the eastern jarrah forest sites in Wellington East have:

- higher mean N concentrations,
- higher mean concentrations of total P and comparable concentrations of available P, and
- higher mean concentrations of extractable K and similar concentrations of total K than the other sites.

Foliar nutrient concentrations measured in Wellington East were generally comparable with those measured in Donnelly and Wellington in previous years.

Table 3. Mean (s.e.m) soil nutrient concentrations for FORESTCHECK grids in Donnelly, Wellington, Perth Hills and Wellington East locations.

District	N Per cent	P_extract ppm	P_total ppm	K_extract ppm	K_total ppm
Donnelly	0.12 (0.01)	2.0 (0.4)	65.4 (13.4)	43.8 (5.6)	94.8 (13.5)
Wellington	0.14 (0.01)	2.3 (0.6)	85.2 (16.3)	54.7 (5.6)	119.0 (11.0)
Perth Hills	0.13 (0.03)	1.4 (0.4)	32.3 (16.7)	53.6 (17.1)	135.7 (49.4)
Wellington East	0.18 (0.02)	2.8 (0.3)	122.3 (13.4)	86.7 (14.1)	112.1 (14.0)

Recommended changes to operating procedure

The WA Chemistry Centre analysed soil and foliar samples this year at similar cost to previous years.

SOIL DISTURBANCE

Kim Whitford

Introduction

This report covers the 2004-05 soil disturbance assessments of the Wellington East FORESTCHECK sites. Only a subset of the sites are sampled as soil disturbance cannot be reliably assessed on all sites. Bulk density is measured on only one harvested site each year. This year the Stockyard shelterwood site (FC33) was measured. The soil bulk density on this site was compared with the bulk density on an adjacent undisturbed site to determine the soil compaction caused by timber harvesting. As snig tracks are the source and location of most soil disturbance produced by logging, mapping of snig track layout and snig track order was attempted on all of the harvested sites.

The objectives of this work were to:

- Record the extent of soil disturbance on FORESTCHECK monitoring sites where machine disturbance (snig tracks) could be readily identified.
- Monitor the intensity of changes to soil physical properties induced by logging, on selected sites.
- Provide base data that could be used to monitor any change in these soil physical properties over time.
- Contribute data that can be used to relate the intensity of disturbance to the operational categories, where intensity of disturbance is measured as the fine earth bulk density of the soil.

Monitoring

Soil bulk density samples were collected in June 2005. Data from the Stockyard shelterwood (FC33) were compared with data from an adjacent unlogged site. These sites were selected for measurement of bulk density because: (a) the surface soils were relatively free of large boulders or sheet caprock, (b) the 2 sites were the same Matiske-Havel vegetation complex and consequently could be expected to have similar soils, and c) the snig tracks could be successfully mapped on the Stockyard shelterwood site, thus providing information on the operational categories of the bulk density sample points. The sampling of bulk density was stratified on the basis of operational categories. Operational categories are classifications of the type of harvesting activities that have occurred at the sample point, e.g. harvested area, unharvested area, log landing, old log landing, old snig track, snig track order.

On the Stockyard shelterwood site, and the remaining 3 sites where bulk density was not measured, mapping of snig tracks provided an assessment of both the amount and the location of the disturbance. Incorrect or low quality mapping gives a poor indication of soil disturbance. Satisfactory mapping of the snig track layout occurred on the Stockyard shelterwood and gap release (FC34), and the Godfrey shelterwood (FC30) and gap release sites (FC31) (see Table 1). The snig tracks could not be mapped on the harvested Bell forest block sites (FC36 and FC37). This is attributed to the amount of surface rock at these sites which prevented depression of the soil under the wheel load of the machinery, the burning of the sites which removed some of the plant evidence of vehicle tracks, and possibly the relatively light harvest that occurred at these sites. The relative quality of the snig track mapping was high on the Stockyard shelterwood site, where primary, secondary, and tertiary snig tracks could be readily identified. In addition some old snig tracks from the previous 2 harvesting events could be identified. The quality of the snig track mapping on the remaining sites (FC30, FC31, and FC34) was reasonable, with primary and

secondary snig tracks identified. The tertiary snig tracks were not well mapped on any of these 3 sites. Surface rock and dispersed harvesting prevented detailed mapping of the tertiary snig tracks on the Stockyard gap release site, though some old snig tracks were identified and mapped on this site. The relatively light harvest on the Godfrey shelterwood resulted in limited mapping of the tertiary snig tracks. Disturbance from what appears to be post harvest pushdown, or possibly the machine harvesting on the Godfrey gap release site obscured some of the tertiary snig tracks on that site and limited the mapping of these snig tracks.

Preliminary results

The sites and treatments assessed and measured are listed in Table 1. Table 2 gives the means and standard errors for total bulk density (TBD) and fine earth bulk density (FEBD), and gravel content for the various operational categories on the Stockyard shelterwood and an adjacent undisturbed unlogged area with similar soil.

Table 1. The type of assessment and the number of samples collected at each site.

Site		Site label	Snig track map	Operational category sample points	Bulk density sample points	Quality of snig track mapping
Godfrey	Coupe Buffer	FC29	No			Not mapped
Godfrey	Shelterwood	FC30	Yes			Satisfactory
Godfrey	Gap Release	FC31	Yes			Satisfactory
Stockyard	External Control	FC32	No			Not mapped
Stockyard	Shelterwood	FC33	Yes	175	175	High
Stockyard	Gap Release	FC34	Yes			Satisfactory
Bell	External Control	FC35	No			No snig tracks
Bell	Shelterwood	FC36	No			Not mapped
Bell	Gap Release	FC37	No			Not mapped
TOTAL				175	175	

Table 3 shows the area of the snig tracks and landings for 4 sites from this years monitoring and 6 sites from previous years monitoring. The snig track maps (Figs. 1, 2, 3 and 4) show the location and extent of snig tracks and landings on the Stockyard and Godfrey shelterwood and gap release sites.

Table 4 provides a summary of the comparisons of soil bulk density that can be made between the control or undisturbed sites and the recently harvested sites over the 4 years of FORESTCHECK monitoring. Six such comparisons can be made. For each of the recently harvested sites 2 values of the mean bulk density are given; one mean for samples collected on the systematically sampled grid points, and another mean for the samples collected from the “harvested area” operational category. The change in bulk density as a result of the harvesting is best estimated by comparison of the mean bulk densities from the unharvested control site and the systematic grid point samples from the recently harvested sites.

Table 2. Number of samples (n), bulk density (g/cm³ and SE) and gravel content (% and SE) for operational categories at the Stockyard shelterwood FORESTCHECK site and an adjacent area of unlogged forest. Operational categories: harvested area (HA), unharvested area (UA), major snig track into landing (ST0), primary snig track (ST1), secondary snig tracks (ST2), tertiary snig track (ST3), old snig track from previous logging (OST), old primary snig track from previous logging (OST1), old tertiary snig track from previous logging (OST3), log landing (LL), old log landing (OLL), access track (Track).

FORESTCHECK site	Operational category	n	Total bulk density (g/cm ³)		Fine earth bulk density (g/cm ³)		Gravel content (%)	
				SE		SE		SE
Stockyard shelterwood FC33	HA	76	1.395	0.029	0.672	0.015	64.8	1.0
	LL	22	1.604	0.034	0.955	0.033	60.6	0.9
	OST	14	1.608	0.081	0.832	0.054	66.2	1.9
	ST1	20	1.709	0.052	0.929	0.039	64.8	0.8
	ST2	21	1.555	0.066	0.804	0.038	64.4	1.7
	ST3	22	1.526	0.060	0.775	0.036	63.9	2.3
Stockyard adjacent unlogged	UA	40	1.396	0.032	0.664	0.018	65.6	1.6

Discussion

Bulk density was measured on the Stockyard shelterwood (logged 3 times, most recently in 1998). These bulk density measurements were compared with measurements from an adjacent area that had never been harvested. The soils on the 2 sites were very similar. The mean gravel content of the Stockyard shelterwood site was 64% and the mean gravel content of the adjacent unlogged site was 66%.

The bulk density values on the Stockyard shelterwood site followed a logical and consistent trend with the compaction increasing as the amount of traffic increased across the operational categories. The bulk density of the log landing was 44% higher than the bulk density of the undisturbed site. The bulk density of the primary snig tracks was 40% higher, the bulk density of the secondary snig tracks was 21% higher, and the bulk density of the tertiary snig tracks was 17% higher than the bulk density of the undisturbed site.

The bulk density of the general harvested area that was not covered by snig track was 0.67 ± 0.015 g/cm³ while the bulk density of the undisturbed soil on the adjacent unlogged area was 0.66 ± 0.018 g/cm³. The bulk density of the general harvested area on the Stockyard shelterwood site was not significantly different from the bulk density of the adjacent undisturbed unlogged area.

The mean bulk density for the Stockyard shelterwood site determined from a systematic grid sampling across the site was 0.71 ± 0.018 g/cm³. This is only slightly higher than the mean bulk density of the adjacent unlogged site, which was 0.66 ± 0.018 g/cm³. This 8% increase in the fine earth bulk density of the surface soil (0-10cm) (Table 4) is relatively small but still a significant increase in bulk density, attributable to the 3 harvesting events that occurred on the Stockyard shelterwood site.

Table 3. The estimated area of the fallers block surrounding each FORESTCHECK site, the area of snig tracks and landings identified on each fallers block, and the areal proportion of the fallers block that has been disturbed by snig tracks and landings. Snig track classes are: primary (ST1), secondary (ST2), tertiary (ST3), and old snig track from a previous logging (OST). Snig track area calculations are based on measurements of snig track lengths and assumed widths of 4.67 m for ST1, 4.46 m for ST2, and 4.13 m for ST3.

Site	Site code	ST1 (m ²)	ST2 (m ²)	ST3 (m ²)	OST (m ²)	Total snig track area (m ²)	Landing Area (m ²)	Fallers block Area (m ²)	Landing area (% of fallers block)	Snig track area (% of fallers block)	% of fallers block disturbed
Edward gap main landing and fallers block	FC11	1,365	1,346	5,326	0	8,037	1,970	55,220	3.6	14.6	18.1
Edwards gap second landing	FC11	n/a	n/a	n/a	n/a	n/a	2,240	n/a			
Edwards gap third landing	FC11	n/a	n/a	n/a	n/a	n/a	600	n/a			
Ross gap	FC12	1,804	2,331	5,256	0	9,391	4,350	105,300	4.1	8.9	13.0
Ross shelterwood eastern fallers block	FC13	1,112	1,560	8,262	0	10,934	2,248	77,130	2.9	14.2	17.1
Surface shelterwood	FC15	2,351	1,761	6,908	0	11,020	1,590	65,760	2.4	16.8	19.2
Chalk shelterwood western fallers block	FC18	322	825	5,995	2,159	9,301	2,120	43,820	4.8	21.2	26.1
Chalk shelterwood eastern fallers block	FC18	444	1,511	3,596	2,739	8,290	680	46,700	1.5	17.8	19.2
Chalk combined, both fallers blocks	FC18	766	2,336	9,591	4,899	17,592	2,805	90,520	3.1	19.4	22.5
Chalk shelterwood west, excluding OST	FC18	322	825	5,995	excluded	7,142	2,120	43,820	4.8	16.3	21.1
Chalk shelterwood east, excluding OST	FC18	444	1,511	3,596	excluded	5,551	680	46,700	1.5	11.9	13.3
Chalk combined, excluding OST	FC18	766	2,336	9,591	excluded	12,693	2,805	90,520	3.1	14.0	17.1
Lesley gap	FC25					9,666	418	120,179	0.4	8.4	
Godfrey Shelterwood Landing 1 (East)	FC30	1,453	1,141	1,817	963	5,374	2,677	74,043	3.6	7.3	10.9
Godfrey Shelterwood Landing 2 (West)	FC30						2,970				
Godfrey Gap Release Landing 1 (East)	FC31	790	2,679	1,590	1,664	6,722	2,366	101,400	2.3	6.6	9.0
Godfrey Gap Release Landing 2 (West)	FC31	220	2,536	1,420	173	4,349	2,044	77,406	2.6	5.6	8.3
Stockyard Shelterwood	FC33	2,491	3,664	10,434	9,233	25,822	1,802	200,990	0.9	12.8	13.7
Stockyard Gap Release Landing 1 (South)	FC34	1,467	1,110	1,197	1,444	5,219	1,358	80,959	1.7	6.4	8.1
Stockyard Gap Release Landing 2 (North)	FC34	3,094	1,810	801		5,704	2,568	181,224	1.4	3.1	4.6
Mean	All	-	-	-	-	-	-	-	2.6	12.1	15.1

Table 4. A summary of the mean change in surface soil (0-10cm) bulk density attributed to timber harvesting operations across all FORESTCHECK sites.

Site	Site Code	Operational Category	Number of times harvested	Year of last harvest	Years since harvest	Mattiske-Havel vegetation complex	Fine earth bulk density (g cm ⁻³)	% Change (grid vs. not harvested)	n ¹	Gravel content (%)	Assessment year
Kingston gap	FC2	Grid points	3	1995/96	7	Corbalup 2	0.82 ± 0.03		77	33	2001
Kingston gap	FC2	Harvested	3	1995/96	7	Corbalup 2	0.80 ± 0.02		68	32	2001
Kingston TEAS	FC4	Not harvested	2	1970s	22-32	Corbalup 2	0.92 ± 0.02	-11	39	10	2001
Thornton gap	FC6	Grid points	2	1991	11	Corbalup 1	1.00 ± 0.02		77	17	2001
Thornton gap	FC6	Harvested	2	1991	11	Corbalup 1	0.98 ± 0.02		75	17	2001
Thornton TEAS	FC7	Not harvested	1	1940s	52-62	Corbalup 1	0.76 ± 0.04	32	40	47	2001
Carter gap	FC8	Harvested	2	1999	3	Collis 1	0.80 ± 0.01		137	35	2001
Carter TEAS	FC9	Not harvested	1	1940s	52-62	Collis 1	0.78 ± 0.02	3	40	55	2001
Chalk shelterwood	FC18	Grid points	3	1992	11	Dwellingup 1	0.81 ± 0.02		81	65	2002
Chalk shelterwood	FC18	Harvested	3	1992	11	Dwellingup 1	0.80 ± 0.02		67	65	2002
Tumlo control	FC19	Not harvested	0	unlogged	n/a	Dwellingup 1	0.60 ± 0.02	35	40	70	2002
Cameron shelterwood	FC23	Grid points	3	1989	15	Dwellingup 1	0.92 ± 0.01		74	53	2003
Cameron shelterwood	FC23	Harvested	3	1989	15	Dwellingup 1	0.92 ± 0.01		73	52	2003
Kennedy control	FC24	Not harvested	2	1930-34	74-70	Dwellingup 1	0.83 ± 0.02	11	40	67	2003
Stockyard shelterwood	FC33	Grid points	3	1998	7	Dwellingup 4	0.71 ± 0.02		75	65	2005
Stockyard shelterwood	FC33	Harvested	3	1998	7	Dwellingup 4	0.67 ± 0.02		76	65	2005
Stockyard unlogged		Not harvested	0	unlogged	n/a	Dwellingup 4	0.66 ± 0.02	7	40	66	2005
Mean								13			

¹ Number of bulk density samples taken.

Table 5. Comparison of soil bulk densities (g/cm³) for the surface soil (0-10cm) following tractor logging as reported in Australia and overseas (Rab, 1992).

Source	Total bulk density (g/cm ³)				Percentage change from undisturbed		
	Undisturbed areas	Harvested areas	Snig tracks	Log landings	Harvested areas	Snig tracks	Log landings
Dickerson (1976)	1.29	1.42	1.55		10.1	18.3	
Froehlich (1979)	0.97		1.14				
Jakobsen (1983)	0.9		1.07				
Gent <i>et al.</i> (1984)	1.14	1.36	1.52		19.3	27.9	
Incerti <i>et al.</i> (1987)	0.96	0.99	1.22	1.33	3.1	26.3	30.3
Rab <i>et al.</i> (1992)	0.94	1.02	1.12	1.19	8.5	17.6	22.3
Anderson <i>et al.</i> (1992)	0.71	0.86	1.1	1.22	21.1	45.3	46.4

Table 4 summarizes the mean change in surface soil bulk density attributed to timber harvesting operations across all of the FORESTCHECK sites where bulk density has been measured. The change in bulk density as a result of timber harvesting is estimated by comparing the mean bulk densities from the grid point samples for the recently harvested sites, with the unharvested control sites. The comparisons in Table 4 are made between sites with the same Mattiske-Havel vegetation complex. Generally soils are similar within a Mattiske-Havel vegetation complex, although this is not always the case with one such exception occurring with the Kingston gap release and the immediately adjacent Kingston coupe buffer (TEAS) sites. The soil on these 2 sites was very different – the Kingston coupe buffer being a sand, and the Kingston gap release a gravely loam. Comparison of bulk density between these 2 sites is not valid because the soils are clearly different.

Excluding the Kingston gap release site, the mean increase in soil bulk density attributable to timber harvesting across these 5 sites was 18%. This is a substantial increase in the mean bulk density of the surface soils, and can be identified as a likely impact of timber harvesting. Similar data from a greater number of sites and further comparisons with unlogged sites is needed to clarify the validity of this observed increase in bulk density.

Table 5 lists actual and per cent increase in total bulk density observed at other harvested forest sites in Australia and overseas. The values indicate that the increase in soil bulk density observed in the jarrah forest (~18%) is within the upper range observed in other forest types (3% to 20%).

Of the 5 comparisons shown in Table 4, only 2, the Chalk shelterwood / Tumlo external control, and the Stockyard shelterwood/adjacent unlogged area, enable a comparison of a unlogged site with a harvested site. These 2 comparisons show surprisingly different responses. Both harvested sites have been logged 3 times, have similar soils with similar gravel contents, yet the Chalk shelterwood site shows a much greater increase in soil bulk

density (35%) attributable to the timber harvesting, than the Stockyard shelterwood site (8%). Factors that influence the impact of timber harvesting on soil bulk density include the soil moisture at the time of harvesting, the intensity of the timber harvest, and soil particle size distribution (Rab *et al.* 2005). Differences in these 3 factors between the 2 harvesting operations, particularly differences in soil moisture, most likely explain the differing impact of these operations on the soil bulk density.

Of the 4 FORESTCHECK monitoring sites that were examined this year, the Stockyard gap release (FC34) and the Godfrey gap release (FC31) have been harvested twice, and the Stockyard shelterwood (FC33) and part of the Godfrey shelterwood (FC30) has been harvested 3 times. The most recent harvest on the Godfrey gap release was a machine harvest in 2000. This type of harvesting operation, which has only recently been introduced into the jarrah forest and is now the predominant mode of timber harvesting, greatly influences the timber extraction pattern and the type, and intensity of soil disturbance that occurs on the harvested area. Machine harvesting generally decreases the amount of tertiary snig tracks on a site reducing the amount of soil compaction attributable to tertiary snig tracks. However, machine harvesting also greatly increase the amount of soil mixing that occurs across the harvested area.

As with the 2003-04 FORESTCHECK sites, the boundaries of the faller's blocks about the 2004-05 FORESTCHECK sites in the northern jarrah forest were not as well delineated as the 2001-02 FORESTCHECK sites in the southern jarrah forest. Consequently the faller's block areas presented in Table 3 are estimates. These areas were estimated from the extent and distribution of the snig tracks and harvesting on these sites. This reduces the accuracy of the last 2 columns in Table 3, i.e. the estimates of the area of the landing as a proportion of the faller's block area, and the estimates of the proportion of the faller's block disturbed by snig tracks and landings.

The network of snig tracks on the Stockyard shelterwood site was extensive (Fig. 3). There was some re-use of old snig tracks on this site, but not as much as could have been achieved. There were crossing tracks, and old snig tracks that were not re-used in the most recent harvest. This site is a particularly good example of how re-use of old snig tracks could reduce the amount of the harvested area compacted. Similarly there could have been greater re-use of old snig tracks on the Stockyard gap release site (Fig. 4). The pattern of timber extraction was generally rational on the Godfrey gap release (Fig. 2) and Godfrey shelterwood sites (Fig. 1); however, there was some duplication of snig tracks on these sites.

Across all of the FORESTCHECK sites, landing size has varied from 0.4% to 4.8% of the total area of the faller's block. The mean landing size was 2.6% of the total area of the faller's block.

The landing size varies according to the type and size of the harvesting operation, and the volume of timber and the range of timber products harvested from the site. Considering the mean landing size as a proportion of the harvested area, the landing on the Stockyard shelterwood was relatively small, occupying only 0.9% of the harvested area. Landing 1 on the Godfrey shelterwood was a particularly large landing, occupying 3.6% of the harvested area. The landings on the remaining sites were of average or less than average size for operations in the jarrah forest.

Conclusions

- Timber harvesting on the Stockyard shelterwood significantly increased the fine earth bulk density of the surface soil (0-10cm) by 7%.
- Considering that 3 harvesting operations have occurred on this site, this is a relatively small increase in soil bulk density and substantial less than that observed at a similar site (Chalk shelterwood) that was also harvested 3 times, resulting in a 35% increase in soil bulk density.
- The bulk density of the general harvested area away from the snig tracks on the Stockyard shelterwood site was not significantly different from the bulk density of the adjacent area that had never been harvested.
- Considering all of the FORESTCHECK sites so far monitored for soil bulk density, harvesting produces a mean increase in bulk density of the surface soil of 18%.
- Similar differences over a larger number of sites are needed to confidently identify the size of any increase in bulk density caused by harvesting, and the range and variation of this increase.
- Understanding the impact of timber harvesting on the soil bulk density of other FORESTCHECK sites would be greatly enhanced when further measurements of soil bulk density are taken on sites with matching soil types that have never been harvested.
- The network of snig tracks on the Stockyard shelterwood site was extensive. Greater re-use of old snig tracks on this and other sites would reduce the amount of soil compaction caused by timber harvesting.
- Across all of the FORESTCHECK sites, landing size has varied from 0.4% to 4.8% of the total area of the faller's block and the mean landing size was 2.6% of the total area of the faller's block.
- The landing on the Stockyard shelterwood site was relatively small, occupying only 0.9% of the harvested area. Landing 1 on the Godfrey shelterwood was a particularly large landing occupying 3.6% of the harvested area. The landings on the remaining sites were on average or less than average size for operations in the jarrah forest.

Future tasks

The planned work for this year's monitoring is complete. The snig track widths used to calculate the area of the snig tracks were mean values determined from earlier work at other sites in the jarrah forest (Whitford 2001). The addition of measurements of snig track widths on the individual FORESTCHECK sites would increase the value of the snig track areas presented here. It could be helpful in the final analysis of this data to have particle size analysis, soil descriptions and possibly soil mapping for all of the FORESTCHECK sites, but particularly for the 10 sites listed in Table 4 where soil bulk density has been measured. As many of the control sites have been harvested at least once, and have thus experienced soil disturbance which is slow to ameliorate, an attempt will be made in the next year's sampling to collect bulk density measurements from some additional sites which have never been harvested. These sites would be used as reference sites for the existing harvested FORESTCHECK sites. This additional sampling would be of areas adjacent to or near existing FORESTCHECK sites.

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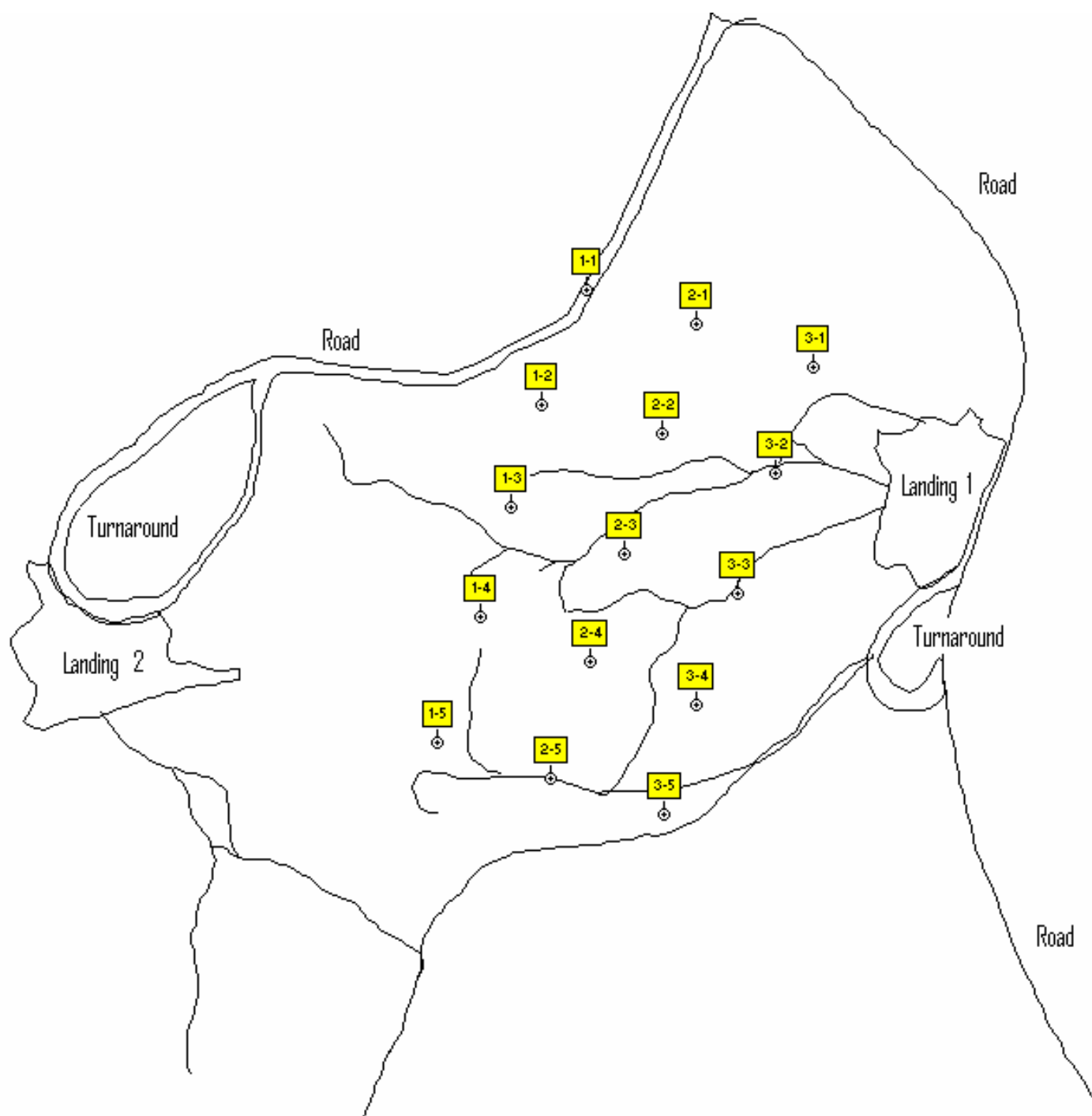


Figure 1. FORESTCHECK site FC30, Godfrey forest block shelterwood showing the layout of the snig tracks, the landings and the FORESTCHECK sampling grid. Scale is given by the grid point spacing of 50 metres.

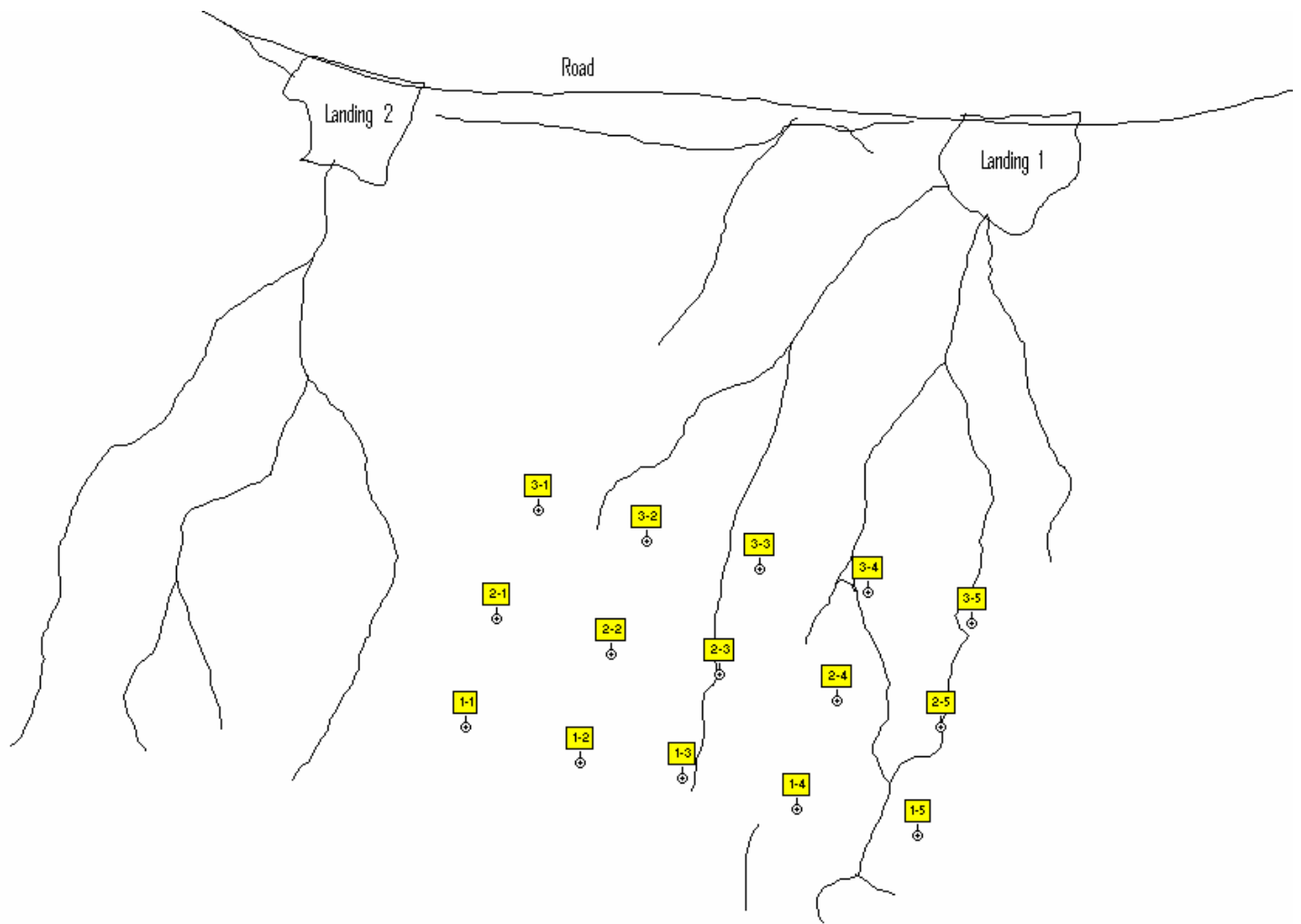


Figure 2. FORESTCHECK site FC31, Godfrey forest block gap release, showing the layout of the snig tracks, landings and the FORESTCHECK sampling grid. Scale is given by the grid point spacing of 50 metres.

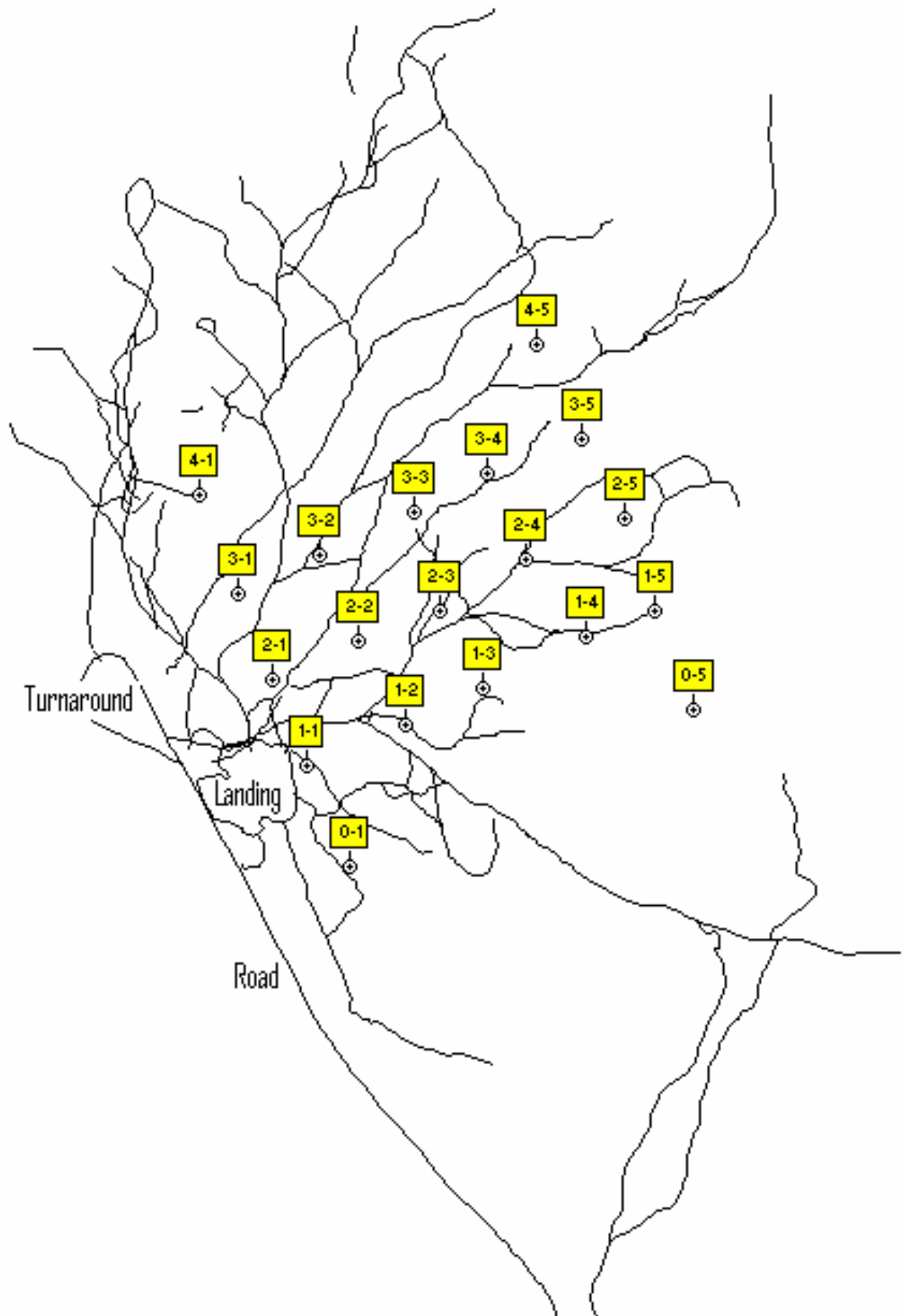


Figure 3. FORESTCHECK site FC33, Stockyard forest block shelterwood, showing the layout of the snig tracks, the landing and the FORESTCHECK sampling grid. Scale is given by the grid point spacing of 50 metres.

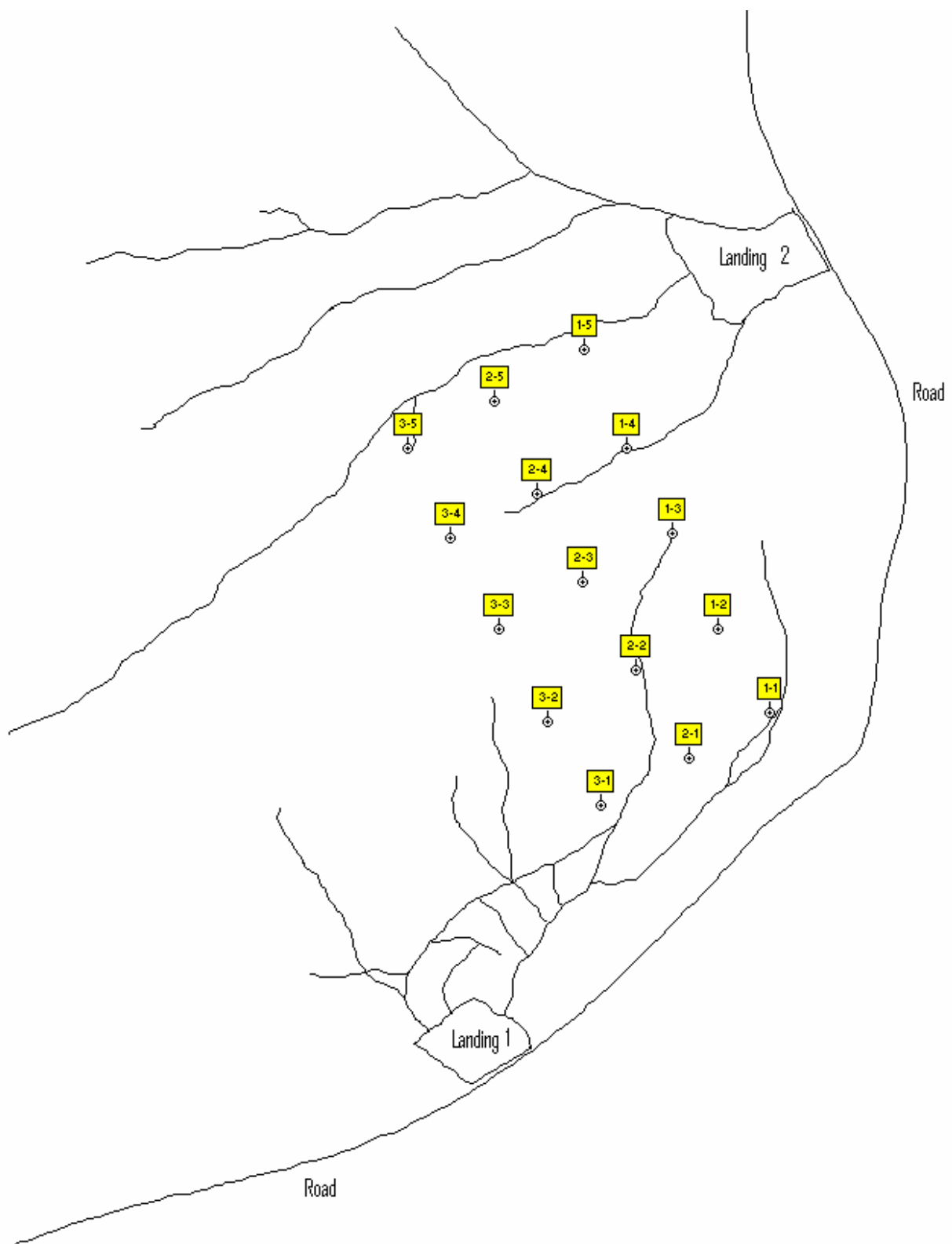


Figure 4. FORESTCHECK site FC34, Stockyard forest block gap release, showing the layout of the snig tracks, the landings and the FORESTCHECK sampling grid. Scale is given by the grid point spacing of 50 metres.

COARSE WOODY DEBRIS, SMALL WOOD AND TWIGS, AND LITTER

Bob Smith and Richard Robinson

Introduction

The quantity of wood and leaf debris on the forest floor contributes to the habitat of fungi, small reptiles, mammals and invertebrates. It is therefore of some importance to ascertain the amount of debris in each area before and after disturbance such as logging and burning. The amount of litter also affects the soil moisture, which in conjunction with micro-organisms affects the soil texture.

This component of FORESTCHECK is intended to:

- Measure and record the amount of litter, small wood and twigs (SWT) and coarse woody debris (CWD) on the ground in the various managed Jarrah forest treatments (i.e.: Gap Release and Shelterwood) and in uncut forest.
- Analyse trends within and between the treatments over time.
- Make the data available for analysis of distribution patterns of other organisms such as invertebrates, small mammals, fungi and cryptogams.

A wildfire burnt through the set of sites in Bell forest block prior to measurement. The litter and SWT were completely destroyed in the fire and as a result were not measured at these sites in 2005. The CWD was measured, however, as the larger logs were still present even though a proportion of them were charred and reduced in size.

Monitoring and Laboratory Work

Sampling in 2004-05 was carried out from 8-10 March 2005, in conjunction with the soil and leaf nutrient sampling and stand structure measurement. The litter and small wood and twigs (SWT) samples were oven dried, weighed in grams and then converted to tonnes per hectare ($t\ ha^{-1}$). The volume of coarse woody debris (CWD) was determined using the line intersect method (van Wagner, 1968) and calculated to cubic metres per hectare ($cm^3\ ha^{-1}$).

Results and Discussion

Litter Weights

Litter weights reflected the time since the last burn (Fig. 1). The heaviest litter weights were found on the external control sites. The Nalyerin site (FC28), with 17-year-old fuels combined with a dense canopy, resulted in the heaviest litter load. Within the silvicultural treatments, the Stockyard shelterwood (FC33) and gap release (FC34) treatments had both been unburnt for 6 years and carried heavier litter loads than the Godfrey forest block treatments (FC30, FC31) that had been unburnt for only 2 years. The Bell forest block sites (FC35, FC36, FC37) were all burnt in a summer wildfire and had no litter to be measured.

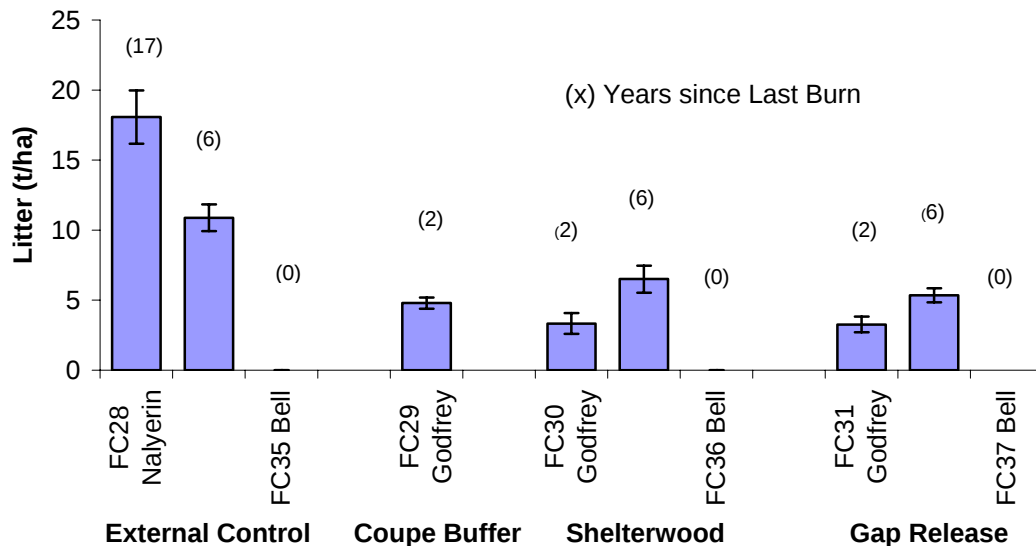


Figure 1. Mean litter loads ($\text{t ha}^{-1} \pm \text{se}$) calculated at each FORESTCHECK site at Wellington East in March 2005. Note that the Bell forest block grids were burnt by wildfire in December 2004.

Small Wood and Twigs

The amount of small wood and twigs carried on all sites was very light (Fig. 2), with the heaviest load being less than 0.7 t ha^{-1} on the Stockyard shelterwood site (FC33). Again, the entire SWT component on the Bell forest block sites was consumed in the fire and not measured.

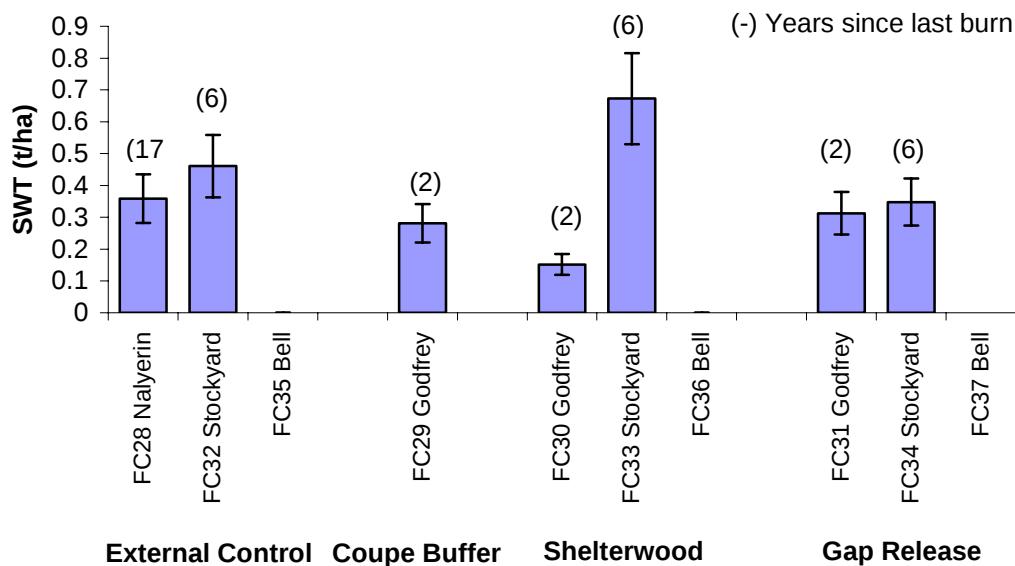


Figure 2. The average weights ($\text{t ha}^{-1} \pm \text{se}$) of small wood and twigs measured at each FORESTCHECK site at Wellington East in March 2005. Note that the Bell forest block grids were burnt by wildfire in December 2004.

Coarse Woody Debris

The largest volume of CWD was found on the Stockyard shelterwood (FC33) and gap (FC34) treatments (Fig. 3). The fire undoubtedly affected the amount of CWD in Bell forest block and a large percentage of the larger logs were obviously reduced in size but not totally consumed. However, both the Bell external control (FC35) and shelterwood (FC36) treatments contained similar or larger volumes of CWD compared to the other grids in those treatments.

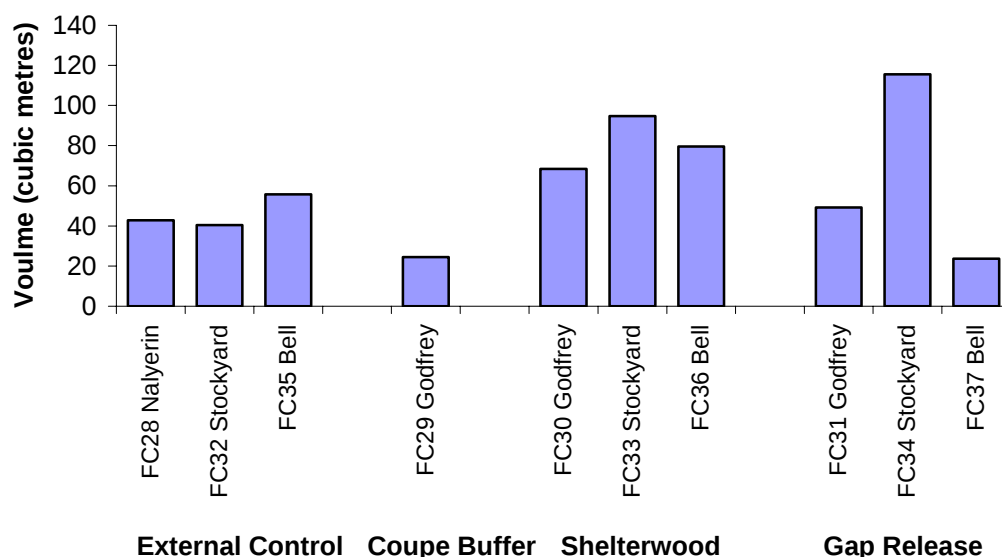


Figure 3. The volume ($\text{m}^3 \text{ ha}^{-1}$) of coarse woody debris measured at each FORESTCHECK site in Wellington East in March 2005. Note that the Bell forest block grids were burnt by wildfire in December 2004.

Conclusions

- Litter and SWT loads reflect the time since last burn
- Recent logging activity may increase the volume of CWD

Data Management

The data were entered into Microsoft Excel worksheets.

Issues

Full assessment was hampered by the fire in Bell forest block. This should not affect future assessments, however, as prescribed burns will be part of the normal silvicultural management within treatments in the future, and natural wildfire may occur at any time.

Reference

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MACROFUNGII

Richard Robinson, Katrina Syme and Bob Smith

Introduction

Fungi are considered one of the most important group of forest organisms in terms of both biodiversity and function. Soil, litter and wood inhabiting fungi play major roles in decomposition and nutrient cycling. Mycorrhizal fungi enhance nutrient uptake of plants and may enhance plant resistance to some pathogens. In addition, underground truffle-like fungi are an important food source for small mammals, especially following disturbance such as fire.

Research on fungi in Western Australia's southern forests is in its infancy. Knowledge on fungal diversity and the ecological roles that fungi play is of vital importance to forest managers making decisions on sustainable forest management.

The objective of this component of the FORESTCHECK program is to:

- Monitor and record the species of macrofungi in the various treatments of managed jarrah forest (gap, shelterwood) and in uncut forest.
- Analyse trends in species composition, richness and abundance and substrate utilization over time.
- Generate detailed descriptions of unknown or unnamed species.

Monitoring and Laboratory work

Transects to monitor macrofungi were installed at the Wellington East sites during grid establishment in September-October 2004. The sites were monitored in June and again in July 2005. Donnelly FORESTCHECK sites (initially established and monitored in 2002) were also monitored in June 2005.

This report deals primarily with the results of monitoring undertaken on the Wellington East sites, however, a brief report on results obtained at the Donnelly sites in 2005 is also included.

1. WELLINGTON EAST

Monitoring

Monitoring dates were selected on the basis of rainfall and soil dryness index (SDI). Statistics were supplied by the CALM Collie Work Centre, and viewed in conjunction with the Bureau of Meteorology online data from the Wandering automatic weather station. Consistent rainfall in early June resulted in the sites being monitored from 22-24 June and again from 4-7 July. Despite consistent rain throughout this period, it was noticeable that the litter dried out considerably between rain events. This is also reflected in the SDI statistic, which increased to about 145 following each rainfall event in late June and throughout July (Fig. 1).

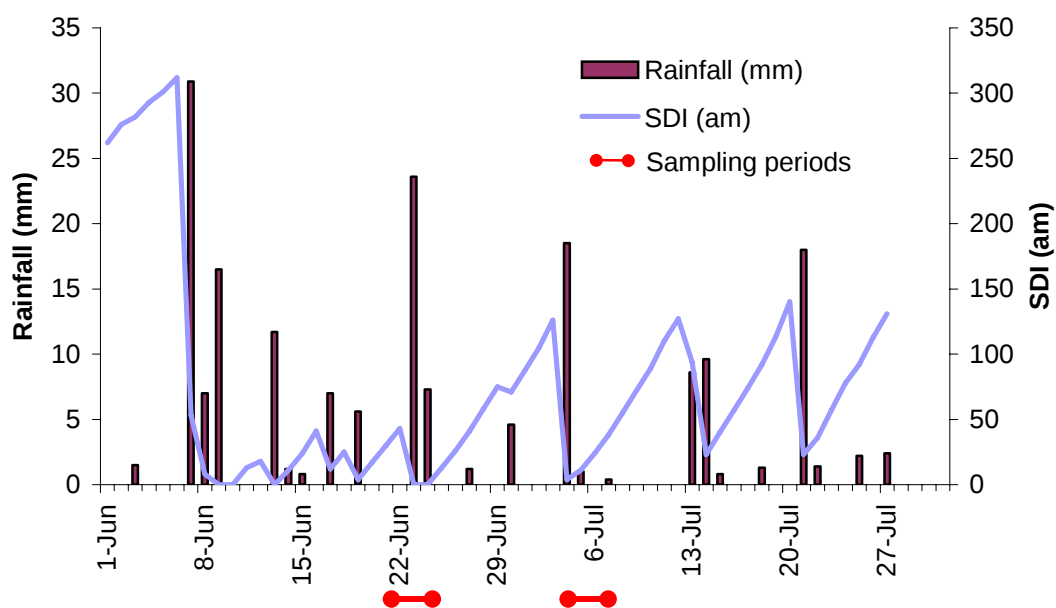


Figure 1. Daily rainfall and soil dryness index (SDI) calculated for Wellington District during the period 1 June to 30 July 2004. The FORESTCHECK sampling period is indicated in red.

Voucher Specimens

Voucher specimens were processed and where possible identified. An overall species list and one for each individual site has been compiled. In total, 138 voucher collections were made representing 103 species. Processing of each voucher was completed on the day of collection or the next day. This included photographing in the field, and preparing morphological descriptions of fresh collections. A total of 68 species had not been encountered previously and detailed descriptions were compiled in order to validate their identity. All collections were air dried at 35° C. Microscopic descriptions were carried out to verify the identity of a number of species and to confirm a number of unnamed species. All vouchers have been entered onto the PERTH database and are housed at the Tony Annels Herbarium at the Manjimup Research Centre.

Results and Discussion

Generally, more species were recorded on each grid in the July monitoring period than in June (Fig. 2). A total of 148 species were recorded in June and 185 in July. The abundance of fruitbodies was consistent and reflected the number of species, 5 204 in June and 6 849 in July. The weather and soil conditions were very similar during each period (Fig. 1).

In all, 240 species of fungi were recorded on the Wellington East sites (Appendix 1). Of these, 28% (68 species) were recorded for the first time in FORESTCHECK. A total of 12 053 fruitbodies were recorded (Appendix 1). This is the most number of species of macrofungi recorded in one year for FORESTCHECK to date, but the abundance of fruitbodies was not as high as that recorded in Wellington and Donnelly in 2003 (18 389 and 14 356 respectively).

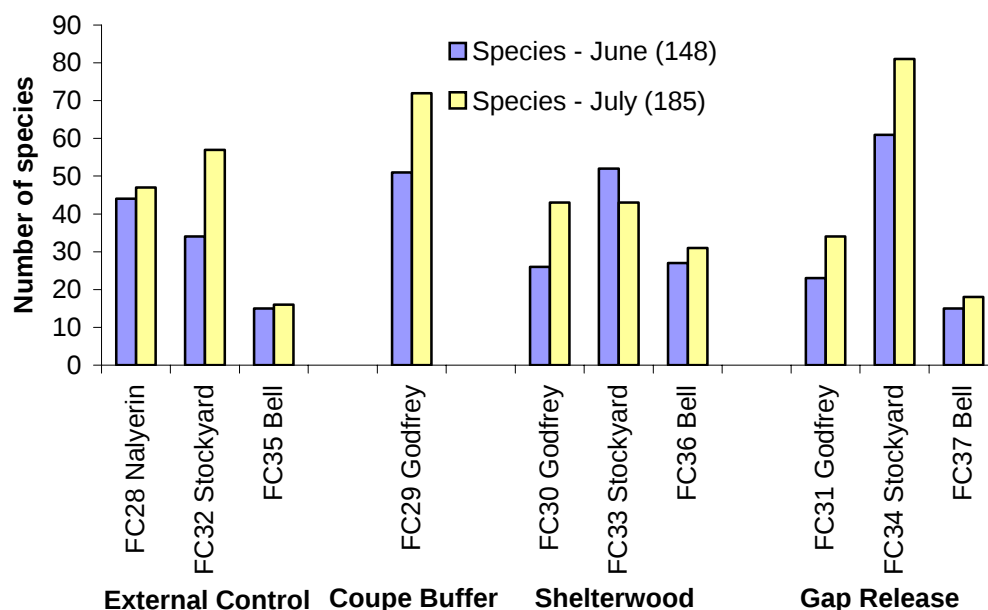


Figure 2. The number of species recorded in June and July at the Wellington East FORESTCHECK sites.

The highest numbers of species were recorded in the Godfrey coupe buffer (FC29) and the Stockyard gap release (FC34). The Bell forest block sites (FC35, FC36 and FC37) all had low numbers of species (Fig. 3). This was due to all 3 sites having been burnt by wildfire on December 28, 2004. On visiting the sites 10 days after the fire, the sclerotial polypore *Laccocephalum tumulosus*, was noted at each site but only recorded and collected from the Bell gap release (FC37) grid. During the regular autumn monitoring, species of typical post-fire macrofungi were recorded on all the burnt grids including Ascomycetes such as *Peziza tennacella*, *Peziza praetervisa*, *Anthrocobia* spp. and *Pulvinula archerii*. In addition a small number of Basidiomycetes such as *Cortinarius* sp., *Hebeloma* sp. and *Phaeocollybia* sp. fruiting on the burnt soil amongst scorched leaf litter and a species of *Dacromyces* fruiting on burnt wood were also recorded.

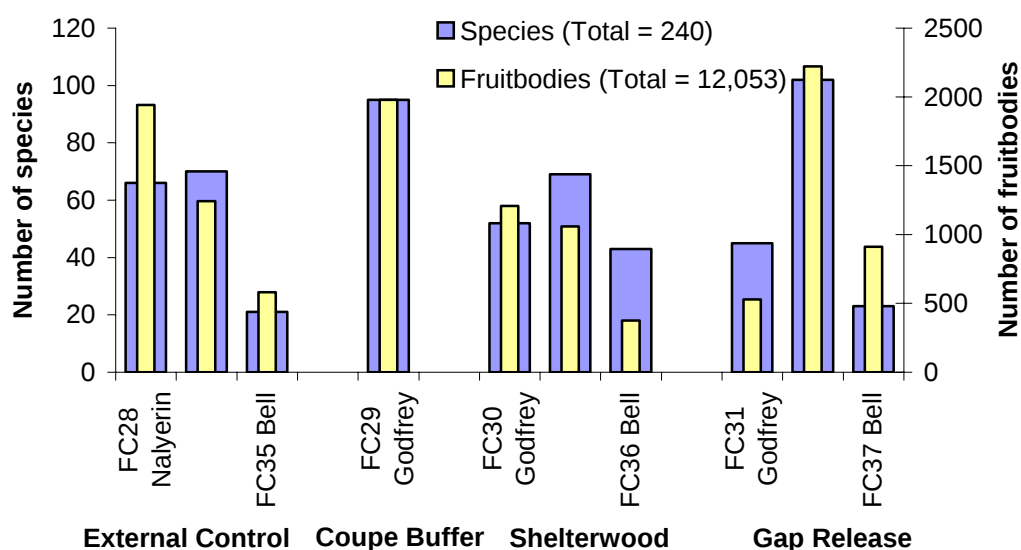


Figure 3. The total number of species and abundance recorded at the Wellington East FORESTCHECK sites in 2005.

Generally, in each treatment the majority of species were recorded fruiting on the soil (Fig. 4), and these species produced the majority of fruitbodies (Fig. 5), except in the Stockyard gap release (FC34) where the most fruitbodies were recorded on wood. The Nalyerin external control had the highest number of fruitbodies associated with the litter layer, about twice as many as recorded in the Stockyard external control. This corresponds well with the fire history of the grids, Nalyerin having been unburnt for 11-15 years longer than the other grids and consequently having the heaviest load of litter recorded (see chapter on CWD and Litter).

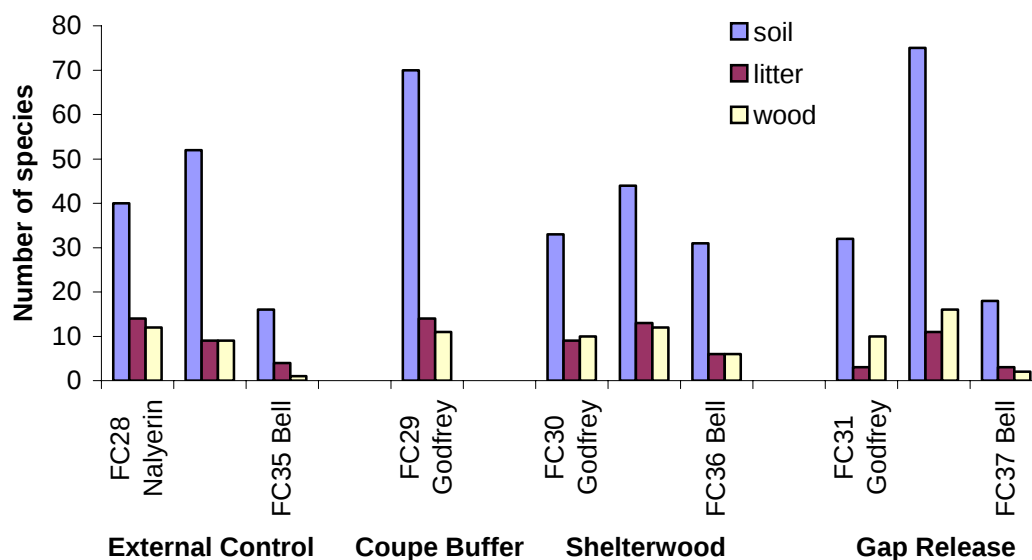


Figure 4. The number of species recorded fruiting on soil, litter and wood on the Wellington East FORESTCHECK sites in 2005.

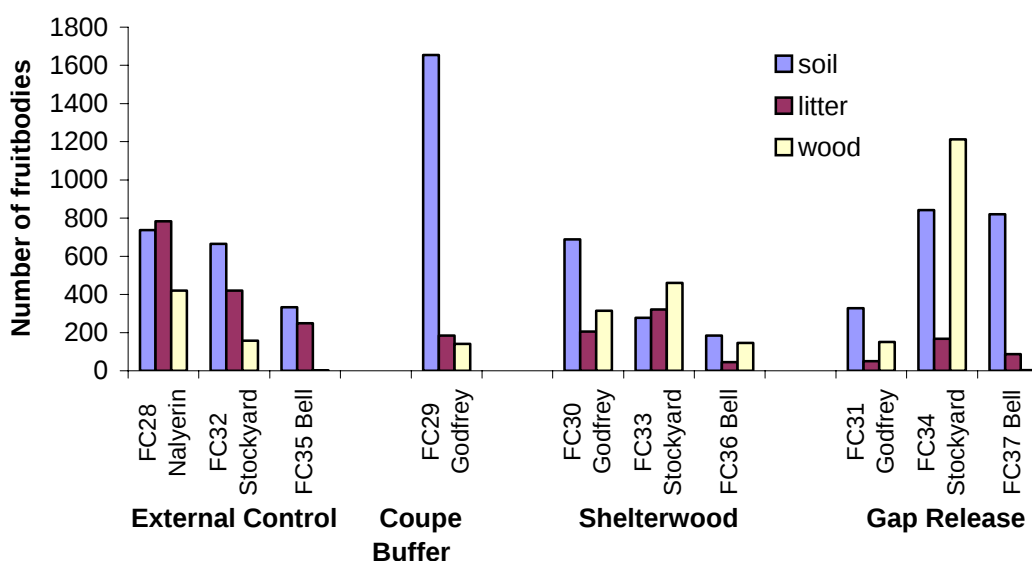


Figure 5. The number of fruitbodies recorded on litter, soil and wood in each grid on the Wellington East FORESTCHECK sites in 2005.

In general, there appears to be no obvious effect of logging on the species richness of macrofungi. However, the low number of species recorded on all the Bell forest block treatments can be attributed to the grids having been burnt. The burnt grids also had a distinct set of species associated with them. This illustrates the importance of monitoring immediately following a disturbance such as wildfire in order to record fluctuations in species composition associated with such disturbance.

2. DONNELLY

Monitoring

The Donnelly sites were established in 2002. They have been monitored annually and in 2005 this was carried out from 13-20 June. Monitoring coincided with a drop in the SDI following consistent rain in early June.

Voucher Specimens

87 voucher collections were made, representing 68 species.

Results and Discussion

A total of 230 species and 15,324 fruitbodies were recorded across all the sites. 43 species were recorded for the first time on the Donnelly sites. There appeared to be no major differences in species richness on grids, either between or within treatments (Fig 6). However, the abundance of fruitbodies was unusually high on the Kingston gap release treatment (FC2). The low number of species and fruitbodies recorded on the Yornup external control (FC5) is likely due to that grid having been burnt 18 months previously.

Although this is the most number of species recorded for any single year at Donnelly, the abundance of fruitbodies was not as high as that recorded in 2003. Variation in species richness within the external controls appears to be very variable from year to year, but is consistently high in the gap release treatment (Fig. 7).

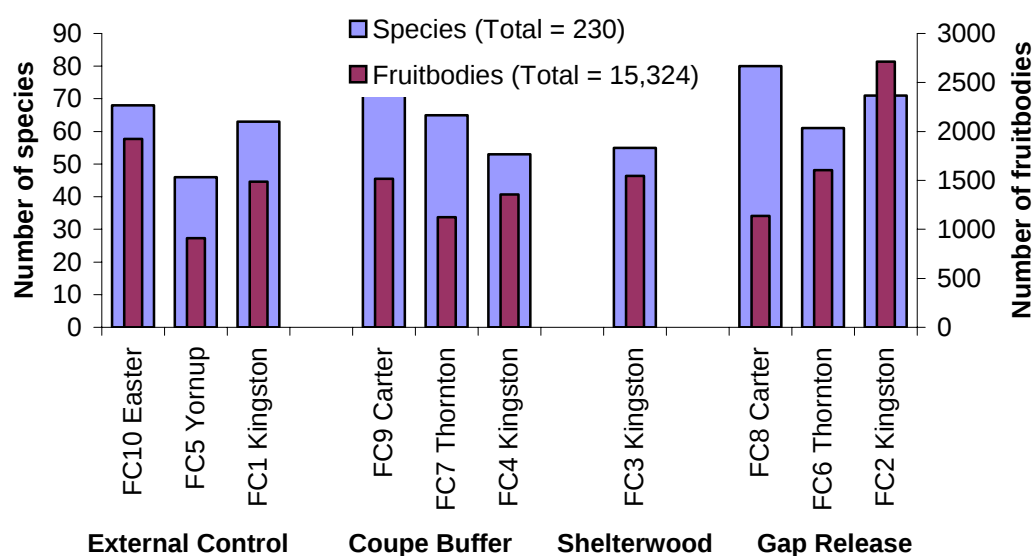


Figure 6. The total number of species and abundance recorded at the Donnelly FORESTCHECK sites in 2005.

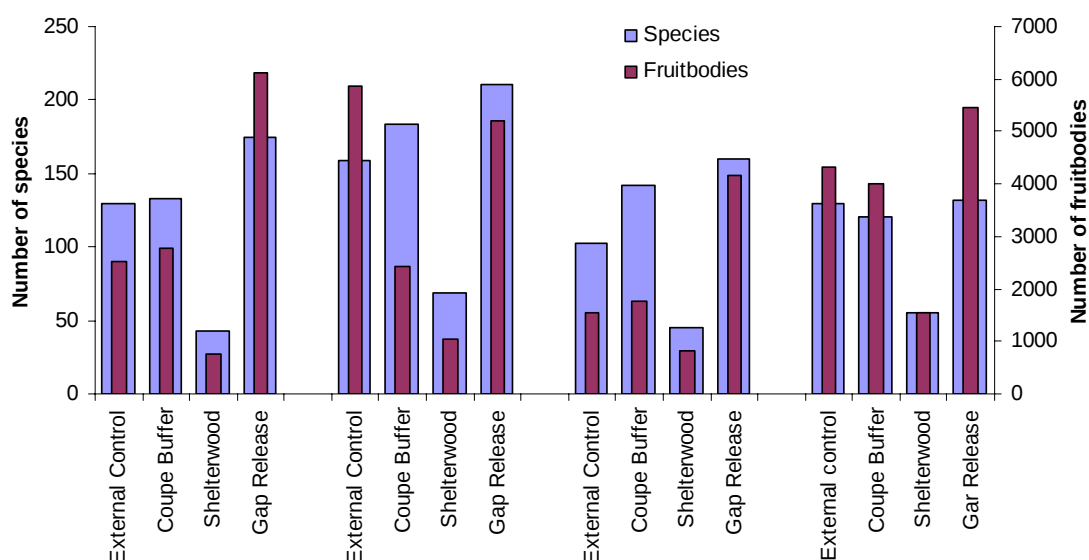


Figure 7. The total number of species and abundance recorded in each treatment in the Donnelly FORESTCHECK sites from 2002-05 (Note that there was only one shelterwood site at Donnelly).

The species list for Donnelly continued to increase, with 43 additions in 2005. This illustrates the value of long-term monitoring in documenting macrofungal diversity on sites and across the landscape.

Conclusions

- A total of 482 species of macrofungi have so far been recorded in FORESTCHECK.
- 352 species were recorded in 2005, 240 at Wellington East and 230 at Donnelly. Of these 118 species were recorded at both locations, 122 were restricted to Wellington East, and 112 were restricted to Donnelly.
- Overall, many species appear to have restricted distribution, but many others are common throughout the various jarrah forest environments.
- Distinct species of macrofungi fruit following recent fire.
- The majority of macrofungi recorded in Wellington East fruit on the soil.
- Gap release treatments in Wellington East varied markedly in macrofungal species richness and abundance.

Data Management

All data have been entered onto a Microsoft Excel worksheet. Species diversity and abundance at each site and a frequency rating of 1 (rare) to 8 (very common) for each species at each site has been determined. The data includes a complete list of species recorded across all the sites, their life modes (mycorrhizal, saprotrophic, parasitic) and the substrate on which they were fruiting. Analysis is ongoing.

Appendix 1. Master list of macrofungi recorded in FORESTCHECK 2002-05, and the species and abundance recorded in each treatment at Wellington East and the species and abundance recorded at Donnelly in 20005

Sp #	Species	Life Mode ¹	Substrate ²	Treatments ³				Well. East	Donnelly
				E.C.	C.B.	SW.	G.R.	Total	Total
349	Agaric "brown, brown decurrent gills"	S?	S				8	8	
475	Agaric "creamy brown scaly cap, white bifurcate gills"	S?	S						
46	Agaric "creamy white"	S	S						
82	Agaric "Lepiota-like, cream-grey"	S	S						
156	Agaric "light brown - red scales on stem"	S	S						
18	Agaric "light brown-olive"	S	S						
473	Agaric "orange"	S?	S						3
97	Agaric "pure white"	?	S						
174	Agaric "red/yellow/red"	S	??						
329	Agaric "viscid buff, long stem"	M?	S						
170	Agaric "yellow brown-moist"	S	S/L						
240	<i>Agaricus</i> sp. "small with red brown fibrils"	S	S			1		1	3
38	<i>Agaricus</i> sp. "small"	S	S	1				1	
71	<i>Agaricus</i> sp. "small, flat- red stain"	S	S						3
33	<i>Agaricus</i> sp. "yellow stainer"	S	S						
39	<i>Agaricus</i> sp. "large cap, purplish scales"	S	S		2	3		5	1
120	<i>Aleuria rhenana</i>	S	S		2	23	43	68	83
126	<i>Aleurina ferruginea</i>	S	S/Moss	2	2		5	9	15
206	<i>Amanita ananiceps</i>	M	S			3	2	5	
186	<i>Amanita brunneibulbosa</i> "grey-brown"	M	S		1			1	
283	<i>Amanita eucalypti</i>	M	S			2		2	
269	<i>Amanita ochrophylloides</i>	M	S						
114	<i>Amanita</i> sp. "apricot-pink margin"	M	S						
518	<i>Amanita</i> sp. "beige with saccate base"	M	S				1	1	
520	<i>Amanita</i> sp. "creamy yellow, sticky cap"	M	S				1	1	
493	<i>Amanita</i> sp. "grey brown robust"	M	S				2	2	
497	<i>Amanita</i> sp. "grey brown with orange yellow veil"	M	S			1		1	
496	<i>Amanita</i> sp. "grey veil"	M	S			7		7	
360	<i>Amanita</i> sp. "large grey-white, robust"	M	S						
526	<i>Amanita</i> sp. "small creamy white, membranous ring"	M	S			1		1	
320	<i>Amanita</i> sp. "small robust, yellow-buff, bulbous base"	M	S		1			1	
368	<i>Amanita</i> sp. "white with mealy stem"	M	S						
371	<i>Amanita</i> sp. "white with saccate volva"	M	S			4		4	
45	<i>Amanita</i> sp. "white, deeply rooted"	M	S		1		1	2	
525	<i>Amanita</i> sp. "white, grey scales, silvery stem"	M	S			1		1	
28	<i>Amanita</i> sp. "white, stout"	M	S						
519	<i>Amanita</i> sp. "yellow brown, long stem, constricted bulb"	M	S			1	1	2	
218	<i>Amanita</i> sp. "powdery - long tapering base"	M	S		1	10	6	17	
196	<i>Amanita umbrinella</i>	M	S						
6	<i>Amanita xanthocephala</i>	M	S	2	13	12	15	42	32

Sp #	Species	Life Mode	Sub- strate	Treatments				Well. East	Donn- elly
				E.C.	C.B.	SW.	G.R.	Total	Total
35	<i>Amanita xanthocephala</i> forma <i>macalpiniana</i>	M	S				2	2	
531	<i>Amanita</i> spp. (Unidentified)	M	S		1	3	1	5	
509	<i>Anthracophyllum archeri</i>	S	T						10
338	<i>Anthrocobia muelleri</i>	S	S	62			300	362	
338b	<i>Anthrocobia muelleri</i> "small yellow"	S	S				130	130	
180	<i>Armillaria luteobubalina</i>	P/S	W						43
188	<i>Austroboletus laccunosa</i>	S	S			2		2	3
200	<i>Austroboletus occidentale</i>	S	S						1
291	<i>Austropaxillus</i> sp. "orange-brown"	M	S	11	13	24	4	52	
436	<i>Beauvaria bassiana</i>	P	Insect	1				1	
93	<i>Boletellus ananiceps</i>	S	S	4		2		6	3
103	<i>Boletellus obscurecoccineus</i>	S	S	1	2			3	2
225	<i>Boletus</i> sp. "creamy pale yellow"	M	S						
29	<i>Boletus</i> sp. "dull maroon"	M	S				2	2	1
345	<i>Boletus</i> sp. "light yellow"	M	S						
350	<i>Boletus</i> sp. "pink maroon cap, yellow/red stem"	M	S						
49	<i>Boletus</i> sp. "red pores and stem"	M	S						
253	<i>Boletus</i> sp. "red-brown/golden yellow - intense blue stain"	M	S				1	1	
95	<i>Boletus</i> sp. "small yellow/cream pores"	M	S						
358	<i>Boletus</i> sp. "viscid brown cap, yellow marshmallow pores"	M	S						
99	<i>Boletus</i> sp. "yellow-red, stains blue"	M	S						1
216	<i>Boletus</i> sp. "brown/yellow pores which stain blue"	M	S						
210	<i>Boletus</i> sp. "maroon/orange pores"	M	S						
195	<i>Boletus</i> sp. "mustard brown-brown stain"	M	S						
193	<i>Boletus</i> sp. "purple brown"	M	S						
284	<i>Boletus</i> sp. "under <i>Allocasuarina</i> "	M	S						
208	<i>Boletus</i> sp. "yellow-brown, cracked/white pores"	M	S						
304	<i>Byssomerulius corium</i> .	S	W/T	2		15		17	36
9	<i>Calocera</i> sp. "yellow"	S	W		11	345	429	785	2587
465	<i>Calostoma fuscum</i>	S	S						3
463	<i>Cantharellus cibarius</i> var. <i>australiensis</i>	M	S						12
470	<i>Ceratiomyxa fruticulosa</i>	Bacteria	W						6
265	<i>Cheilymenia</i> sp. "eyelash on roo poo"	C	Dung						
243	<i>Cheilymenia</i> sp. "orange disks on marri nuts"	S	Fruits						57
364	<i>Chlorociboria aeruginascens</i>	S	W						
377	<i>Clavaria</i> "small lemon yellow"	S	S	1		2	1	4	39
316	<i>Clavaria</i> (<i>Clavulinopsis</i>) aff. <i>aurantiaca</i> "orange"	M?	S						10
319	<i>Clavaria</i> (<i>Clavulinopsis</i>) sp. "grey- brown with black tips"	M?	S			1	3	4	16
534	<i>Clavulina amethystina</i>	S	S				1	1	
81	<i>Clavulina</i> cf. <i>cinerea</i> "grey-brown"	S	S	1			4	5	138
344	<i>Clavulina</i> sp. "cream, fluffy tips"	M?	S						20
140	<i>Clavulina</i> sp. "pink-buff coral"	S	S			2		2	
458	<i>Clavulina</i> sp. "pinkish brown, red-brown tips"	S	S	8	2		6	19	147

Sp #	Species	Life Mode	Substrate	Treatments				Well. East	Donnelly
				E.C.	C.B.	SW.	G.R.	Total	Total
362	<i>Clavulinopsis</i> "grey brown, black tips"	M?	S						54
472	<i>Clavulinopsis</i> sp. "coral pink"	S	S						10
261	<i>Clavulinopsis</i> sp. "cream"	S	S						
262	<i>Clavulinopsis</i> sp. "tiny white candles"	S	S						
197	<i>Clitocybe semi oculata</i>	S	W						4
197b	<i>Clitocybe semi-occulta</i> "large"								
14	<i>Clitocybe</i> sp.	S	S						
23	<i>Clitocybe</i> sp.	S	S						
370	<i>Clitocybe</i> sp. "creamy beige"	S	S/L						
324	<i>Clitocybe</i> sp. "grey robust"	S	S						
301	<i>Clitocybe</i> sp. "dark grey with dimple"	S	S		1		1	2	
181	<i>Collybia</i> aff. <i>butracea</i>	S	S						1
143	<i>Collybia</i> sp. "buff funnel"	S	S						10
249	<i>Collybia</i> sp. "grey"	S	S				3	13	2
233	<i>Collybia</i> sp. "grey/dimple"	S	S				1	1	
489	<i>Collybia</i> sp. "large brown, tan gills"	S	S				2	2	
151	<i>Collybia</i> sp. "large"	S	S				1	1	
513	<i>Collybia/Clitocybe</i> "dark grey-brown convex"	S	S		1	4		5	
15	<i>Coltricia oblectans</i>	S	S	48	67	45	60	220	303
532	<i>Coltriciella dependens</i>	S	W				3	3	
32	<i>Coprinus</i> sp.	S	S/L						
128	<i>Coprinus</i> sp. "basal hairs"	S	S						112
224	<i>Coprinus</i> sp. "micacus"	S	S						
460	<i>Cordyceps</i> sp. "brown club"	P	Insect						11
467	<i>Cordyceps</i> sp. "orange-brown club"	P	Insect						2
282	<i>Corinarius</i> sp. "honey-brown dome /long stem"	M	S						
303	<i>Cortinaius</i> sp. "stubby domes"	M	S						
511	<i>Cortinaius</i> sp. "multi red"	M	S		12			12	
146	<i>Cortinarius (Myxacium)</i> sp. "orange-brown"	M	S		8	3	1	12	3
125	<i>Cortinarius (Phlegmacium)</i> sp. "purple-grey"	M	S						
158	<i>Cortinarius</i> aff. <i>micro archerii</i>	M	S		6			6	1
314	<i>Cortinarius archerii</i>	M	S						
207	<i>Cortinarius australiensis</i>	M	S				1	1	
173c	<i>Cortinarius basirubescens</i> " brown large"	M	S		1			1	
173	<i>Cortinarius basirubescens</i> (red cap)	M	S						
173b	<i>Cortinarius basirubescens</i> (brown cap)	M	S				5	5	
115	<i>Cortinarius fibrillosus</i>	M	S						1
7	<i>Cortinarius radicans</i>	M	S		2			2	
293	<i>Cortinarius rotundisporus</i>	M	S	1				1	
357	<i>Cortinarius sinapicolor</i>	M	S			1		1	2
234	<i>Cortinarius</i> sp.	M	S						
485	<i>Cortinarius</i> sp.	M	S		4			4	
421	<i>Cortinarius</i> sp. "brown cap, lilac white stem"	M	S	8	1	4	3	16	
355	<i>Cortinarius</i> sp. "brown with lavender margin and stem"	M	S						
73	<i>Cortinarius</i> sp. "brown with purplish tints"	M	S						

Sp #	Species	Life Mode	Sub- strate	Treatments				Well. East	Donn- elly
				E.C.	C.B.	SW.	G.R.	Total	Total
466	<i>Cortinarius</i> sp. "brown with white margin"	M	S						2
68	<i>Cortinarius</i> sp. "brown" ?(34)	M	S	1				1	
346	<i>Cortinarius</i> sp. "brown, grey-lavender gills"	M	S						
232	<i>Cortinarius</i> sp. "cf sinapicolor"	M	S		1			1	
500	<i>Cortinarius</i> sp. "chestnut large"	M	S	101				101	
382	<i>Cortinarius</i> sp. "chestnut with yellow margin and yellow flesh"	M	S						
154	<i>Cortinarius</i> sp. "chestnut"	M	S	30	15	6	10	61	9
453	<i>Cortinarius</i> sp. "decurrent gills, deep stem with double ring"	M	S			12		12	6
334	<i>Cortinarius</i> sp. "fawn brown"	M	S						
252	<i>Cortinarius</i> sp. "glutinous cap/rooting stem"	M	S						
348	<i>Cortinarius</i> sp. "golden tan, long stem"	M	S						
374	<i>Cortinarius</i> sp. "golden-tan"	M	S	2				2	1
257	<i>Cortinarius</i> sp. "honey-brown"	M	S						
369	<i>Cortinarius</i> sp. "large red-brown"	M	S						
379	<i>Cortinarius</i> sp. "lilac-brown with yellow gills"	M	S						
404	<i>Cortinarius</i> sp. "orange cap, white floccose stem"	M	S				1	1	
251	<i>Cortinarius</i> sp. "orange-brown 2"	M	S						
98	<i>Cortinarius</i> sp. "pointy cap"	S	S	18	19	4	5	46	34
515	<i>Cortinarius</i> sp. "red brown cap, slender lavender stem"	M	S		5			5	
121	<i>Cortinarius</i> sp. "slender brown"	M	S						
131	<i>Cortinarius</i> sp. "slender lilac"	M	S						
432	<i>Cortinarius</i> sp. "tan cap with chocolate gills"	M	S	4				4	
171b	<i>Cortinarius</i> sp. "vinaceus lilac"	M	S	5				5	18
96/259	<i>Cortinarius</i> sp. "viscid - pink"	M	S						3
375	<i>Cortinarius</i> sp. "yellow with brown fibrils and orange ring"	M	S						
237	<i>Cortinarius</i> sp. "yellow with orange brown fibrils"	M	S	5				5	6
492	<i>Cortinarius</i> sp. "yellow with yellow stem"	M	S	4		1	2	7	
354	<i>Cortinarius</i> sp. "yellow-brown cap, lavender gills and stem"	M	S	3	1			4	
231	<i>Cortinarius</i> sp. "yellow-brown/tan margin"	M	S		2			2	
124	<i>Cortinarius</i> sp. "yellow-olive"	M	S						
255	<i>Cortinarius</i> sp. "yellow-orange"	M	S						
279	<i>Cortinarius</i> sp. "brown fibrillose"	M	S						
244	<i>Cortinarius</i> sp. "brown umbonate"	M							
299	<i>Cortinarius</i> sp. "chocolate brown with mustard gills"	M	S	1	12			13	
201	<i>Cortinarius</i> sp. "cream with orange gills"	M	S						
212	<i>Cortinarius</i> sp. "orange brown"	M	S						
230	<i>Cortinarius</i> sp. "orange viscid"	M	S						
223	<i>Cortinarius</i> sp. "orange"	M	S						
205	<i>Cortinarius</i> sp. "orange/yellow flesh/yellow gills"	M	S		2			2	

Sp #	Species	Life Mode	Substrate	Treatments				Well. East	Donnelly
				E.C.	C.B.	SW.	G.R.	Total	Total
267	<i>Cortinarius</i> sp."snowy chestnut"	M	S	11	2			13	2
270	<i>Cortinarius</i> sp."viscid, yellow-red-brown, white stem"	M	S		1			1	1
273	<i>Cortinarius</i> sp."white with deep rooting stem"	M	S						
199	<i>Cortinarius</i> sp."yellow orange"	M	S						
184	<i>Cortinarius</i> spp. (unidentified)	M	S		9	5	9	23	11
171	<i>Cortinarius vinaceolamellatus</i>	M	S						
290	<i>Cortinarius violaceus</i>	M	S						
469	<i>Crepidotus</i> sp. "chestnut with fringed margin"	S	W						9
118	<i>Crepidotus</i> sp. "large creamy-tan"	S	W	4				4	37
323	<i>Crepidotus</i> sp. "rusty brown suede"	S	W						
61	<i>Crepidotus</i> sp. "small brown"	S	W/Bark						19
83	<i>Crepidotus</i> sp. "small creamy tan"	S	Bark/W						5
21	<i>Crepidotus</i> sp. "small white"	S	W	75				75	25
241	<i>Crepidotus variabilis</i>	S	T/W	215				215	1799
148	<i>Crucibulum laeve</i>	S	T/L	47	12	222	51	332	12
296	<i>Cyathus</i> sp."on roo poo"	S/C	Dung		10	58		80	
307	<i>Cyathus</i> sp.	S	L	28	2			30	90
138	<i>Daldina concentrica</i>	S	W						5
110	<i>Dermocybe</i> aff. <i>sanguinea</i>	M	S						
147	<i>Dermocybe austroveneta</i>	M	S						2
340	<i>Dermocybe clelandii</i> "mini"	M	S	17	25			42	
57b	<i>Dermocybe clelandii</i> "olive brown"								
57/34	<i>Dermocybe clelandii</i> (white mycelium)	M	S	5	7	1	2	15	3
172b	<i>Dermocybe clelandii</i> (yellow mycelium - glutinous cap)	M	S						
172	<i>Dermocybe clelandii</i> (yellow mycelium)	M	S				1	1	1
328	<i>Dermocybe</i> sp. "small olive"	M	S						
486	<i>Dermocybe</i> sp. "yellow stipe, yellow mycelium"	M	S						
168	<i>Dermocybe</i> sp. (<i>D. clelandii</i> ?) "brown with mustard yellow gills"	M	S	16				16	
40	<i>Dermocybe</i> sp."chestnut"	M	S						
310	<i>Dermocybe splendida</i>	M	S						1
294	Discomycete "small yellow on <i>Banksia grandis</i> leaf"	S	L						135
508	Discomycete "tiny cream disks on leaves"	S	L						20
462	Discomycete "tiny white on marri nut"	S	Fruits						1
123	Discomycete "yellow stalked"	S	S	88	11	12	22	133	873
31	<i>Entoloma (Leptonia) moongum</i> "blue-black"	S	S			1	5	6	2
78	<i>Entoloma (Leptonia)</i> sp. "grey/decurrent gills"	S	S						
153	<i>Entoloma (Leptonia)</i> sp. "small dark grey-brown"	S	S						
409	<i>Entoloma</i> aff. <i>incana</i>	S	S				6	6	
222	<i>Entoloma</i> sp. "black with grey-white gills"	S	S						
410	<i>Entoloma</i> sp. "blue-black, marginate gills"	S	S				1	1	5
347	<i>Entoloma</i> sp. "brown striate cap"	S	S						

Sp #	Species	Life Mode	Substrate	Treatments				Well. East	Donnelly
				E.C.	C.B.	SW.	G.R.	Total	Total
227	<i>Entoloma</i> sp. "brown-black with tan gills"	S	S				6	6	
530	<i>Entoloma</i> sp. "brown-black, marginate gills, bluish-grey stem"	S	S				2	2	
471	<i>Entoloma</i> sp. "buff with dimple"	S	S						10
30	<i>Entoloma</i> sp. "creamy white"	S	S	5		3	3	11	3
167	<i>Entoloma</i> sp. "dark grey/blue gill edge"	S	S						
25	<i>Entoloma</i> sp. "grey-brown/blue stem"	S	S			1		1	
77	<i>Entoloma</i> sp. "grey-brown/brown stem"	S	S						7
235	<i>Entoloma</i> sp. "grey-brown/grey stem"	S	S	3	3	7	4	17	9
135	<i>Entoloma</i> sp. "tall, grey-brown"	S	S			2	7	9	4
514	<i>Entoloma</i> sp. "very large brown-grey"	S	S		1			1	
198	<i>Entoloma</i> sp. "brown black/tan/blue"	S	S				1	1	
194	<i>Entoloma</i> sp. "brown"	S	S						1
272	<i>Entoloma</i> sp. "grey-brown with dimple"	S	S			4	4	8	4
278	<i>Entoloma</i> sp. "suede grey-brown with dimple"	S	S				6	6	
274	<i>Entoloma viridomarginatum</i>	S	S			5		5	1
159	<i>Exidia glandulosus</i>	S	W						55
187	<i>Favodashia</i> sp. "grey"	S	W						
41	<i>Fistulina hepatica</i>	S	W			1	1	2	2
91	<i>Fistulinella mollis</i>	S	W						3
19	<i>Formitopsis lilacino-gilva</i>	S	W	8	1	2		11	12
11	<i>Galerina</i> sp. "hanging gills" and "conic"	S	S/L	261	57	43	88	449	480
111	<i>Galerina</i> sp. "large"	S	S						
58	<i>Galerina</i> sp. "small cap, eccentric stipe - on wood"	S	W	10	46	10	43	109	571
42	<i>Galerina</i> sp. "small on bark"	S	Bark						
228	<i>Geastrum</i> sp.	S	S/L						8
533	<i>Geoglossum</i> sp.	S	S				1	1	
8	<i>Gymnopilus allantopus</i>	S	W	367	4	58	153	582	168
365	<i>Gymnopilus junionus</i>	S	W						
43	<i>Gymnopilus</i> sp.	S	W						
105	<i>Gymnopilus</i> sp. "chestnut scales, forked gills"	S	W						
517	<i>Gymnopilus</i> sp. "red cap, yellow gills, yellow stem"	S	W			1		1	
26	<i>Gymnopilus</i> sp. "reddish cap, orange gills"	S	W						4
85	<i>Gymnopilus</i> sp. "slender"	S	W	28	19	54	57	158	183
217	<i>Gyroporus</i> aff. <i>cyanescens</i> "yellow suede - intense blue stain"	M	S						
292	<i>Gyroporus</i> sp. "beige-yellow, blue stain"	M	S						
433	<i>Hebeloma</i> aff. <i>westraliensis</i>	S	S				1	1	
498	<i>Hebeloma</i> sp. "small"	S	S	14				14	
56	<i>Heterotexus peziziformis</i>	S	W/T	8		4	45	57	136
422	<i>Hohenbuehelia</i> aff. <i>atracaerulea</i> "grey brown"	S	W		4		10	14	
480	<i>Hydnellum</i> sp. "orange tipped spines"	S	L		1			1	2
87	<i>Hydnellum</i> sp. "red brown"	S	L/S				1	1	169
275	Hydnoid "fleshy funnel"	?	S/L						
297	<i>Hydnum repandum</i>	S?	S				3	3	40
380	<i>Hydnum</i> sp. "chestnut"	S?	S						
476	<i>Hygrocybe</i> aff. <i>astatogala</i>	S	S						2

Sp #	Species	Life Mode	Sub- strate	Treatments				Well. East	Donn- elly
				E.C.	C.B.	SW.	G.R.	Total	Total
381	<i>Hygrocybe cantharellus</i>	M	S	1				1	1
317	<i>Hygrocybe conica</i>	S	S						
122	<i>Hygrocybe</i> sp. "yellow-orange"	S	Moss						2
281	<i>Hygrocybe</i> sp. "pallid yellow"	S	S						
100	<i>Hypholoma australe</i>	S	W						24
59	<i>Hypholoma brunneum</i>	S	W						6
268	<i>Hyphomyces</i> sp. "brown/yellow-orange"	M	Fungus						16
108	<i>Hypomyces chrysospermus</i>	P	Bolete			4	2	6	9
516	<i>Ileodictyon gracile</i>	S	S			5		5	
204	<i>Innonotus</i> sp.	S	W						
1	<i>Inocybe australiensis</i>	M	S	130	201	51	91	473	191
203	<i>Inocybe geophylla</i>	M	S	5				5	
487	<i>Inocybe</i> sp. "brown fibrillose, yellow gills"	M	S	6	10	6	10	32	
378	<i>Inocybe</i> sp. "chocolate umbonate"	M	S						
137	<i>Inocybe</i> sp. "creamy-brown"	M	S						
48	<i>Inocybe</i> sp. "grey"	M	S	3	3	6	11	23	30
484	<i>Inocybe</i> sp. "large firillose, umbonate, yellow-tan gills"	M	S	4	2			6	
65	<i>Inocybe</i> sp. "large scaly cap"	M	S						2
398	<i>Inocybe</i> sp. "large scaly umbonate cap"	M	S				1	1	
226	<i>Inocybe</i> sp. "orange brown"	M	S						
113	<i>Inocybe</i> sp. "radially fibrillose, pink stem"	M	S						17
20	<i>Inocybe</i> sp. "scaly cap" see sp. 277 Fire Fungi	M	S	1	27	3	4	35	53
169	<i>Inocybe</i> sp. "shaggy stem"	M	S	4				4	2
162	<i>Inocybe</i> sp. "small light brown, fibrillose"	M	S						
53	<i>Inocybe</i> sp. "tan skirt"	M	S	2	51	12	4	69	26
286	<i>Inocybe</i> sp. "umbonate, shaggy"	M	S	2			7	9	5
74	<i>Laccaria</i> aff. <i>masonii</i>	M	S	127	512	422	286	1347	974
36	<i>Laccaria lateritia</i>	M	S	18	52	17	3	90	57
384	<i>Laccocephalum basilapiloides</i>	S	W						
383	<i>Laccocephalum tumulosum</i>	S	W						
221	<i>Lactarius clarkeae</i>	M	S	13		2		15	10
142	<i>Lactarius eucalypti</i>	M	S	2				2	11
245	<i>Lactarius</i> sp. "cream yellow"	M	S	33	20	75	22	150	22
215/220	<i>Lactarius</i> sp. "cream custard"	M	S	2		1		3	
478	<i>Laetiporus potentosus</i>	S	W	4		1	1	6	3
335	<i>Lentinellus</i> sp. "brown cap, saw-toothed gills"	S	W						
457	<i>Lentinellus</i> sp. "brown fan, white saw-gills"	S	W						1
468	<i>Lentinellus</i> sp. "brown lobed, hirsute, brown gills"	S	W						6
271	<i>Lepiota</i> aff. <i>haemorrhagica</i> "red stainer"	S	S						1
185	<i>Lepiota cristata</i>	S	S						1
264	<i>Lepiota</i> sp. "cream-grey"	S	S	1				1	
246	<i>Lepiota</i> sp. "purple-grey"	S	S						1
166	<i>Lepiota</i> sp. "creamy-brown"	S	S	4		6	2	12	10
76	<i>Lepiota</i> sp. "orange with brick red scales/white gills"	S	S		2		3	5	5
117	<i>Lepista</i> sp.	M?	S						

Sp #	Species	Life Mode	Substrate	Treatments				Well. East	Donnelly
				E.C.	C.B.	SW.	G.R.	Total	Total
214	<i>Leucapaxillus lilacinus</i>	M	S						
24	<i>Lycoperdon</i> sp.	S	S	1	4	6	11	22	5
190	<i>Macrolepiota konradii</i>	S	S						1
318	<i>Marasmellus</i> sp. "small white, on twigs & leaves"	S	L/T						2
191	<i>Marasmiellus</i> sp. "white umbrella"	S	T/W						1
239	<i>Marasmiellus</i> sp. "on zamia"	S	T						
55	<i>Marasmius crinis-equi</i>	S	L				1	1	1183
183	<i>Marasmius elegans</i>	S	S						
341	<i>Marasmius</i> sp. "tiny red on twigs"	S	L/T						26
309	<i>Marasmius</i> sp. (see 223 Fire fungi)	S	S/L						
75	<i>Marasmius</i> sp. "large brown, on Zamia stems"	S	T						
529	<i>Melanoleuca</i> sp. "grey-brown/white/white"	S	S				3	3	
22	<i>Melanotus hepatocrous</i> (<i>Crepidotus subhaustellaris</i>)	S	W						
373	<i>Merulius</i> sp "creamy yellow, on jarrah stick"	S	W						
101	<i>Merulius</i> sp. "pink-buff"	S	W/Bark		2			2	2
238	<i>Mycana yuulongicola</i>	S	W	39	28		6	73	23
503	Mycelium "creamy-grey crust on charcoal and leaves"	S	L/W	13		7	3	23	
504	Mycelium "creamy-orange mycelium under well rotted litter"	S	L/W	39	1	20	59	119	
44	<i>Mycena</i> aff. <i>atrata</i>	S	W	1		1	50	52	43
372	<i>Mycena</i> aff. <i>fumosa</i>	S	W						
134	<i>Mycena albidocapillaris</i> (aff. <i>subcapillaris</i>)	S	L			25	6	31	24
80	<i>Mycena carmeliana</i>	S	W	10	15		48	73	63
327	<i>Mycena maldea</i> (<i>austrocapillaris</i> (bleach))	S	L		7			7	8
50	<i>Mycena mijoii</i>	S	L	22	1	99		122	1269
66	<i>Mycena pura</i>	S	S/L	41				41	6
144	<i>Mycena sanguinolenta</i>	S	S	20	310	35	10	375	17
491	<i>Mycena</i> sp. "brown pointy cap"	S	L	156				156	
521	<i>Mycena</i> sp. "brown pura"	S	L	15				15	
523	<i>Mycena</i> sp. "brown striate with dark umbo"	S	L	29				29	
456	<i>Mycena</i> sp. "brown-grey, viscid conic"	S	L		5			5	10
51	<i>Mycena</i> sp. "buff umbrella"	S	L/T	637	43	47	19	746	276
336	<i>Mycena</i> sp. "dk brown on burnt ground"	S	S						
285	<i>Mycena</i> sp. "light brown striate/white stems, on wood"	S	W				12	12	
27	<i>Mycena</i> sp. "long stem"	S	W						
376	<i>Mycena</i> sp. "small brown with decurrent gills"	S	W						2
326	<i>Mycena</i> sp. "small buff on wood - bleach"	S	W						
352	<i>Mycena</i> sp. "small creamy yellow-white"	S	L/Bark						
165	<i>Mycena</i> sp. "small grey - bleach"	S	S/L	1	1			2	50
502	<i>Mycena</i> sp. "striate cap, decurrent gills, on burnt ground"	S	S			3	4	7	
477	<i>Mycena</i> sp. "tiny rosy pink"	S	S						1

Sp #	Species	Life Mode	Sub-strate	Treatments				Well. East	Donnelly
				E.C.	C.B.	SW.	G.R.	Total	Total
88	<i>Mycena</i> sp. "tiny white with decurrent gills"	S	S						19
64	<i>Mycena</i> sp. "tiny white, on twigs"	S	T				3	3	46
308	<i>Mycena</i> sp. "grey-brown,/no bleach"	S	S	3	1	1	8	13	5
302	<i>Mycena</i> sp. "nipple umbrellas"	S	W						
312	<i>Mycena</i> sp. "pink,bleach"	S	S/L						
295	<i>Mycena</i> sp. "small buff"	S	L	6		5	35	46	3
182	<i>Mycena</i> spp. (unidentified)	S		3		8		11	
163/260	<i>Mycena subgallericulata</i>	S	W				13	13	19
510	<i>Mycoacia subceracea</i>	S	T/W						1
164	<i>Nidula candida</i>	S	L/T				19	19	
413	<i>Nidularia</i> aff. <i>farcta</i> "white dots on roo poo"	S	Dung		8	21		29	12
127	<i>Omphalina</i> aff. <i>umbellifera</i>	S	S			3	71	74	52
112	<i>Omphalina chromacea</i>	S	S						39
461	<i>Omphalina</i> sp.	S	S						8
213	<i>Omphalotus nidiformis</i>	S	W						
130	Orange parasite on white resupinate polypore (sp.116)	P	Fungus						
104	<i>Panellus ligulatus</i>	S	W						14
339	<i>Panellus</i> sp. - on ground	S	S						
343	<i>Panellus</i> sp. "soft brown"	S	W						
311	<i>Panus fasciatus</i>	S	W			4	5	9	89
356	<i>Paxillus</i> sp. "robust with bulbous base"	M	S						
522	<i>Paxillus</i> sp. "yellow" (<i>P. infundibuliformis</i> ?)	M	S	1		7	5	13	
179	<i>Paxillus</i> sp. "yellow, brown scales"	M	S		1			1	
332	<i>Peziza "praetervisa"</i>	S	S	73		36	134	243	
524	<i>Peziza</i> aff. <i>thozetii</i>	S	S	1				1	
501	<i>Peziza</i> sp. "black, flat"	S	S				2	2	
455	<i>Peziza</i> sp. "brown"	S	S						2
527	<i>Peziza</i> sp. "dark brown, smooth"	S	S				2	2	
499	<i>Peziza</i> sp. "dark brown-burgandy, tan underside"	S	S	8			31	39	
256	<i>Peziza</i> sp. "white cup"	?	S						
330	<i>Peziza tenacella</i>	S	S	34		22	31	87	
300	<i>Peziza whitei</i>	M	S	1			5	6	1
488	<i>Phaeocollybia raticauda</i>	S	W/S				14	14	
479	<i>Phelledon</i> sp. "niger brown"	S	L			12		12	154
136	<i>Phellinus gilvus</i>	S	W						20
37	<i>Phellinus</i> sp. "yellow rim"	S	W						
70	<i>Phellodon</i> aff. <i>niger</i>	S	L/S						153
160	<i>Pholiota highlandensis</i>	S	S	2	32	49	61	144	16
119	<i>Pholiota multicingulata</i>	S	W						2
506	<i>Pholiota</i> sp. "water soaked gills"	S	S						2
481	<i>Phylloporus</i> sp.	M	S		1			1	
363	<i>Piptoporus australiensis</i>	S	W						1
403	<i>Pisolithus</i> aff. <i>arhizus</i> "black-yellow"	M	S	1				1	
353	<i>Pisolithus</i> sp. 'small, stalked"	M	S						
192	<i>Plectania</i> sp. "black"	S	L						
133	<i>Pluteus attromarginata</i>	S	W						1
248	<i>Pluteus cervinus</i>	S	W						1

Sp #	Species	Life Mode	Sub- strate	Treatments				Well. East	Donn- elly
				E.C.	C.B.	SW.	G.R.	Total	Total
47	<i>Pluteus lutescens</i> "orange"	S	W						6
47b	<i>Pluteus lutescens</i> "yellow-green"	S	W						2
4	<i>Pluteus</i> sp. "brown velvet"	S	S				1	1	1
157	<i>Podoserpula pusio</i>	S/M?	L/S						18
277	Polypore "beige"	S	W	1				1	
13	Polypore "brown with white margin"	S	W						
3	Polypore "long white shelf"	S	W						
333	Polypore "on dead waterbush"	S	W	1				1	
474	Polypore "soft pored bracket"	S	W						9
361	Polypore "white resupinate on twig"	S	W/T						
116	Polypore "white resupinate"	S	T/W			1	1	2	6
313	<i>Polyporus citreus</i>	S	W/T						
494	<i>Polyporus</i> sp. "brown stalked"	S	W			1		1	
109	<i>Poria</i> sp. "purple splash"	S	W						
145	<i>Poronia ericii</i>	C	Dung				8	8	99
236/219	<i>Postia (Tyromyces) peliculosa</i>	S	W				3	3	1
155	<i>Protuberia canescens</i>	M?	S			1	2	3	
17	<i>Psathyrella</i> sp.	S	S/L						
229	<i>Psathyrella</i> sp.	S	L	3				3	6
250	<i>Psathyrella</i> sp.	S	L						50
337	<i>Psathyrella</i> sp.	S	S						1
359	<i>Psathyrella</i> sp. "brown with white skirt"	S	S						1
177	<i>Psilocybe coprophila</i>	C	Dung	23	83	19	34	159	16
331	<i>Pulvinula archerii</i>	S	S	95	5		118	218	23
280	<i>Pulvinula</i> sp.	S	S						
129	<i>Pulvinula</i> sp. (<i>P. tetraspora</i> ?)	S	S				15	15	1
176	<i>Pycnoporus coccineus</i>	S	W	2		49	25	76	12
366	<i>Pyronemas</i> sp.	S	S		1			1	
351	<i>Ramaria</i> / <i>Clavulina</i> "creamy white"	M	S						
52	<i>Ramaria</i> aff. <i>aurea</i> "yellow, flat tops"	M	S						4
72	<i>Ramaria holorubella</i> "purple-pink with pink tips"	M	S				1	1	17
102	<i>Ramaria ochroceosalmonicolor</i>	M	S	3	35	25	27	90	30
139	<i>Ramaria</i> sp. "bright-yellow"	M	S						
242	<i>Ramaria</i> sp. "cream/flat"	M	S		1			1	1
247	<i>Ramaria</i> sp. "lemon yellow"	M	S	5		5	8	18	
86	<i>Ramaria</i> sp. "orange-red, yellow stem"	M	S	2			1	3	3
367	<i>Ramaria</i> sp. "small yellow"	M?	S						
490	<i>Ramaria</i> sp. "tan, Allocasuarina litter"	S	S	291				291	
254	<i>Ramaria vesatilis</i> "purple"	M	S						
79	<i>Resupinatus cineroscens</i>	S	T/Bark		5			5	66
452	<i>Rhizopogon</i> sp.	M	S						2
397	<i>Rhodocybe</i> ? sp. "grey agaric"	S	S			1		1	
209	<i>Rickenella fibula</i>	S	Moss						31
69	<i>Russula adusta</i>	M	S						10
90	<i>Russula</i> aff. <i>cyanoxantha</i>	M	S	1				1	5
89	<i>Russula clelandii</i> group	M	S	12	2	3	1	18	40
202	<i>Russula flocktoniae</i>	M	S	53	2	18	6	79	4
92	<i>Russula neerimea</i>	M	S	2				2	22
178	<i>Russula persanguinea</i> (white stem)	M	W						1
107	<i>Russula</i> sp. "grey-white"	M	S		1			1	2

Sp #	Species	Life Mode	Sub-strate	Treatments				Well. East	Donnelly
				E.C.	C.B.	SW.	G.R.	Total	Total
10a	<i>Russula</i> sp. "small white-white-white"								5
10	<i>Russula</i> sp. "white/white/white"	S	S	18	5	9	3	35	72
276	<i>Russula</i> sp. "purple-mottled"	M	S						
342	<i>Ryvardinia campyla</i>	S	W						1
263	<i>Sarcodon</i> sp. "brown"	S	S						4
315	<i>Scleroderma</i> sp. "yellow/yellow mycelium"	M	S						1
150	<i>Scutellina</i> aff. <i>margaritacea</i>	S	W/T						14
12	<i>Simocybe</i> sp. "olive"	S	W						3
306	<i>Sphaerobolus stellatus</i>	S	L				20	20	
132	<i>Steccherinum</i> sp. "creamy yellow crust"	S	W			1		1	
94	<i>Steccherinum</i> sp. "tiered white shelves"	S	W		5			5	30
106	<i>Stemonitis herbatica</i>		Fruits						
62	<i>Stereum hirsutum</i>	S	W	21		58	102	181	123
149/141/152	<i>Stereum illudens</i>	S	W		5	140	83	228	185
5&84	<i>Stereum</i> sp. "grey-brown white hirsute, purple fertile layer"	S	W						
325	<i>Stereum</i> sp. "purple margin - algae"	S	W						15
16	<i>Stereum</i> sp. "translucent funnels"	S	S/Moss				67	67	16
67	<i>Stropharia semiglobata</i>	C	Dung	2	5			7	11
451	<i>Stropharia</i> sp. "shaggy stem, on roo poo"	S	Dung						1
507	<i>Tapinella curtisii</i>	S	W						6
266	<i>Thelephora</i> sp. "white with orange margin"	M	S/Host	1	35	5	10	51	40
454	<i>Thelephora</i> "creamy jagged-ridged crust"	S	S				2	2	7
482	<i>Torrendia</i> sp.	M	S		2	33	9	44	
63	<i>Trametes versicolor</i> (brown or grey)	S	W			1	21	22	152
60	<i>Tremella mesentericia</i>	S	W						16
287	<i>Tremella</i> sp. "cloudy yellowish white"	S	W			2		2	23
289	<i>Tremella</i> sp. "tiny yellow knobs"	S	W			112	300	412	30
288	<i>Tremella</i> sp. "yellow buttons"	S	W			81		81	
161	<i>Tricholoma</i> aff. <i>virgatum</i>	M	S						1
483	<i>Tricholoma</i> sp.	S	S		2			2	
211	<i>Tricholoma</i> sp.	M	S						
446	<i>Tricholoma</i> sp. "orange with ring"	S	S		3			3	1
495	<i>Tricholoma</i> sp. "orange with white stem"	S	S			4		4	
54	<i>Tricoloma eucalypticum</i>	M	S	4	6	1	15	26	15
322	Truffle "black gleba"	M	S						
464	Truffle "pale yellow"	M	S						3
321	Truffle "pink gleba"	M	S		1			1	
258	Truffle "sticky"	M	S						
189	<i>Tubaria rufofulva</i>	S	W	9				9	1
305	<i>Tylopilus</i> sp. "yellow"	M	S				2	2	6
505	<i>Tyromyces caesius</i>	S	W						2
440	White mycelium on roo poo	S	Dung			1		1	
2	<i>Xerula australis</i>	S	S						
175	<i>Xylaria hypoxylon</i>	S	W						2
459	<i>Xylaria</i> sp. "black and white spears"	S	W/S						
528	Yellow discs on Emu poo	S	Dung	150				150	

Sp #	Species	Life Mode	Sub- strate	Treatments				Well. East	Donn- elly
				E.C.	C.B.	S.W.	G.R.	Total	Total
482	Number of species			110	95	108	137	240	230
	Number of fruitbodies			3755	1979	2632	3662	12053	15324

¹ S = saprotrophic, M = mycorrhizal, P = parasitic, C = coprophilous

² S = soil, L = litter, T = twigs, W = wood

³ E.C. = external control, C.B. = coupe buffer, S.W. = shelterwood, G.R. = gap release

CRYPTOGAMS

Ray Cranfield

Introduction

Cryptogam is the collective name for the lichen, moss and liverwort flora. Some species of lichens are important indicators of ecosystem health because they are sensitive to changes and disturbance in the environment in which they grow. The majority of cryptogams are resurrection species and although fully desiccated in the drier months, they rapidly re-hydrate with any opportunistic rain event. Many colonize primary substrates such as rocks and bare organic matter and are active in the initial breakdown of these materials. Mosses also play an important role in the stabilization of bare soil. Cryptogams are a major component of the biodiversity of forest ecosystems, and most species in Western Australia have unknown distributions and many are yet to be determined.

The object of this component of FORESTCHECK is to:

- Record species richness and abundance in each FORESTCHECK grid and treatment
- Record species habitat and substrate preference, and
- Monitor the effect of disturbance on cryptogam communities

Monitoring

Usually cryptogam monitoring is carried out in the autumn, but in 2004-05, monitoring was conducted in October (spring) 2004. At each of the 10 grids, the presence and frequency of each species along with the macro and microhabitat that each species occurred on was recorded.

Voucher Specimen Processing

All specimens collected this year have been identified or phrase-named for ease of redetermination. Information gained from several ongoing cryptogam studies enabled the current species list to be reviewed. This report reflects these changes. Several samples of terrestrial algae were also collected and prepared for incorporation in the state collection held at the WA Herbarium.

All collections were prepared for database entry and label generation prior to submitting vouchers to the Herbarium. A total of 413 collections (369 lichens, 39 mosses and 5 liverworts) were made from the 10 grids. The collections represent 162 species of lichens, 13 species of moss and 4 species of liverworts, making a total of 179 species of cryptogams. A further 6 collections of terrestrial algae were collected and 7 fungal collections. It should be noted that monitoring in spring rather than autumn resulted in a reduction in microscopic fungi collections but no real impact upon the other cryptogams normally sampled.

Results and Discussion

Species and habitats

Although the monitoring was conducted in spring it appears that the flora was well represented. A total of 179 species of cryptogams were recorded on the Wellington East grids (Appendix 1). However, the number of liverworts species recorded was very low. It may be that they have a naturally low occurrence in the drier eastern jarrah forest. Liverworts were located outside the grids, but appeared to be restricted to the sheltered banks of creek lines. Harvesting (Shelterwood and gap formation) also results in increased

exposure to the drying elements such as sun and wind. How such disturbances affect liverworts and other sensitive cryptogams is not yet known.

In general, a decline in species numbers occurred from external control to Shelterwood to gap release treatment (Fig. 1). In the external controls the long unburnt Nalyerin grid (FC28) had the highest number of species. In the treatments (shelterwood and gap release) there appeared to be no pattern or relationship associated with time since logging or burning. For example, the Godfrey grids had higher numbers and the Stockyard grids had lower numbers of species than would be expected from having respectively been burnt 2 and 6 years previously (compared to the controls). The unusual relationship between the treatments and time since treatment may also be due to species composition on the respective sites, which will be investigated in detail during the full analysis in 2006.

Wood (old logs), the bark of older trees and stone were the most utilised substrates in the control grids (Fig. 2). Generally, all 7 specified substrate types were present in each of the treatment grids but differences in colonization of the substrates were related to the age of the substrate. For cryptogams, time after fire is important and species substitution can be observed when primary species are replaced over time by subsequent species. On the gap release sites, although suitable substrates were present they were not always colonised because they were not yet mature enough to be colonised. On the other hand, the colonization of termite mounds was recorded on the majority of treatment grids but on the external controls the mounds were not colonized.

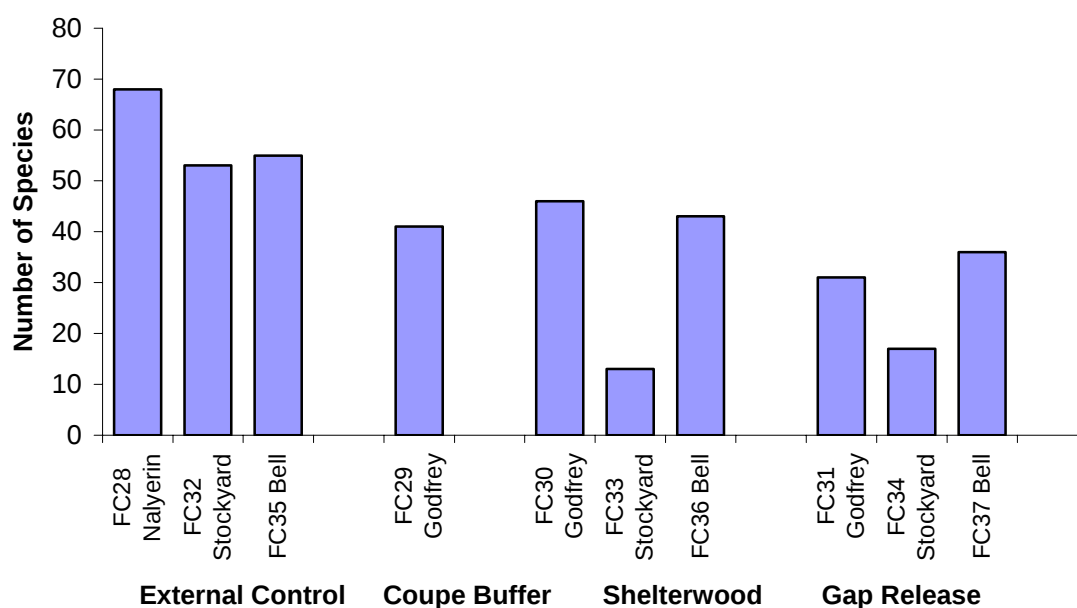


Figure 1. The number of species of cryptogams on each FORESTCHECK grid in Wellington East.

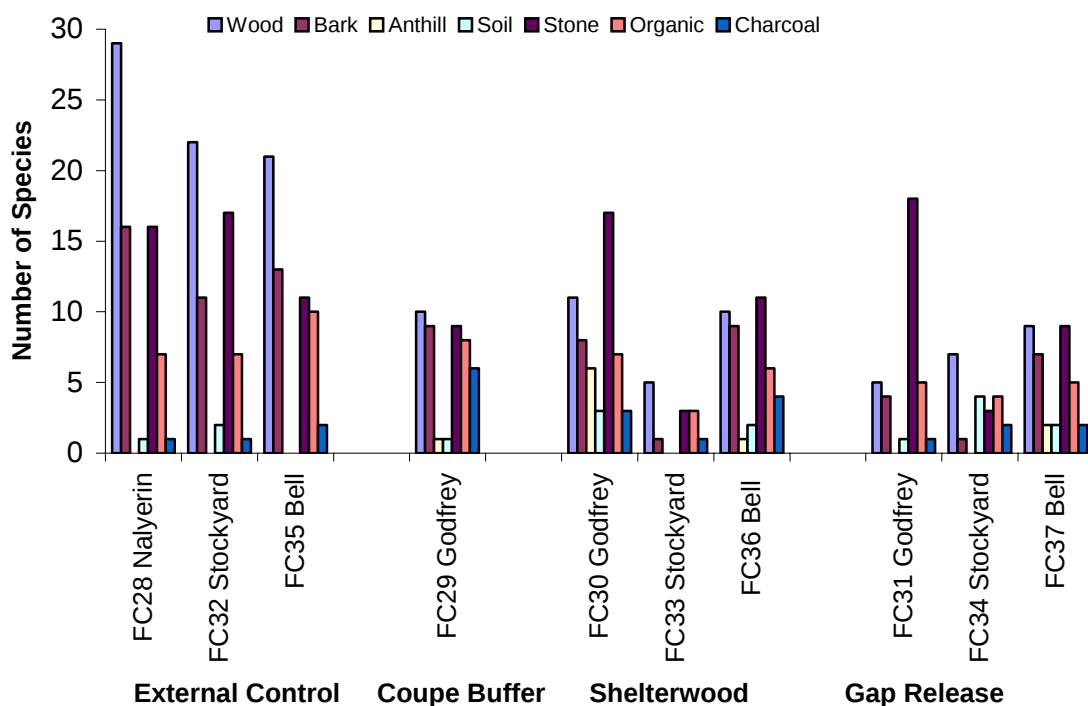


Figure 2. The number of cryptogam species colonizing the available microhabitats (substrates) on FORESTCHECK grids of Wellington East.

The position each species within strata was also recorded (Fig. 3). The number of species recorded at ground level (0-30 cm) was similar across all grids. At shrub level (31 cm – 3 m) the numbers of species in the grids appeared to be related to the type of treatment, with lower numbers of species being recorded in the Shelterwood and Gap release treatments respectively. This may be related to the degree of exposure and the age and structure of the regrowth. At tree level (>3.1 m) there were fewer species and they were recorded in lower numbers in the treatment (Shelterwood and gap release) grids than in the controls. This is possibly related to the age of the regrowth trees and to the number of older trees retained in the Shelterwood treatments. However, the accuracy of accessing the tree (>3.1 m) species is variable, as it depends on sampling material that occurs on fallen limbs, which may restrict the number of species actually recorded. A more thorough survey of canopy species should be considered where tree removal has occurred, as they may be a high-risk group.

An understanding of the growth requirements of individual cryptogam species and the effects of modified external influences is required to ensure the long term viability and diversity of these taxa.

The number of species of lichens, mosses and liverworts occurring on each substrate and in each level of strata is included in the summary in Appendix 1.

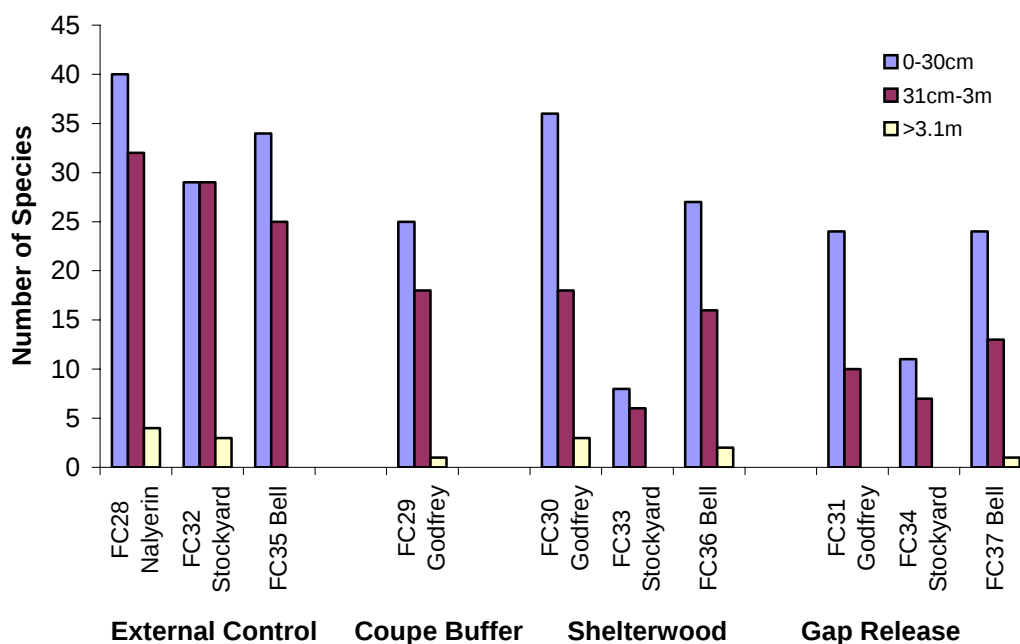


Figure 3. The number of cryptogam species recorded at each stratum on FORESTCHECK grids of Wellington East.

Indicator Species

At Wellington East investigation into the use of possible indicator species continued. The list of potential indicators includes 28 species, of which 21 are lichens, 6 are mosses and 2 are liverworts (see 2002-03 Wellington report). The species were chosen on the basis of their preferred substrate and the position they occupy in the strata.

Several species on the original list have undergone taxonomic review and as a result have been subject to name changes, these changes are detailed in Table 1. An additional species that colonises old decaying wood, *Graphis* sp. (black rays), was added to the list in 2003-04, this species has now been determined as an un-named species of *Opegrapha* not previously recorded for Australia.

At each grid the presence of each potential indicator species was noted and in most instances the list of species present was close to the possible twenty-eight species listed (Fig. 4). Preliminary examination of the indicator species indicates that species not being present on a grid represents a habitat that has been lost or modified. However, the possibility that a species may have a limited geographical range cannot as yet be ruled out. FORESTCHECK includes jarrah forest that extends over a large geographical range and as yet we do not have detailed information on the extent of the range of all the potential indicator species.

Table 1. Recent name changes (a) of potential FORESTCHECK Indicator Species and additional taxa (b) that were added to the list in 2003-4 (B = moss, H = liverwort and L = lichen).

Code No.	Group	Taxon	Redetermined Name
(a) <u>Redetermined Indicator Taxa</u>			
23	H	<i>Fossombronia</i> sp. (leafy & lettuce)	= <i>Fossombronia altilamellosa</i>
8	L	<i>Neuropogon</i> sp	= <i>Usnea</i> sp. (leuco)
10	L	<i>Parmotrema cooperi</i>	= <i>Rimelia reticulata</i>
18	B	<i>Sematophyllum contiguum</i>	= <i>Sematophyllum subhumile</i> var. <i>contiguum</i>
21	B	<i>Ceratodon purpureus</i>	= <i>Ceratodon purpureus</i> subsp. <i>convolutes</i>
28	L	<i>Graphis</i> sp. (blackrays)	= <i>Opegrapha</i> sp.

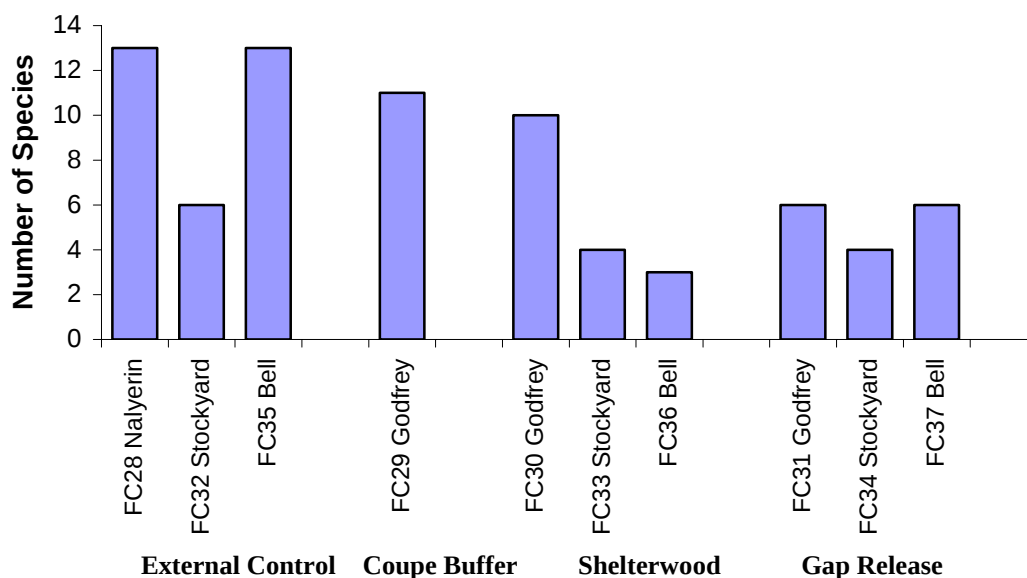


Figure 4. The number of indicator species (from a possible 28) recorded on each FORESTCHECK grid in Wellington East.

Other species that show up as possible indicators are included in Appendix 1 (bold type). In establishing this and previous lists an attempt has been made to include species that have specific requirements of either substrate or landscape position. It is anticipated that in the final report in 2006 that a list of indicator species can be presented along with the rationale as to why they are selected and what their absence may indicate.

Comparison of all FORESTCHECK Locations

An overview of all FORESTCHECK locations (Donnelly, Wellington, Perth Hills and Wellington East) was undertaken in an attempt to see possible similarities and differences between the 4 areas.

Common species

17 species of lichens, 3 species of moss and 2 species of liverworts were found to be common to 90% of the grids in the 4 FORESTCHECK locations. Many of these species appear on the proposed indicator species list. The common lichens were: *Cladia aggregata*, *Cladia schizopora*, *Cladonia krempelhuberi*, *Cladonia ramulosa*, *Cladonia rigida*, *Cladonia sulcata*, *Graphis* sp. (blackrays) [= *Opegrapha* sp.], *Hypocenomyce foveata*, *Hypocenomyce scalaris*, *Hypogymnia subphysodes* var. *subphysodes*, *Ochrolechia* sp. (G. Kantavilis 306/92), *Pannoparmelia wilsonii*, *Parapropidia glauca*, *Ramboldia stuartii*, *Tephromela atra*, *Thysanothecium scutellatum* and *Usnea inermis*. The common mosses were: *Barbula calycina*, *Campylopus introflexus* and *Sematophyllum subhumile* var. *contiguum* and the common liverworts: *Cephaloziella exiliflora* and *Chiloscyphus semiteres*.

External control and gap release grids

Species richness

There was some variation observed in the number of species recorded on the control grids at each location and between locations (Fig. 5). As the preferred habitat substrates for most species are represented on most grids, the variation may be the result of varying climatic conditions and vegetation densities on each grid and between locations. The lower number of liverwort species recorded at Wellington, Wellington East and the Perth Hills for example may be indicative of the warmer drier conditions on these grids.

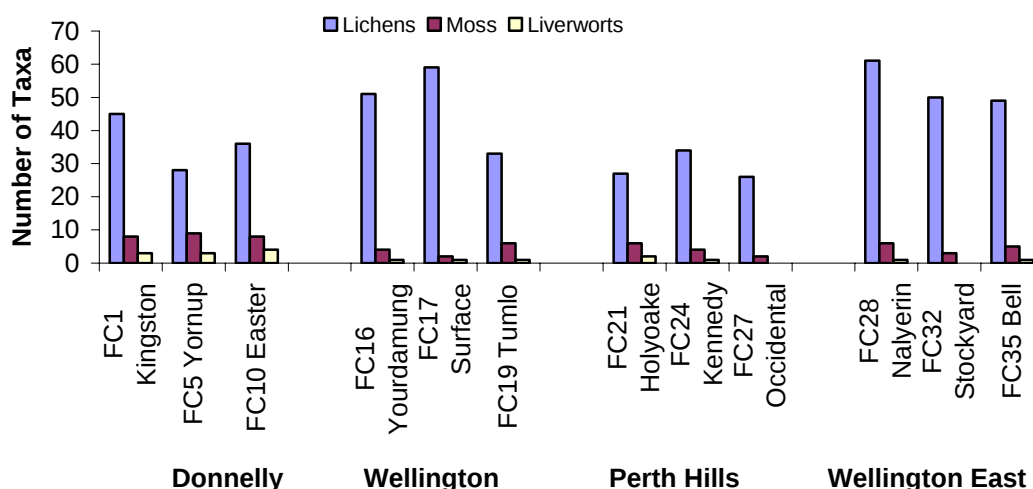


Figure 5. The number of cryptogam taxa recorded on external control sites at the Donnelly, Wellington, Perth Hills and Wellington East FORESTCHECK locations.

The lower number of taxa recorded on the gap release sites (Fig. 6) is a direct result of disturbance and habitat loss, or by either tree harvesting or fire or both. The loss of micro climatic niches is known to impact on several cryptogam species resulting in the loss of species or reduced population sizes. The time since treatment on the gap release sites is important in the recovery of the cryptogam flora. On disturbed sites recovery is influenced directly by the increase in size and density of understorey vascular flora and by tree species densities. Early strong regrowth of tree saplings can retard the establishment of many cryptogams as a direct result of too much shading and lack of aged substrates. Further management practices, such as thinning, may have the effect of either delaying or facilitating the recovery of the cryptogam flora on these sites and further investigation is needed in order to understand what impact this intervention would imply.

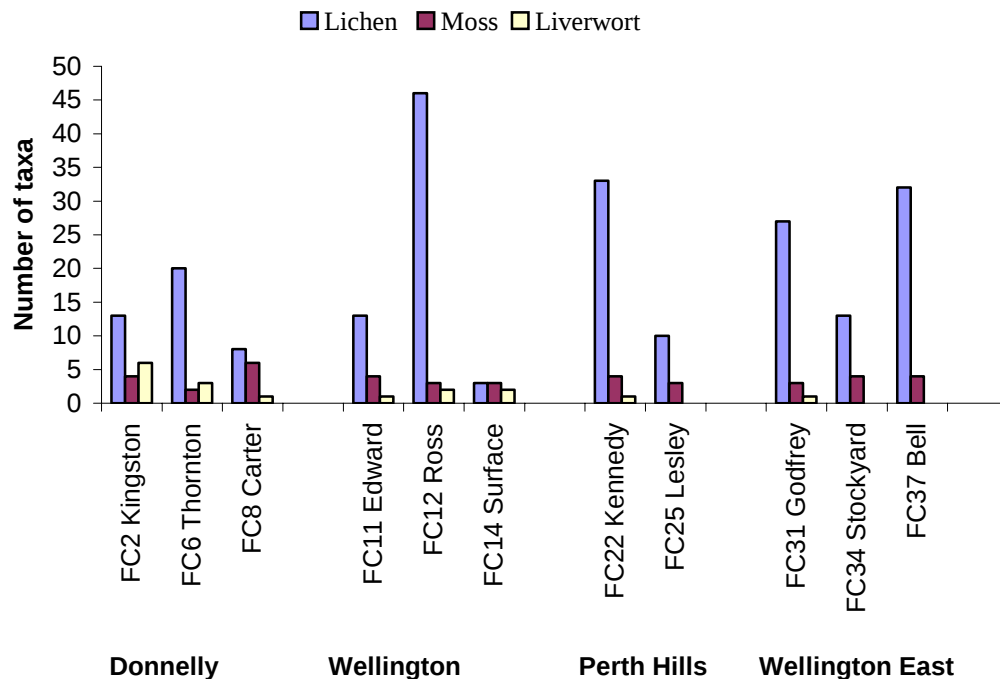


Figure 6. The number of cryptogam taxa recorded on gap release sites at the Donnelly, Wellington, Perth Hills and Wellington East FORESTCHECK locations.

Strata occupied

The use of particular strata by various cryptogam flora in the external control grids varied between each location. At Donnelly, Perth Hills and Wellington East the majority of species were generally recorded on the ground layer (0-30 cm) while at Wellington it was the shrub layer (30 cm- 3 m) stratum was colonized by the majority of species (Fig. 7). The number of species recorded in the tree layer (>3 m) was low at all 4 locations but noticeably lower in the Perth Hills. This variation may be a reflection of the change in moisture and temperature from north to south in the locations.

On the gap release sites (Fig. 8), the loss of older trees, and the mature shrub cover, has reduced the availability of suitable substrates for cryptogam establishment. Rapid tree re-growth, dense canopy cover, and the removal of debris by fire in the early stages appear to restrict the establishment of ground and shrub layer occupation. On the wetter sites, however, the ground level appears to be the desired location for cryptogams but it is really a reflection of the predominance of moss species and liverworts.

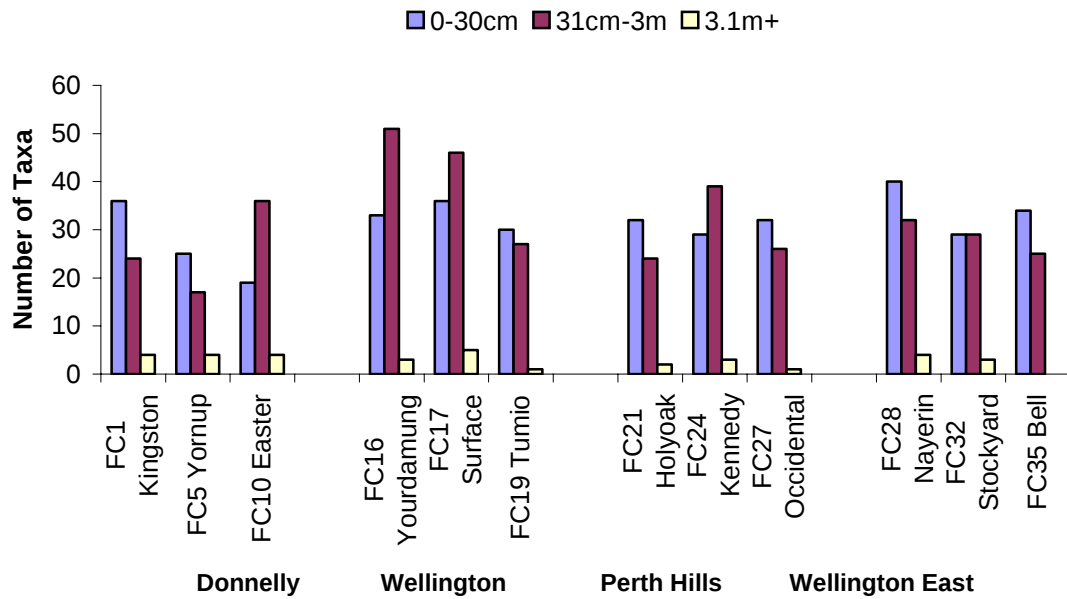


Figure 7. The number of cryptogam taxa occupying the different levels of forest strata in the control grids at the Donnelly, Wellington, Perth Hills and Wellington East FORESTCHECK locations.

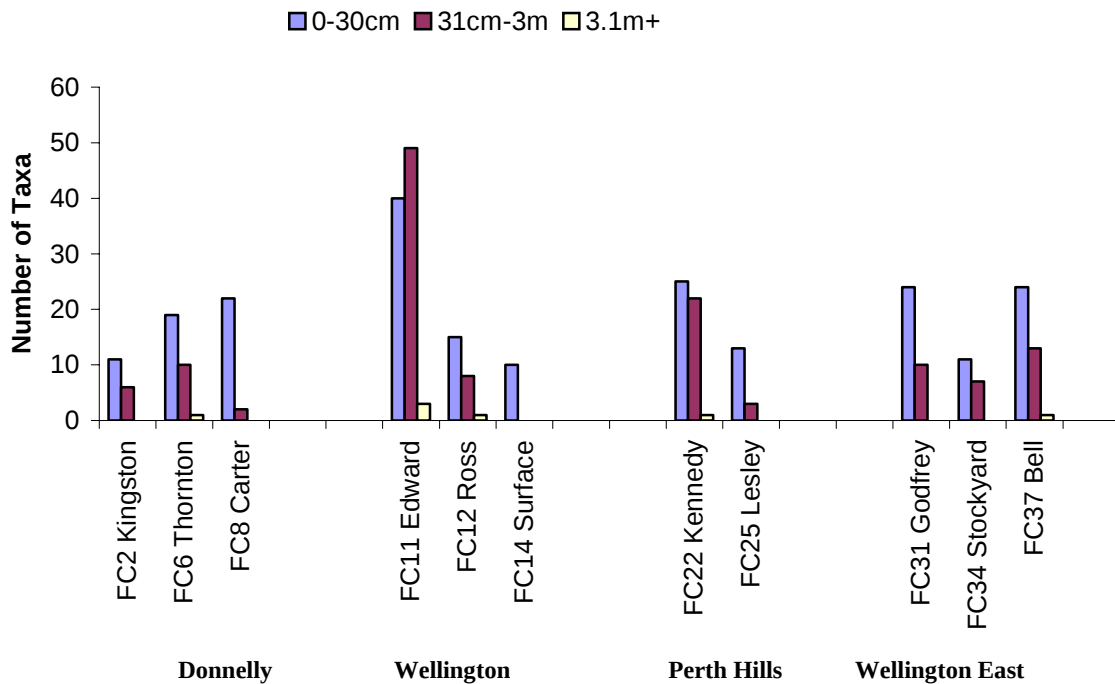


Figure 8. The number of cryptogam taxa occupying the different levels of forest strata in the gap release grids at the Donnelly, Wellington, Perth Hills and Wellington East FORESTCHECK locations.

Conclusions

- Cryptogam species richness was higher in external control grids
- Anthills, a common substrate for several species of lichens and mosses, were not utilized in the external controls – reason not clear
- The most utilized substrates in all treatments were rocks and decaying logs

- Only one species of liverwort was recorded, likely due to the drier environment of the eastern jarrah forest

Comparing Locations

- 17 species of lichens, 3 species of moss and 2 species of liverworts were found to be common to 90% of the grids in the 4 FORESTCHECK locations
- External controls are consistently higher in species richness
- The lower number of taxa recorded on the gap release sites appears to be due to disturbance and habitat loss
- Donnelly location had a higher number of moss and liverwort species, this is due to the wetter environment

Appendix 1. Presence/absence and summary of Cryptogam taxa located on each FORESTCHECK grid (Names in **bold text** are the nominated indicator species).

Taxa	External Control				Coupe Buffer	Shelterwood		Gap Release		
	FC28 Nalyerin	FC32 Stockyard	FC35 Bell	FC29 Godfrey		FC30 Godfrey	FC33 Stockyard	FC36 Bell	FC31 Godfrey	FC34 Stockyard
Mosses (B)										
<i>Barbula calycina</i>		✓		✓	✓	✓	✓		✓	✓
<i>Campylopus introflexus</i>	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓
<i>Ceratodon purpureus</i> ssp. <i>convolutus</i>	✓							✓	✓	
<i>Didymodon torquatus</i>							✓			✓
<i>Funaria hygrometrica</i>			✓		✓			✓		
<i>Rhynchostegium subhumile</i> var. <i>contiguum</i>	✓	✓	✓	✓						
<i>Rhynchostegium tenuifolium</i> var. <i>tenuifolium</i>							✓			
<i>Rosulabryum billarderi</i>	✓		✓						✓	
<i>Rosulabryum</i> sp. Stockyard (R.J. Cranfield 20799)						✓				
<i>Weissia controversa</i>							✓			✓
Liverworts (H)										
<i>Cephaloziella exiliflora</i>	✓		✓	✓	✓			✓		
Lichens (L)										
<i>Acarospora</i> sp. brown (R.J. Cranfield 20633)					✓					
<i>Biatropsis usneanum</i>			✓							
<i>Biatropsis</i> sp. Bell (R.J. Cranfield 20994)							✓			
<i>Buellia stellulata</i>	✓	✓	✓		✓	✓	✓	✓		✓
<i>Buellia</i> sp.					✓		✓			
<i>Buellia</i> sp. brown apo (R.J. Cranfield 20714b)				✓						
<i>Buellia</i> sp. sparse (R.J. Cranfield 20619)					✓					
<i>Calicium abietinum</i>	✓	✓								
<i>Calicium glaucellum</i>		✓	✓					✓		
<i>Calicium victorianum</i> subsp. <i>desidiosum</i>		✓								
<i>Calicium victorianum</i> subsp. <i>victorianum</i>	✓	✓		✓					✓	
<i>Calicium tricolor</i>	✓			✓						
<i>Caloplaca marina</i>	✓	✓	✓				✓	✓		✓
<i>Chaenotheca brunneola</i>					✓					
<i>Chaenothecopsis pusilla</i>	✓									
<i>Cladia aggregata</i>	✓		✓	✓	✓					
<i>Cladia schizopora</i>	✓		✓	✓	✓			✓		
<i>Cladonia angustata</i>				✓	✓					
<i>Cladonia capitellata</i>					✓					
<i>Cladonia cervicornis</i> var. <i>verticillata</i>			✓							
<i>Cladonia crispata</i> var. <i>cetrariiiformis</i>	✓	✓							✓	
<i>Cladonia humilis</i> var. <i>humilis</i>							✓			
<i>Cladonia krempelhuberi</i>	✓	✓		✓	✓		✓	✓		✓
<i>Cladonia macilenta</i>	✓			✓						
<i>Cladonia polycarpoides</i>			✓							
<i>Cladonia praetermissa</i>				✓						
<i>Cladonia ramulosa</i>							✓		✓	
<i>Cladonia rigida</i>	✓	✓	✓	✓	✓		✓			✓
<i>Cladonia scabriuscula</i>			✓				✓			

Taxa	External Control			Coupe Buffer	Shelterwood			Gap Release		
	FC28 Nalyerin	FC32 Stockyard	FC35 Bell		FC29 Godfrey	FC30 Godfrey	FC33 Stockyard	FC36 Bell	FC31 Godfrey	FC34 Stockyard
<i>Cladonia southlandica</i>	✓									✓
<i>Cladonia sulcata</i>						✓		✓		
<i>Cladonia ustulata</i>	✓									✓
<i>Cladonia</i> sp. Nalyerin (R.J. Cranfield 20895)	✓									
<i>Cladonia</i> sp. Stockyard (R.J. Cranfield 20788)		✓								
<i>Cladonia</i> sp. Stockyard (R.J. Cranfield 20737)										✓
<i>Cyphelium trachylioides</i>			✓					✓		
<i>Diploschistes ocellatus</i>										✓
<i>Diploschistes scruposus</i>	✓									
<i>Diploschistes strictus</i>		✓	✓			✓			✓	✓
<i>Diploschistes</i> sp.				✓					✓	
<i>Diploschistes</i> sp.lead (R.J. Cranfield 20796)							✓			
<i>Ephebe</i> sp.						✓				
<i>Flavoparmelia springtonensis</i>		✓								
<i>Graphis</i> sp. (black beans)		✓		✓					✓	
<i>Graphis</i> sp. (black buns)										
Graphis sp. (black rays)	✓		✓				✓			
<i>Graphis</i> sp. (tram lines)	✓	✓						✓		✓
<i>Graphis</i> sp. (writhing mass)				✓						
<i>Hafellia tetrapla</i>			✓							
<i>Hypocenomyce australis</i>	✓	✓	✓						✓	✓
<i>Hypocenomyce foveata</i>	✓	✓	✓	✓			✓			✓
<i>Hypocenomyce scalaris</i>	✓	✓	✓	✓	✓		✓			✓
Hypogymnia subphysodes var. <i>subphysodes</i>	✓		✓	✓				✓		
<i>Imshaugia aleurites</i>										✓
<i>Lecanora farinacea</i>										✓
<i>Lecidea orosthea</i>						✓				
<i>Lecidea</i> sp. Godfrey (R.J. Cranfield 20657)									✓	
<i>Lecidea</i> sp. grey smudge (R.J. Cranfield 20645)						✓				
<i>Lecidella stigmata</i>		✓	✓	✓						
<i>Megalaria</i> sp. Nalyerin (R.J. Cranfield 20906)	✓									
Menegazzia platytrema	✓	✓								
<i>Neofuscelia verrucella</i>		✓	✓	✓				✓		
<i>Ochrolechia</i> sp. (G.S. Kantavilas 306/92)	✓	✓	✓				✓	✓		✓
<i>Pannaria elixii</i>										✓
Pannoparmelia wilsonii	✓	✓	✓	✓	✓			✓		
Paraporphidia glauca	✓			✓	✓			✓		
<i>Parmelia erumpens</i>								✓		
<i>Parminopsis minarum</i>								✓		
<i>Parmotrema praesorediosum</i>	✓		✓	✓			✓		✓	
<i>Peltula euploca</i>										✓
<i>Pertusaria trimera</i>						✓				
<i>Pyrrhospora laeta</i>						✓				
<i>Ramalea cochleata</i>						✓				
<i>Ramboldia stuartii</i>	✓	✓	✓	✓	✓	✓	✓			✓
<i>Rhizocarpon geographicum</i>		✓								
<i>Rhizocarpon tinei</i>		✓	✓						✓	

Taxa	External Control			Coupe Buffer	Shelterwood			Gap Release		
	FC28 Nalyerin	FC32 Stockyard	FC35 Bell		FC29 Godfrey	FC30 Godfrey	FC33 Stockyard	FC36 Bell	FC31 Godfrey	FC34 Stockyard
<i>Rhizocarpon</i> sp. grey (R.J. Cranfield 20894)	✓									
<i>Rimelia reticulata</i>		✓	✓							
<i>Rinodina gennari</i>	✓	✓	✓							
<i>Tephromela atra</i>	✓									
<i>Thysanothecium hookeri</i>					✓		✓			
<i>Thysanothecium scutellatum</i>	✓	✓	✓	✓			✓			✓
<i>Thysanothecium</i> sp. green scales (R.J. Cranfield 20953)										✓
<i>Toninia sedifolia</i>		✓		✓						
<i>Toninia</i> sp.		✓								✓
<i>Trapelia</i> sp. Godfrey (R.J. Cranfield 20612)					✓					
<i>Trapelia</i> sp. Godfrey (R.J. Cranfield 20718)				✓						
<i>Trapelia</i> sp. Bell (R.J. Cranfield 20850)			✓							
<i>Trapeliopsis</i> sp. Godfrey (R.J. Cranfield 20649)					✓					
<i>Trapeliopsis</i> sp. Godfrey (R.J. Cranfield 20714)				✓						
<i>Trapeliopsis</i> sp. Bell (R.J. Cranfield 20853)			✓							
<i>Trapeliopsis</i> sp.	✓	✓		✓	✓					
<i>Usnea inermis</i>	✓	✓	✓	✓	✓	✓	✓		✓	✓
<i>Usnea subalpina</i>	✓	✓	✓							
<i>Usnea subciliata</i>							✓			
<i>Usnea</i> sp. (leuco)	✓				✓					
<i>Usnea</i> sp. Nalyerin (R.J. Cranfield 20883)	✓									
<i>Xanthoparmelia congensis</i>				✓						
<i>Xanthoparmelia flindersiana</i>								✓		
<i>Xanthoparmelia mexicana</i>	✓	✓		✓	✓	✓	✓	✓	✓	✓
<i>Xanthoparmelia nana</i>			✓		✓			✓		✓
<i>Xanthoparmelia parvoincerta</i>	✓									
<i>Xylographa</i> sp. Stockyard (R.J. Cranfield 20739)									✓	
Genus sp. balls (R.J. Cranfield 20676)								✓		
Genus sp. Bell (R.J. Cranfield 20855)			✓							
Genus sp. Bell (R.J. Cranfield 20946)										✓
Genus sp. black folds (R.J. Cranfield 20914)	✓									
Genus sp. black freckles (R.J. Cranfield 20747)		✓								
Genus sp (black hairy stepping stones)			✓					✓		✓
Genus sp. blobs (R.J. Cranfield 20982)								✓		
Genus sp. (blue apo)							✓			✓
Genus sp. bread crumbs (R.J. Cranfield 20889)	✓									
Genus sp. brown crust (R.J. Cranfield 21003)							✓			
Genus sp. brown wax (R.J. Cranfield 20854a)			✓							
Genus sp. coke (R.J. Cranfield 20633)					✓					
Genus sp. Godfrey (R.J. Cranfield 20633)					✓					
Genus sp. Godfrey (R.J. Cranfield 20640)					✓					
Genus sp. Godfrey (R.J. Cranfield 20704)				✓						
Genus sp. (green crust)		✓	✓					✓		
Genus sp. green wax (R.J. Cranfield 20714a)				✓						

Taxa	External Control			Coupe Buffer	Shelterwood			Gap Release		
	FC28 Nalyerin	FC32 Stockyard	FC35 Bell		FC29 Godfrey	FC30 Godfrey	FC33 Stockyard	FC36 Bell	FC31 Godfrey	FC34 Stockyard
Genus sp. grey (R.J. Cranfield 20915)	✓									
Genus sp. grey brain (R.J. Cranfield 20616)		✓			✓					
Genus sp. (grey corn cobs)	✓		✓		✓					
Genus sp. (grey crust)		✓	✓				✓			
Genus sp. grey scale (R.J. Cranfield 20741)		✓								
Genus sp. Stockyard (R.J. Cranfield 20803)						✓				
Genus sp. stromats (R.J. Cranfield 20639)					✓					
Genus sp. yellow crust (R.J. Cranfield 20922)	✓	✓	✓				✓			✓
Genus sp. yellow wax (R.J. Cranfield 20770)		✓	✓							
Genus sp. (U7)							✓			
Genus sp. RC20872	✓									
SUMMARY	External Control			Coupe Buffer	Shelterwood			Gap Release		
	FC28 Nalyerin	FC32 Stockyard	FC35 Bell		FC29 Godfrey	FC30 Godfrey	FC33 Stockyard	FC36 Bell	FC31 Godfrey	FC34 Stockyard
Number of voucher collections	68	53	55	41	46	13	43	31	17	36
GROUPS (number of species)										
Lichen (L)	61	50	49	36	43	10	38	27	13	32
Mosses (B)	6	3	5	4	2	3	5	3	4	4
Liverwort H)	1	0	1	1	1	0	0	1	0	0
Habitats										
(number of individual cryptogam records)										
Wood	29	22	21	10	11	5	10	5	7	9
Bark	16	11	13	9	8	1	9	4	1	7
Anthill	0	0	0	1	6	0	1	0	0	2
Soil	1	2	0	1	3	0	2	1	4	2
Stone	16	17	11	9	17	3	11	18	3	9
Organic Material	7	7	10	8	7	3	6	5	4	5
Charcoal	1	1	2	6	3	1	4	1	2	2
Stratal Position										
(number of individual cryptogam records)										
0-30 cm	40	29	34	25	36	8	27	24	11	24
31 cm-3 m	32	29	25	18	18	6	16	10	7	13
> 3.1 m	4	3	0	1	3	0	2	0	0	1
Indicator Species (from a total of 27)	13	6	13	11	10	4	3	6	4	6

VASCULAR PLANTS

Bruce Ward and Ray Cranfield

Introduction

The vegetation complexes of jarrah forest are considered to be relatively stable and resilient to natural disturbances such as fire. In most circumstances, and in time, the species that were present before the disturbance are generally present after the event, although abundances may change. Where logging is concerned, disturbance might include soil movement, mixing and compaction. This level of disturbance may impact more severely and cause a loss in diversity either through a loss in species richness or a shift in species abundance.

The object of this component of the FORESTCHECK program is to:

- Monitor vascular plant species richness and abundance on each of the FORESTCHECK grids
- Compare species richness, abundance and composition recorded on treatment grids (shelterwood and gap release) to those on unlogged grids (external control)

As the pyric succession of plants also affects which species are present, differences found may be successional rather than from logging. Therefore an allowance for time since fire needs to be considered in determining species richness.

This report highlights the results from the fourth round of monitoring for FORESTCHECK.

Monitoring

Spring is the preferred time to monitor understorey vascular plants as it coincides with the peak flowering time for most plants. Species richness and understorey vegetation structure were determined, by recording each species, estimating its area of cover, and measuring its position in the understorey strata, in 4 30 x 30 m plots in each grid (40 in total). Species density was measured by recording species occurrence and abundance in twenty 1 x 1 m plots in each grid (800 in total) and Vegetation structure is determined from levy contact data at various height categories in the understorey (Levy and Madden 1933). The area around each grid was used to search for voucher flowering plant specimens to aid in or confirm their identification.

Voucher Collections

A total of 127 vouchers were collected, representing 58% of the species recorded. All specimens have been processed and following entry onto the database will be lodged with the WA Herbarium (PERTH).

Results and Discussion

Species Richness

A total of 159 species were recorded in the 30 x 30 m plots and 137 species in the 1 x 1 m plots, 28 of which were unique and not found in the 30 x 30 m plots. The total number of species recorded was 187 (see Table 1 for comparison of species numbers and Appendix 1 for the complete species list). A further 36 species collected from outside the plot areas were unique and increased the species numbers on site to 223.

Table 1. The number of vascular plant species recorded in plots on all the FORESTCHECK grids at Donnelly, Wellington, Perth Hills and Wellington East.

Location	30 x 30 m Plots Species Numbers	1 x 1 m Plots Species Numbers	% Difference	Number of weed species
Donnelly 2002	203	158	22	24
Wellington 2003	181	116	36	10
Perth Hills 2004	188	126	33	4
Wellington East 2005	159	137	14	10

Species richness across treatments showed little variation and indicated that there has been no loss of species due to logging activities. Any loss in species numbers would be reflected in lower species richness recorded on the treatment grids (shelterwood and gap release) than was recorded for external controls (Fig. 1).

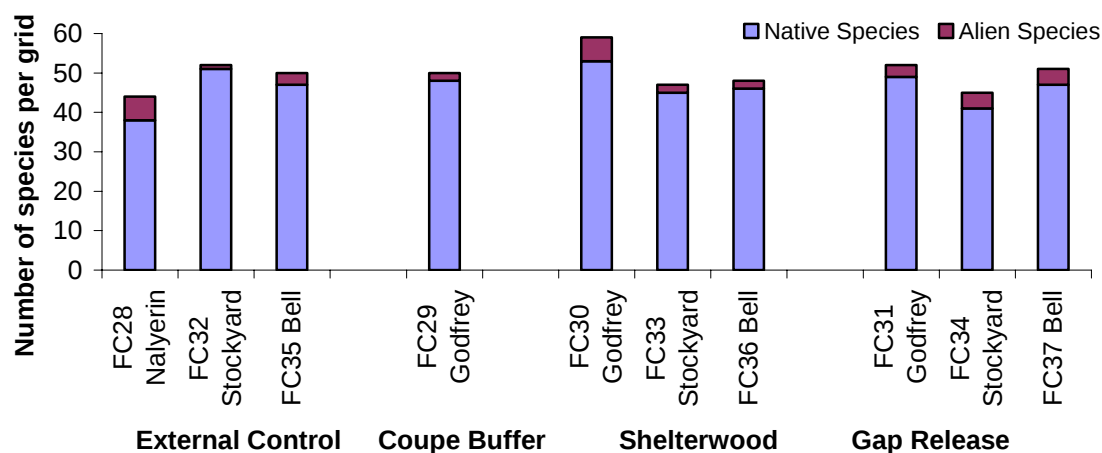


Figure 1. Vascular plant species richness recorded on each of the FORESTCHECK grids in Wellington East.

The minor variation in species richness is likely due to time since fire. The plots that were most recently burnt (2 years prior) had the highest species richness and the lowest species richness was in the Nalyerin external control grid (FC28), which was burnt 17 years previously. Introduced species are present in all grids, and when expressed as a proportion of the total they show a level of about 6-8%, with no difference between any of the treatments (including external control plots).

Species Density

Species density showed similar trends across all treatments and there appeared to be no impact associated with logging treatments (Fig. 2).

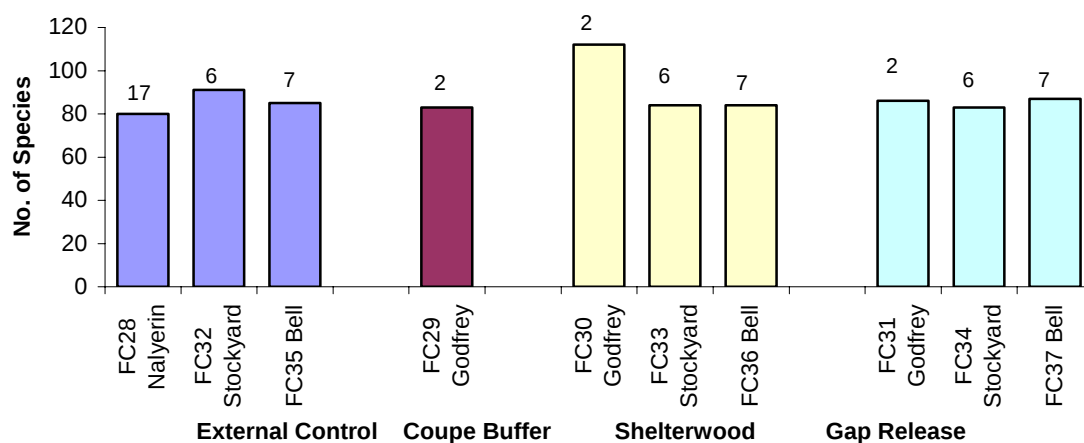


Figure 2. Mean species density (abundance) on each FORESTCHECK grid in Wellington East. Numbers above the columns represent years since fire.

With density (abundance) of plants, there was a clear effect of time since fire with the exception of the external control grids. The Nalyerin external control, which was unburnt for 17 years, contained fewer species at a higher density than the other external control grids. In the other treatments the most recently burnt grids had the highest plant density, which decreased with time since fire.

In order to determine the contribution that each species made to the vegetation complex, importance values were calculated. Importance values are based on cover, frequency and density of each plant. Each of the 3 attributes is rated according to predetermined ranges and the importance value is the proportion of the total for each plot. Cover, frequency and density importance values summed to provide a total importance value for each species (Mueller-Dombois and Ellenberg 1974). A mean of the total importance value was plotted for each grid (Fig. 3) and showed a high consistency across all treatments.

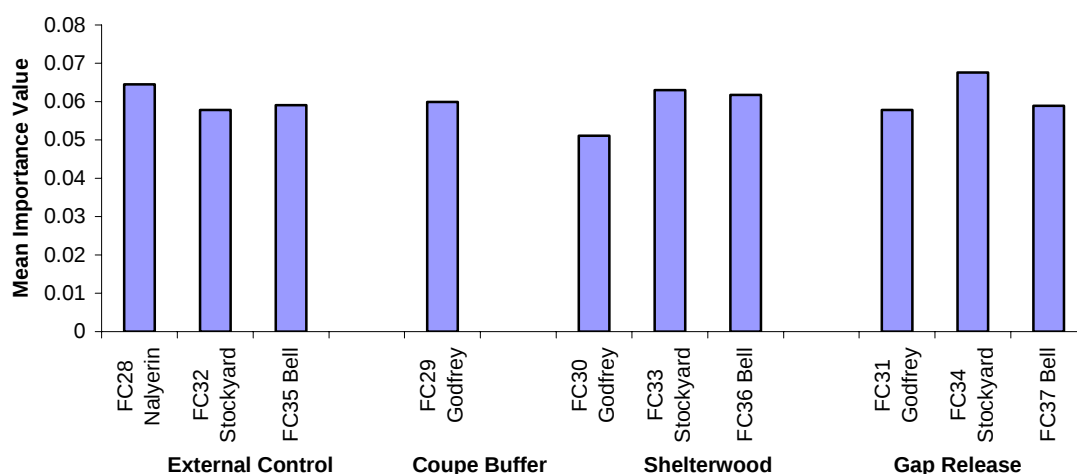


Figure 3. SMean total importance values for each grid.

Lifeform composition was generally consistent within each treatment, indicating that there have not been any major impacts associated with logging (Fig. 4).

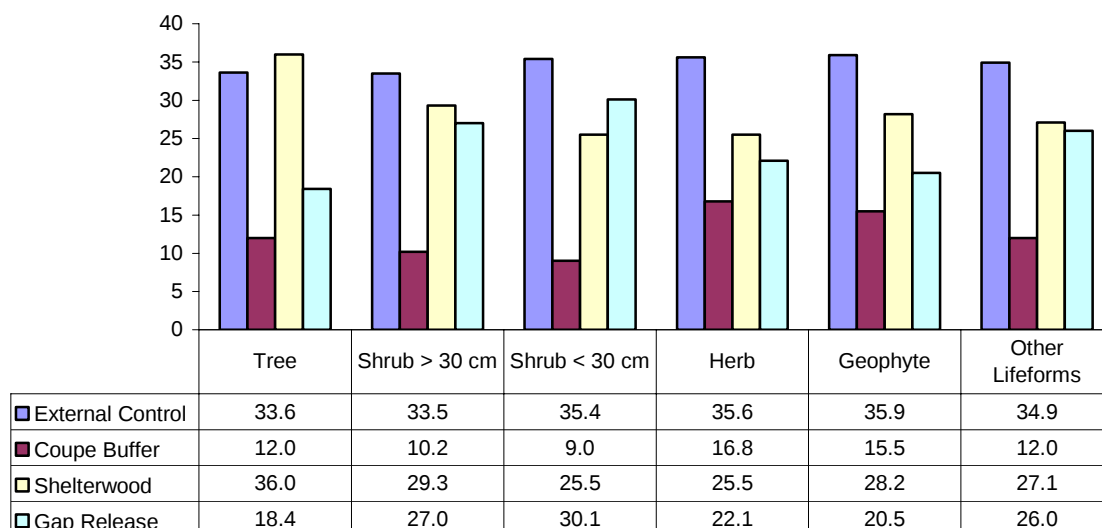


Figure 4. The proportion of plants in each life-form category in each treatment.

In each lifeform category the number of plants were summed and expressed as a percentage of the total. For the control grids there was a consistent representation across all lifeform categories. The coupe buffer, however, had low representation across all categories. This may be an artifact of there being only one coupe buffer compared to 3 grids in each of the other treatments.

Vegetation height was highly variable across the various grids (Fig. 5), and appeared to exhibit a successional effect linked to time since fire. Plant heights were lowest in the grids burnt 2 years previously and higher 6-7 years following fire, after which they declined as particular species reached the end of their life span. This is also reflected in the structural measurements where Levy contacts are most abundant in the lower height classes for the youngest age group (Fig. 6). Vegetation structure was similar within treatments but varied between treatments, and appeared to be related to time since fire.

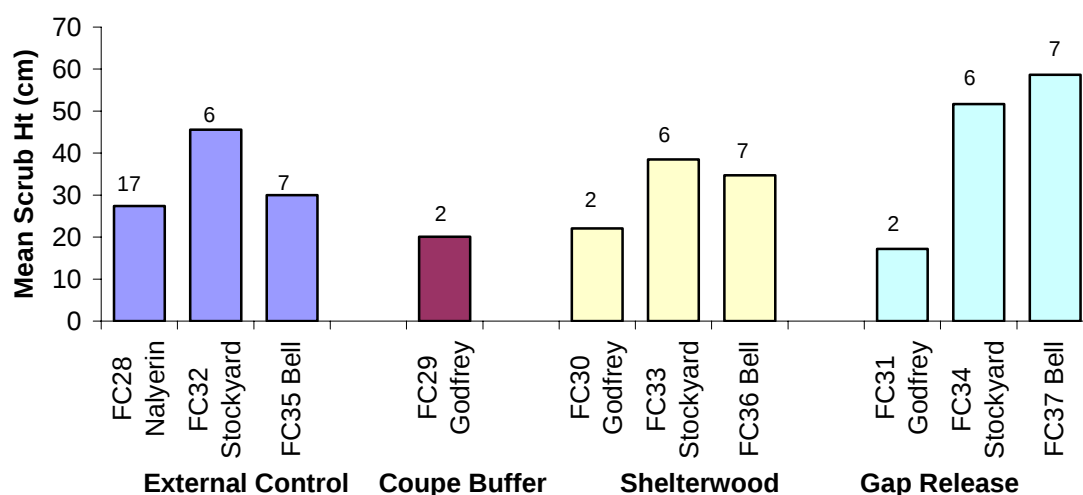


Figure 5. Mean scrub height (cm) measured in each FORESTCHECK grid in Wellington East. Numbers above the columns represent years since fire.

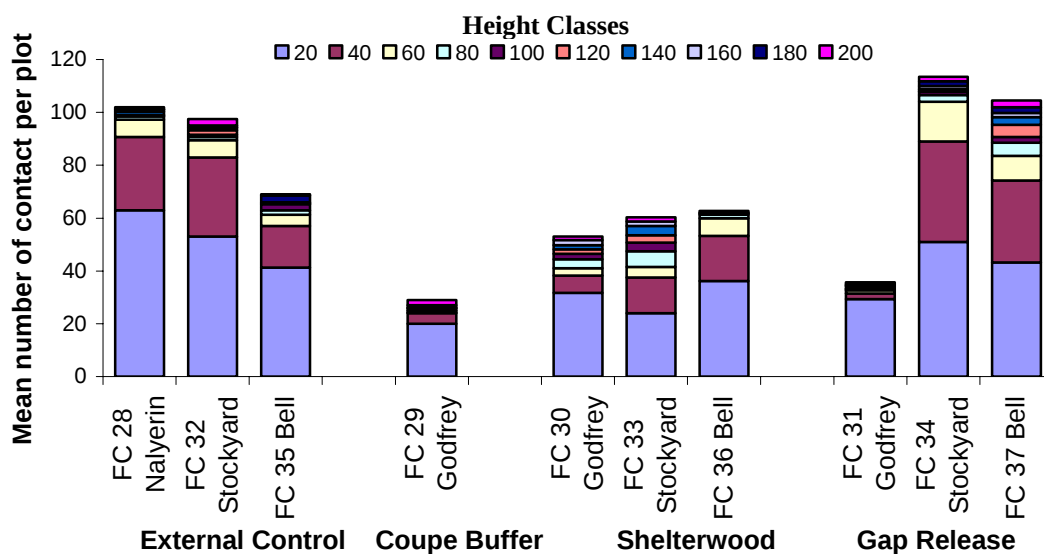


Figure 6. Vegetation structure in each grid (calculated from mean numbers of Levy contacts for height classes up to 2 m).

Conclusions

- There has been no apparent impact of logging on species richness and most variation appears to be due to time since fire.
- Plant density in shelterwood shows a trend in time since fire, with highest density recorded in the most recently burnt plot.
- Some categories of lifeform may have been impacted by logging treatments. Further analysis is needed to confirm this.

References

- Levy, E.B. and Madden, E.A. 1933. The point method of pasture analysis. *New Zealand Journal of Agriculture* **46**: 267-279.
- Mueller-Dombois, D. and Ellenberg, H. 1974. *Aims and Methods of Vegetation Ecology*. John Wiley & Sons, Sydney.

APPENDIX 1

TOTAL SPECIES LIST 2004-2005 SAMPLING

Introduced	Taxon Name	Taxon ID	SpCode	Lifeform	Lifestyle	Fire Response
	<i>Acacia alata</i>	3207	ACAALA	S	P	A1
	<i>Acacia browniana</i>	3247	ACABRO	S	P	A1
	<i>Acacia celastrifolia</i>	3254	ACACEL	S	P	A1
	<i>Acacia drummondii</i>	3311	ACADRU	S	P	A1
	<i>Acacia drummondii</i>	11661	ACADRUDR	S	P	A1
	<i>Acacia extensa</i>	3331	ACAEXT	S	P	A1
	<i>Acacia myrtifolia</i>	3453	ACAMYR	S	P	A1
	<i>Acacia preissiana</i>	3456	ACAPRE	DS	P	A1
	<i>Acacia pulchella</i>	3502	ACAPUL	S	P	A1
	<i>Acacia saligna</i>	3527	ACASAL	S	P	A1
*	<i>Acaena echinata</i>	3184	ACAECH	DS	P	A1
	<i>Actinotus glomeratus</i>	6203	ACTGLO	DS	P	A1
	<i>Agrostocrinum scabrum</i>	1261	AGRSCA	G	P	B3
*	<i>Aira cupaniana</i>	185	AIRCUP	GR	A	A1
	<i>Allocasuarina fraseriana</i>	1728	ALLFRA	T	P	B1
	<i>Anigozanthos manglesii</i>	1411	ANIMAN	S	P	B3
*	<i>Arctotheca calendula</i>	7838	ARCCAL	H	A	A1
	<i>Astroloma ciliatum</i>	6323	ASTCIL	DS	P	B2
	<i>Astroloma drummodii</i>	6325	ASTDRU	DS	P	B2
	<i>Astroloma pallidum</i>	6334	ASTPAL	DS	P	B2
	<i>Austrodanthonia caespitosa</i>	17950	AUSCAE	GR	P	B3
	<i>Austrostipa campylachne</i>	17233	AUSCAM	GR	P	B3
	<i>Banksia grandis</i>	1819	BANGRA	T	P	A2
	<i>Banksia shpaerocarpa</i>	1851	BANSPH	S	P	A2
	<i>Billardiera variifolia</i>	3165	BILVAR	V	P	A1
	<i>Boronia crenulata</i>	4413	BORCRE	DS	P	A1
	<i>Boronia gracilipes</i>	4422	BORGRA	DS	P	A1
	<i>Boronia ramosa</i>	4438	BORRAM	S	P	A1
	<i>Boronia spathulata</i>	4441	BORSPA	S	P	B2
	<i>Bossiaea eriocarpa</i>	3710	BOSERI	S	P	A1
	<i>Bossiaea ornata</i>	3714	BOSORN	S	P	A1
*	<i>Briza minor</i>	245	BRIMIN	GR	A	A1
	<i>Burchardia umbellata</i>	1387	BURUMB	G	P	B3
	<i>Caesia micrantha</i>	1276	CAEMIC	G	P	B3
	<i>Caladenia flava</i>	1592	CALFLA	G	P	B3
	<i>Caladenia macrostylis</i>	1604	CALMAC	G	P	B3
	<i>Caladenia reptans</i>	1613	CALREP	G	P	B3
	<i>Caladenia splendens</i>	15380	CALSPL	G	P	B3
	<i>Cassytha racemosa</i>	2957	CASRAC	P	P	A1
	<i>Cassytha sp.</i>		CASSP.	P	P	A1
*	<i>Centaurium erythraea</i>	6539	CENERY	H	A	A1
	<i>Centrolepis polygyna</i>	1134	CENPOL	H	A	A1
	<i>Chamaescilla corymbosa</i>	1280	CHACOR	G	P	B3
	<i>Clematis pubescens</i>	2929	CLEPUB	V	P	A1
	<i>Comesperma calymega</i>	4550	COMCAL	DS	P	B2
	<i>Conospermum caerulesens</i>	1862	CONCAE	S	P	A1
	<i>Conostylis aculeata</i>	1418	CONACU	DS	P	B3
	<i>Conostylis serrulata</i>	1453	CONSER	DS	P	B2
	<i>Conostylis setigera</i>	1454	CONSET	DS	P	B3
	<i>Corymbia calophylla</i>	17104	CORCAL	T	P	A2
	<i>Craspedia variabilis</i>	13354	CRAVAR	G	P	B3

Introduced	Taxon Name	Taxon ID	SpCode	Lifeform	Lifestyle	Fire Response
	<i>Cyanicula sericea</i>	15404	CYASER	G	P	B3
	<i>Cyrtostylis hugelii</i>	10916	CYRHUE	G	P	B3
	<i>Dampiera alata</i>	7420	DAMALA	DS	P	A1
	<i>Dampiera linearis</i>	7454	DAMLIN	DS	P	A1
	<i>Daucus glochidiatus</i>	6218	DAUGLO	H	A	A1
	<i>Daviesia incrassata</i>	3816	DAVINC	S	P	A1
	<i>Daviesia preissii</i>	3835	DAVPRE	S	P	A1
	<i>Desmocladus fasciculatus</i>	17691	DESFAS	Z	P	B3
	<i>Desmocladus flexuosus</i>	16595	DESFLE	Z	P	B3
	<i>Dianella revoluta</i>	1259	DIAREV	H	P	B3
	<i>Diuris corymbosa</i>	11049	DIUCOR	G	P	B3
	<i>Diuris longifolia</i>	1635	DIULON	G	P	B3
	<i>Drakaea gracilis</i> ms	15406	DRAGRA	G	P	B3
	<i>Drosera erythrorhiza</i>	3095	DROERY	G	P	B3
	<i>Drosera huegelii</i>	3102	DROHUE	G	P	B3
	<i>Drosera mensiesii</i> subsp. <i>menziesii</i>	11853	DROMENME	G	P	B3
	<i>Drosera pallida</i>	3118	DROPAL	G	P	B3
	<i>Drosera stolonifera</i>	3131	DROSTO	G	P	B3
	<i>Dryandra bipinnatifida</i>	1891	DRYBIP	DS	P	B2
	<i>Dryandra lindleyana</i>	16672	DRYLIN	S	P	B2
	<i>Dryandra sessilis</i>	1932	DRYSES	S	P	A1
	<i>Elythranthera brunonis</i>	1643	ELYBRU	G	P	B3
	<i>Eucalyptus marginata</i>	5708	EUCMAR	T	P	A2
	<i>Glischrocaryon aureum</i>	6143	GLIAUR	S	p	B2
	<i>Gompholobium capitatum</i>	3948	GOMCAP	DS	P	A1
	<i>Gompholobium confertum</i>	10909	GOMCON	S	P	A1
	<i>Gompholobium knightianum</i>	3950	GOMKNI	DS	P	A1
	<i>Gompholobium marginatum</i>	3951	GOMMAR	DS	P	A1
	<i>Gompholobium preissii</i>	3955	GOMPRE	DS	P	A1
	<i>Gompholobium scabrum</i>	11083	GOMSCA	S	P	A1
	<i>Hakea amplexicaulis</i>	2128	HAKAMP	S	P	B2
	<i>Hakea cyclocarpa</i>	2152	HAKCYC	S	P	B2
	<i>Hakea lissocarpha</i>	2175	HAKLIS	S	P	B2
	<i>Hakea undulata</i>	2215	HAKUND	S	P	B2
	<i>Helichrysum macranthum</i>	8027	HELMAC	H	A	A1
	<i>Hemiandra pungens</i>	6839	HEMPUN	S	P	A1
	<i>Hemigenia rigida</i>	6868	HEMRIG	DS	P	B2
	<i>Hibbertia amplexicaulis</i>	5109	HIBAMP	S	P	B2
	<i>Hibbertia commutata</i>	5114	HIBCOM	S	P	B2
	<i>Hibbertia hypericoides</i>	5135	HIBHYP	S	P	B2
	<i>Hibbertia racemosa</i>	5162	HIBRAC	S	P	A1
	<i>Hibbertia spicata</i>	5171	HIBSPI	S	P	B2
	<i>Homalosciadium homalocarpum</i>	6222	HOMHOM	H	A	A1
	<i>Hovea chorizemifolia</i>	3964	HOVCHO	DS	P	B2
	<i>Hovea elliptica</i>	3965	HOVELL	S	P	B2
	<i>Hovea trisperma</i>	3968	HOVTRI	S	P	A1
	<i>Hyalopserma cotula</i>	12714	HYACOT	H	A	A1
	<i>Hydrocotyle diantha</i>	6229	HYDDIA	H	A	A1
	<i>Hypocalymma angustifolium</i>	5817	HYPANG	S	P	B2
*	<i>Hypochaeris glabra</i>	8086	HYPGLA	H	A	A1
	<i>Hypoxis occidentalis</i>	1503	HYPOCC	H	P	B3
	<i>Isolepis marginata</i>	917	ISOMAR	R	A	A1
	<i>Isotoma hypocrateriformis</i>	7396	ISOHYP	H	A	A1
	<i>Kennedia coccinea</i>	4037	KENCOC	V	P	A1
	<i>Kennedia prostrata</i>	4044	KENPRO	V	P	A1

Introduced	Taxon Name	Taxon ID	SpCode	Lifeform	Lifestyle	Fire Response
	<i>Labichea punctata</i>	3669	LABPUN	DS	P	B2
	<i>Lagenophora huegelii</i>	18585	LAGHUE	G	P	B3
	<i>Lasiopetalum floribundum</i>	5033	LASFLO	S	P	A1
	<i>Laxmannia squarrosa</i>	1309	LAXSQU	DS	P	A1
	<i>Lechenaultia biloba</i>	7568	LECBIL	S	P	A1
	<i>Lepidosperma leptostachyum</i>	936	LEPLEP	Z	P	B3
	<i>Lepidosperma squamatum</i>	945	LEPSQU	Z	P	B3
	<i>Leptomeria cunninghamii</i>	2342	LEPCUN	S	P	A1
	<i>Leucopogon capitellatus</i>	6367	LEUCAP	S	P	B2
	<i>Leucopogon propinquus</i>	6436	LEUPRO	S	P	B2
	<i>Levenhookia pusilla</i>	7676	LEVPUS	H	A	A1
	<i>Lobelia rarifolia</i>	7405	LOBRAR	H	A	A1
	<i>Logania serpyllifolia</i>	6511	LOGSER	DS	P	B2
	<i>Lomandra caespitosa</i>	1223	LOMCAE	DS	P	B3
	<i>Lomandra drummondii</i>	1225	LOMDRU	DS	P	B3
	<i>Lomandra hermaphrodita</i>	1228	LOMHER	DS	P	B2
	<i>Lomandra integra</i>	1229	LOMINT	DS	P	B3
	<i>Lomandra preissii</i>	1239	LOMPRE	DS	P	B3
	<i>Lomandra purpurea</i>	1240	LOMPUR	DS	P	B3
	<i>Lomandra sericea</i>	1243	LOMSER	DS	P	B3
	<i>Lomandra sp.</i>	-801	LOMSP	DS	P	B3
	<i>Lomandra sparteae</i>	1245	LOMSPA	DS	P	B3
	<i>Lomandra suaveolens</i>	1246	LOMSUA	DS	P	B2
	<i>Luzula merioionalis</i>	1198	LUZMER	R	P	B3
	<i>Macrozamia riedlei</i>	85	MACRIE	C	P	B3
	<i>Millotia tenuifolia</i>	8106	MILTEN	H	A	A1
	<i>Monotaxis grandiflora</i>	4662	MONGRA	DS	P	A1
	<i>Neurachne alopecuroidea</i>	492	NEUALO	GR	P	B3
	<i>Olax benthamiana</i>	2365	OLABEN	S	P	A1
	<i>Opercularia hispidula</i>	7348	OPEHIS	S	P	A1
	<i>Opercularia vaginata</i>	18255	OPEVAG	S	P	A1
	<i>Orchid sp.</i>		ORCSP.	G	P	B3
*	<i>Oxalis corniculata</i>	4349	OXACOR	G	P	B3
*	<i>Parentucellia latifolia</i>	7089	PARLAT	H	A	A1
	<i>Patersonia babianoides</i>	1542	PATBAB	G	P	B3
	<i>Patersonia juncea</i>	1546	PATJUN	DS	P	B3
	<i>Patersonia pygmaea</i>	1551	PATPYG	DS	P	B3
	<i>Patersonia umbrosa</i>	1553	PATUMB	DS	P	B3
	<i>Pelargonium littorale</i>	4346	PELLIT	DS	p	a1
	<i>Pentapeltis peltigera</i>	6245	PENPEL	DS	P	B2
	<i>Pentapeltis silvatica</i>	6246	PENSIL	S	P	B2
	<i>Persoonia longifolia</i>	2267	PERLON	S	P	B2
	<i>Petrophile diversifolia</i>	2293	PETDIV	S	P	A1
	<i>Petrophile serruriae</i>	2309	PETSER	S	P	A1
	<i>Petrophile striata</i>	2312	PETSTR	S	P	A1
	<i>Pheladenia deformis</i>	20460	PHEDEF	G	P	B3
	<i>Phyllangium paradoxum</i>	16177	PHYPAR	H	A	A1
	<i>Phyllangium paradoxum</i>	16177	PHYPAR	H	A	A1
	<i>Phyllanthus calycinus</i>	4675	PHYCAL	DS	P	B2
	<i>Pimelea angustifolia</i>	5231	PIMANG	S	P	A1
	<i>Pimelea imbricata</i>	5251	PIMIMB	S	P	A1
	<i>Pimelea preissii</i>	5259	PIMPRE	S	P	A1
	<i>Pimelea rosea</i>	5261	PIMROS	S	P	A1
	<i>Pimelea sp.</i>	-42	PIMSP.	S	P	A1
	<i>Pimelea suaveolens</i>	5266	PIMSUA	S	P	B2

Introduced	Taxon Name	Taxon ID	SpCode	Lifeform	Lifestyle	Fire Response
	<i>Pimelea suaveolens</i> subsp. <i>suaveolens</i>	12041	PIMSUASU	S	P	B2
	<i>Poa drummoniana</i>	573	POADRU	GR	P	B3
	<i>Podolepis canescens</i>	8172	PODCAN	H	A	A1
	<i>Podotheca angustifolia</i>	8182	PODANG	H	A	A1
	<i>Podotheca gnaphalioides</i>	8184	PODGNA	H	A	A1
	<i>Poranthera huegelii</i>	4690	PORHUE	DS	A	A1
	<i>Poranthera microphylla</i>	4691	PORMIC	DS	A	A1
	<i>Pterostylis pyramidalis</i>	11118	PTEPYR	G	P	B3
	<i>Pterostylis recurva</i>	1693	PTEREC	G	P	B3
	<i>Pterostylis vittatus</i>	1698	PTEVIT	G	P	B3
	<i>Ptilotus drummondii</i>	2718	PTIDRU	G	P	B3
	<i>Ptilotus manglesii</i>	2742	PTIMAN	G	P	B3
	<i>Pyrorchis nigricans</i>	16367	PYRNIG	G	P	B3
	<i>Quinetia urvillei</i>	8195	QUIURV	H	A	A1
	<i>Ranunculus colonorum</i>	2932	RANCOL	G	P	B3
	<i>Rhodanthe citrina</i>	13300	RHOCIT	H	A	A1
	<i>Rhodanthe manglesii</i>	13234	RHOMAN	H	A	A1
	<i>Scaevola striata</i>	7646	SCASTR	DS	A	A1
	<i>Senecio glomeratus</i>	8206	SENGLO	H	A	A1
	<i>Senecio hispidulus</i>	8208	SENHIS	S	A	A1
	<i>Senecio leucoglossus</i>	8212	SENLEU	S	A	A1
	<i>Sollya fusiformis</i>	8922	SOLFUS	S	P	U
*	<i>Sonchus oleraceus</i>	8231	SONOLE	H	A	A1
	<i>Sowerbaea laxiflora</i>	1312	SOWLAX	G	P	B3
	<i>Stackhousia monogyna</i>	4733	STAMON	S	P	B2
	<i>Stylidium amoenum</i>	7684	STYAMO	DS	P	A1
	<i>Stylidium brunonianum</i>	7693	STYBRU	DS	P	A1
	<i>Stylidium calcaratum</i>	7696	STYCAL	H	A	A1
	<i>Stylidium ciliatum</i>	7702	STYCIL	DS	P	A1
	<i>Stylidium lineatum</i>	7752	STYLIN	DS	P	B2
	<i>Stylidium piliferum</i>	7774	STYPIL	DS	P	A1
	<i>Stylidium schoenoides</i>	7798	STYSCH	DS	P	A1
	<i>Stylidium spathulatum</i>	7799	STYSPA	DS	P	A2
	<i>Stypandra glauca</i>	1260	STYGLA	S	P	B3
	<i>Styphelia tenuiflora</i>	6476	STYTEN	S	P	A1
	<i>Synaphea petiolaris</i>	2324	SYNPET	S	P	B3
	<i>Tetraria capillaris</i>	1034	TETCAP	S	P	B3
	<i>Tetraria octandra</i>	1036	TETOCT	Z	P	B3
	<i>Tetrarrhena laevis</i>	667	TETLAE	GR	P	B3
	<i>Tetratheca hirsuta</i>	4535	TETHIR	S	P	A1
	<i>Thelymitra crinita</i>	1705	THECRI	G	P	B3
	<i>Thysanotus manglesianus</i>	1338	THYMAN	G	P	B3
	<i>Thysanotus multiflorus</i>	1339	THYMUL	G	P	B3
	<i>Thysanotus patersonii</i>	1343	THYPAT	G	P	B3
	<i>Thysanotus sparteus</i>	1351	THYSPA	G	P	B3
	<i>Trachymene pilosa</i>	6280	TRAPIL	H	A	A1
	<i>Trichocline spathulata</i>	8251	TRISPA	G	P	B3
	<i>Tricoryne elatior</i>	1361	TRIELA	DS	P	A1
	<i>Tricoryne humilis</i>	1362	TRIHUM	DS	P	A1
	<i>Trymalium ledifolium</i>	4842	TRYLED	S	P	A1
	<i>Velleia trinervis</i>	7665	VELTRI	DS	A	A1
*	<i>Vulpia bromoides</i>	722	VULBRO	GR	A	A1
	<i>Wahlenbergia gracilentia</i>	7386	WAHGRA	H	A	A1
	<i>Waitzia acuminata</i>	8275	WAIACU	H	A	A1
	<i>Wurmbia dioica</i>	1394	WURDIO	G	P	B3

Introduced	Taxon Name	Taxon ID	SpCode	Lifeform	Lifestyle	Fire Response
	<i>Xanthorrhoea gracilis</i>	1253	XANGRA	X	P	B2
	<i>Xanthorrhoea preissii</i>	1256	XANPRE	X	P	B2
	<i>Xanthosia atkinsoniana</i>	6283	XANATK	S	P	B2
	<i>Xanthosia candida</i>	6284	XANCAN	DS	P	B2
	<i>Xanthosia huegelii</i>	6289	XANHUE	DS	P	A1

* 10 introduced species

INVERTEBRATES

Janet Farr, Allan Wills, Tom Burbidge and Paul Van Heurck

Introduction

Invertebrates, including the class Insecta, comprise > 75% of Earth's biodiversity and therefore represent a crucial component of any ecosystem. Invertebrates play a major role in decomposition, nutrient recycling and provide an important food source for vertebrates. In addition a wide range of species are already known to be exclusive to the southwest forests of Western Australia, and some of these are Gondwanan relicts. Despite this, current knowledge of the invertebrate taxa present in the jarrah forest is limited.

The objectives of this component of FORESTCHECK monitoring are to:

- Monitor and record the species of invertebrates in the various treatments of managed jarrah (Gap Release, Shelterwood) and unlogged forest.
- Analyse trends in species composition, richness and abundance
- Monitor the presence of Gondwanan relict and affinity invertebrate species with respect to the above treatments
- Monitor the presence of known insect pest species.

Monitoring and Laboratory Work

Sampling at Wellington East was carried out in November (spring) 2004 and April (autumn) 2005 using the protocol formerly established at Donnelly. To briefly summarize: active capture samples, involving sweeping, beating, and habitat searches of coarse woody debris (CWD) and litter were conducted once at each site for a total time of 1 person hour per capture/habitat method. Light traps were run for 3 nights simultaneously at each site (there was no light trap failure), achieving one trap night per week for 3 weeks; pitfall traps were opened for 10 days simultaneously at each site. Captures were bagged and labelled according to site and other capture details in the field, then transported in an insulated container back to a base camp, where they were stored in a portable freezer. At the conclusion of a sampling period, specimens were transported to the laboratory in Manjimup where they were sorted and assigned to morphospecies. This report examines invertebrate captures made in the eastern jarrah forest at Wellington East and includes some comparison with the Donnelly and Wellington sites.

This is a preliminary report, and previously sampled results are from data as at July 2005; morphospecies assignment and Gondwanan relationships will change following ongoing taxonomic examination of specimens.

Results

Total Species

The Wellington East samples collected in 2004-05 increased the number of morphospecies to 1 271 (Appendix 1). Fig. 1 shows the cumulative captures for the successive sampling locations.

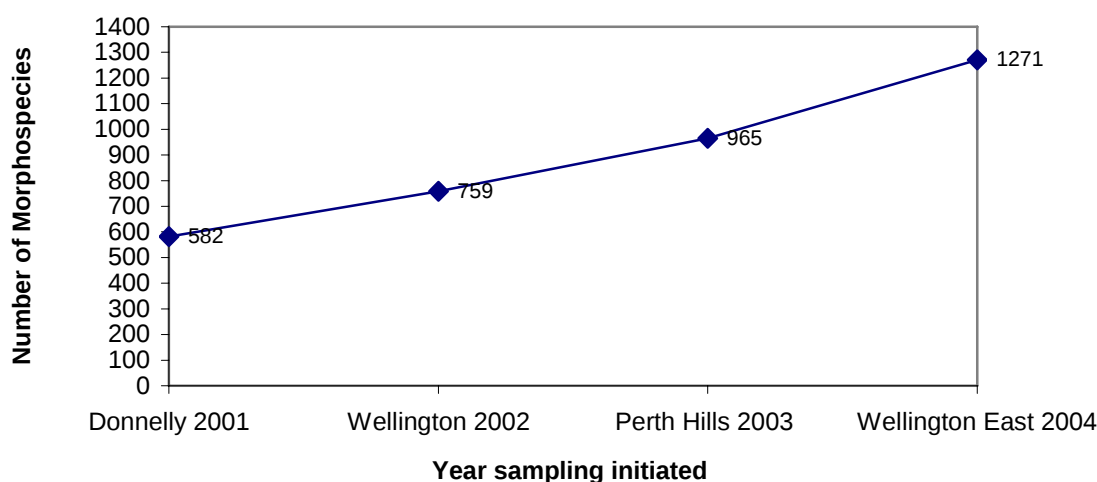


Figure 1. Cumulative morphospecies for all FORESTCHECK locations (2001-2005).

A total of 670 morphospecies were collected from Wellington East compared to 450, 381 and 582 from Perth Hills, Wellington and Donnelly locations respectively. Of those morphospecies collected from Wellington East, 21 were considered Gondwanan relicts and a further 65 species had Gondwanan affinities compared to 13 and 48 respectively for Donnelly. The numbers of morphospecies for orders where 10 or more morphospecies have been assigned are compared for Wellington East, Perth Hills, Wellington and Donnelly locations in Fig. 2.

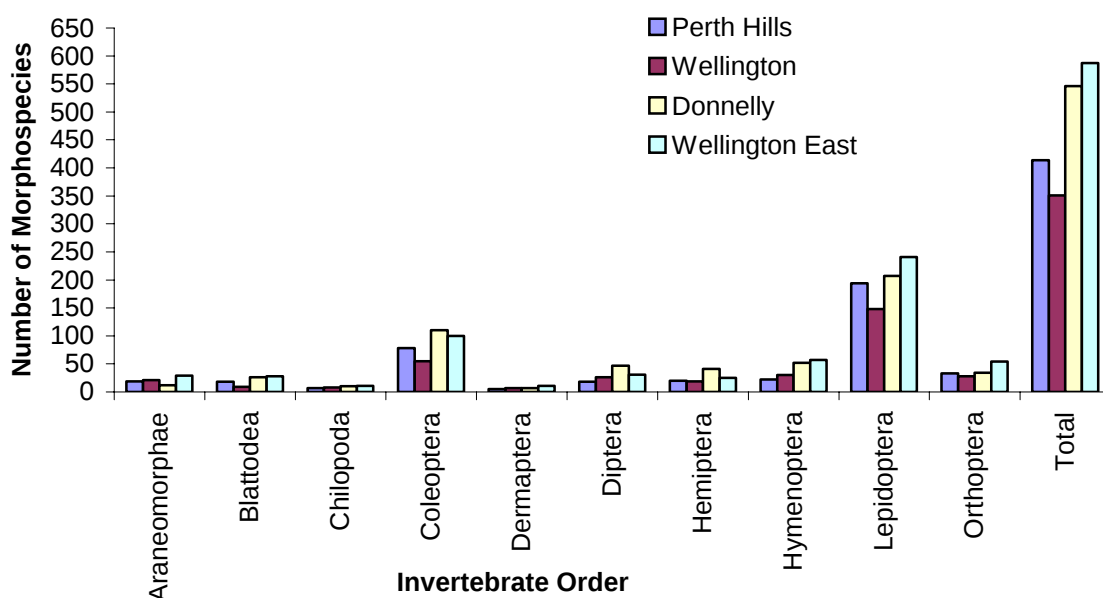


Figure 2. Comparison of all FORESTCHECK locations (2001-2005) morphospecies numbers for invertebrate orders where ten or greater morphospecies have been assigned.

Overall, Wellington East had the greatest species diversity, and Donnelly second, with this pattern repeated in Araneomorpha (spiders), Dermaptera (earwigs), Lepidoptera (moths) and Orthoptera (crickets and grasshoppers). Species diversity in Blattodea (cockroaches), Chilopoda (centipedes) and Hymenoptera (wasps) were similar for both Wellington East and Donnelly, whereas Donnelly had the greatest diversity in Coleoptera (beetles), Diptera (flies) and Hemiptera (bugs).

Comparing capture methods

Light trapping resulted in the most abundant and diverse captures (Table 1) with a spring capture of 300 morphospecies comprising 26 163 individuals. This high number of individuals was mainly due to bulk captures of a hydrophilid beetle (species # 14) of which 19 656 individuals were caught in light traps. As expected, diversity and abundance for all capture methods were highest in spring.

Table 1. Number of morphospecies and specimen abundance for Wellington East in spring and autumn for each capture method (CWD = coarse woody debris).

Capture Method	No of Morphospecies		Abundance	
	Spring	Autumn	Spring	Autumn
Light	300	167	26 163	1 536
Pitfall	69	40	262	127
Sweep	49	23	130	38
Beat	49	17	103	25
CWD	52	34	77	53
Litter	31	18	59	20
Targeted pursuit	13	7	15	7
Total			26 809	1 806

Fig. 3 shows morphospecies comparisons for capture methods from Wellington East, Perth Hills, Wellington and Donnelly locations. Abundance comparisons are shown in Fig. 4. In spring, with the exception of the light trap and coarse woody debris (CWD) captures, the Donnelly captures are more diverse (Fig. 3a). Pit fall and Sweep captures for both Wellington samples are similar. Coarse woody debris captures are greatest at Wellington East but this could be due to a variation in the collection method for CWD at the Wellington sites, where one operator was dedicated to CWD compared to dividing CWD search between 3 operators on the other sites. However, for autumn, Wellington East and Donnelly diversity levels (number of morphospecies) for CWD are comparable (Fig. 3b) as are the other sampling methods with the exception of light traps. Also, the abundance data for autumn show Donnelly CWD captures are much greater than those for Wellington East (Fig. 4b). Therefore we may conclude that varying the active capture method by dedicating a single operator to this task has not significantly altered capture results.

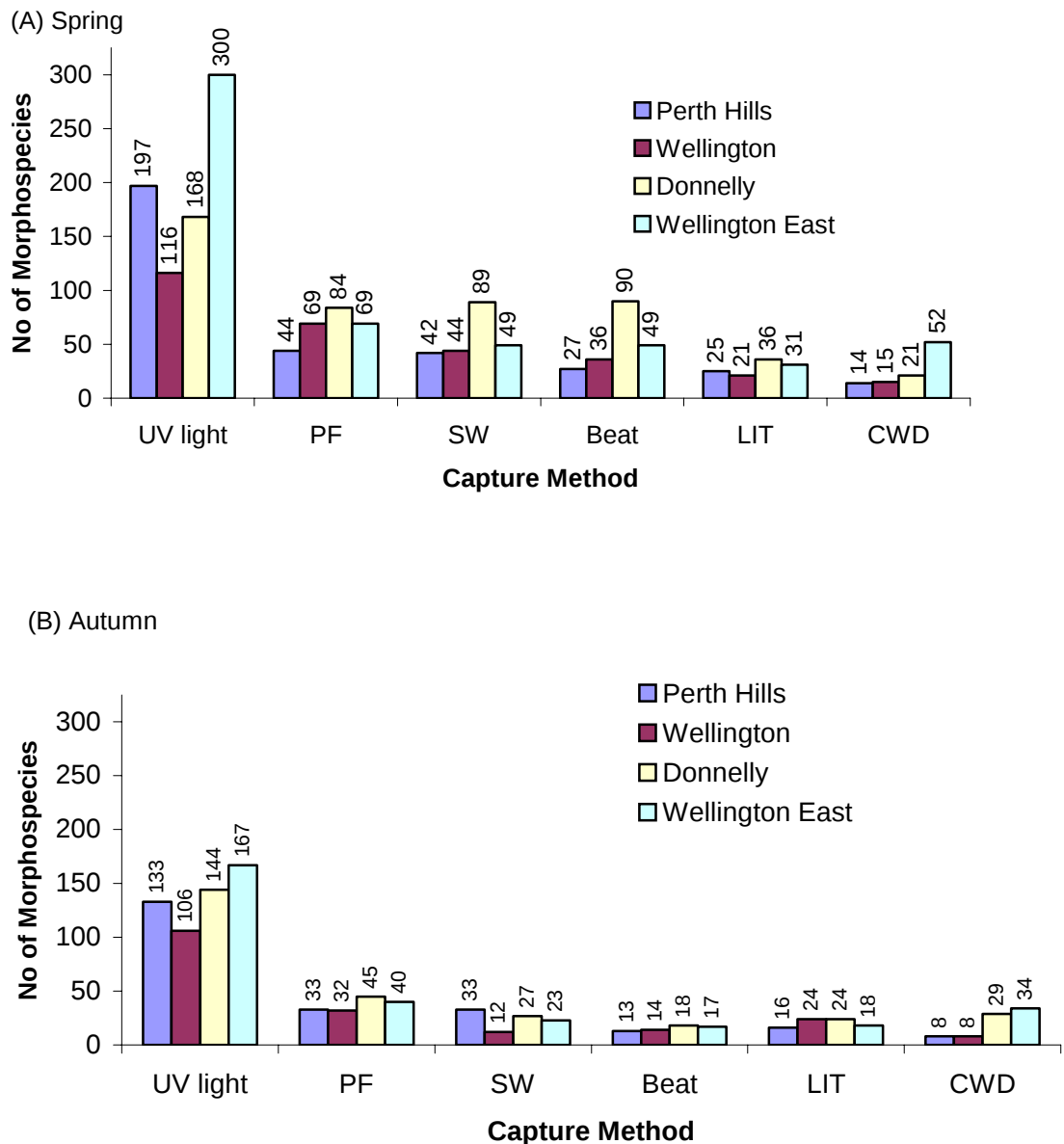


Figure 3. The number of morphospecies in spring (A) and autumn (B) for each capture method; Wellington East Perth Hills, Wellington and Donnelly compared. (PF= pitfall trap; SW = sweep net; Beat = beating tray, LIT = litter search, CWD = coarse woody debris search).

For abundance, spring light trap captures were most abundant for Wellington East with 26 163 individuals compared to 3 500 individuals for Perth Hills (Fig. 4a). Correcting for the high number of the hydrophilid beetle (species # 14) from spring light traps, captures reduce to 6 507 which is still much greater than the number of individuals caught at other sites. In autumn, light trap capture abundance for Wellington East was also high compared to other sampling districts (Fig. 4b). Abundances from pitfall captures were comparable between both Wellington samples. For sweep, beat and CWD, Donnelly had the highest abundance.

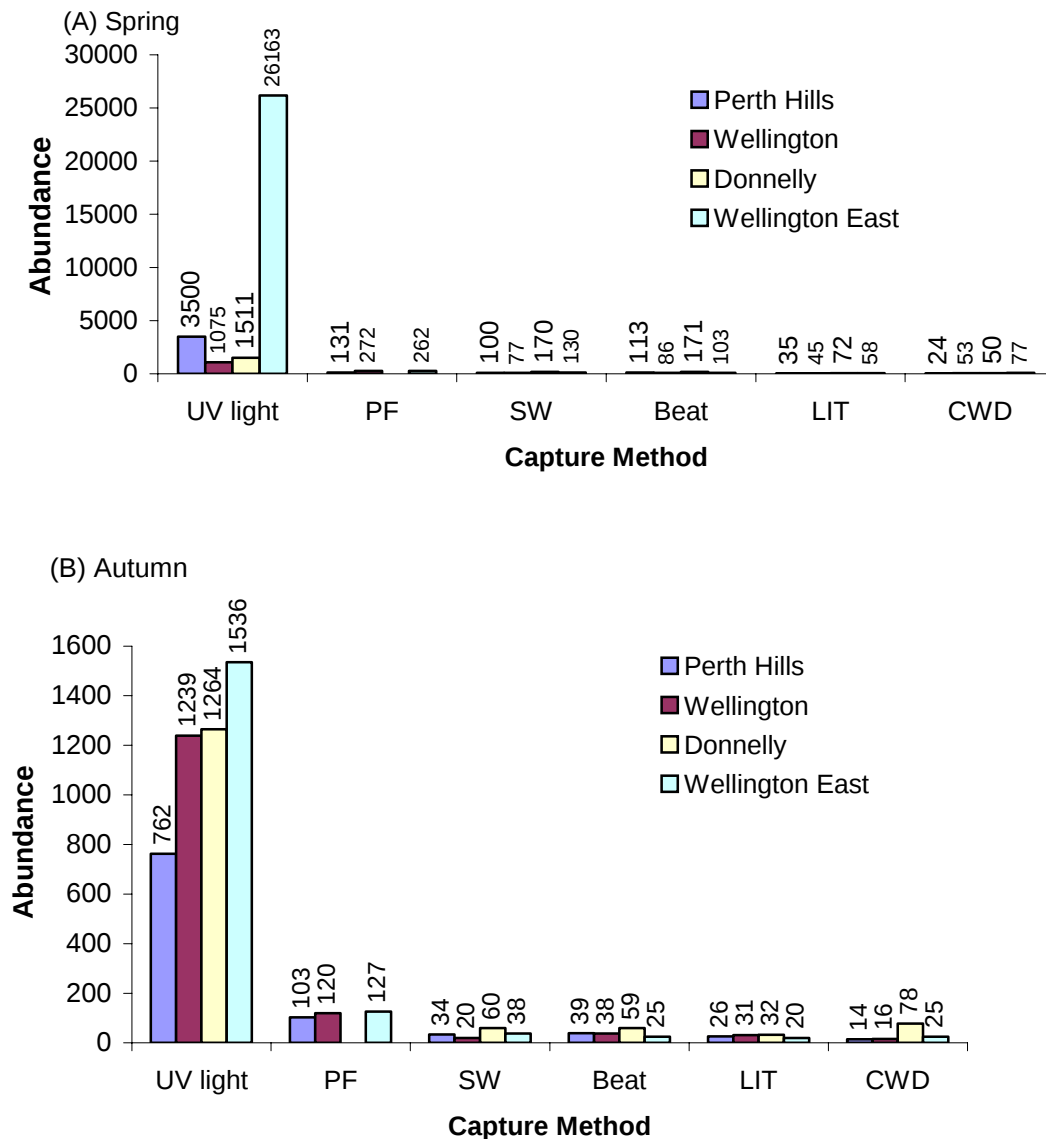


Figure 4. Abundance (measured as the number of individual specimens captured) in spring (A) and autumn (B) for each capture method, from Wellington East, Perth Hills, Wellington and Donnelly. (UV Light = light trap captures; PF= pitfall trap captures; SW = sweep net; Beat = beating tray; LIT = litter search; CWD = coarse woody debris search).

Comparing sample grids and silvicultural treatments

Fig. 5 shows Wellington East grid comparisons for silvicultural treatments expressed as the total morphospecies and abundance for all capture methods and summed for spring and autumn seasons. There was very little difference in species diversity between sites. The Godfrey shelterwood (FC30) had the greatest number of species (278), followed by Stockyard gap release (FC34) treatment and the Bell external control (FC35). Abundance was highest at the Stockyard gap release grid, followed by the Stockyard shelterwood (FC33). The lowest diversity and abundance was at the Godfrey gap release (FC 31). A species of Onychophora (species # 1481) was found in the Nalyerin external control grid (FC28).

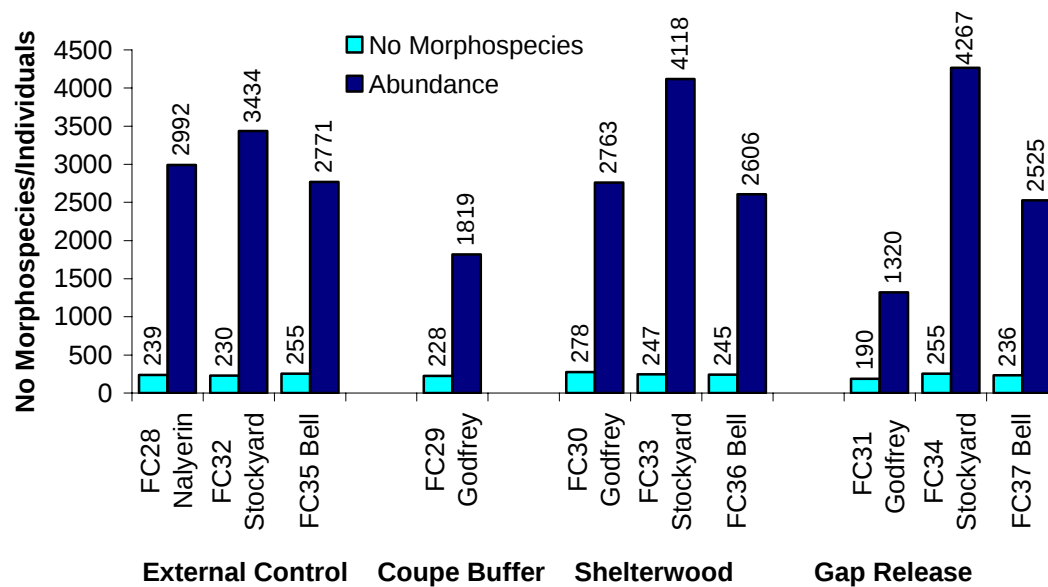


Figure 5. Comparison of individual Wellington East treatment grids for total morphospecies (no. of species) and abundance (no. of individuals), for all capture methods, combining both seasons.

Comparison of the means between treatments indicates few distinct patterns. For Wellington the controls are less diverse but have comparable invertebrate abundance with the shelterwood sites (Table 2). For Wellington East species number and abundance is similar across most treatments, with the gap release treatment showing a lower abundance.

Table 2. Comparison of means ($\pm$ SE) for number of species and abundance (number of individuals) at Wellington East and Wellington in respect to silvicultural treatment (the coupe buffer site was not included in the analysis).

Silvicultural Treatment	Wellington		Wellington East	
	No. of Species	Abundance	No. of Species	Abundance
External Control	108.7 (12.8)	337.3 (31.7)	245.0 (4.9)	3065.7 (194.9)
Shelterwood	116.3 (0.9)	329.7 (16.3)	227.3 (19.0)	3162.3 (480.0)
Gap Release	120.0 (7.2)	357.0 (53.2)	257.0 (11.0)	2704.0 (855.4)

The number of morphospecies and abundance (number of individuals captured at each grid) for active (beat, sweep, coarse woody debris and litter searches), light and pitfall trap capture techniques in spring and autumn is shown in Table 3.

Fig. 6 shows comparisons for silvicultural treatments for light trap captures between Wellington East and Donnelly in spring. For Wellington East, species diversity and abundance are lowest in the coupe buffer (single sample), and the gap treatment also has a lower diversity and abundance compared with the external control and shelterwood treatment. Lower captures in the coupe buffer site are also apparent for the active capture and pit falls (Table 3). For Donnelly spring light trap samples the shelterwood treatment had lower diversity and abundance. However in autumn, the external control treatment

was lowest in diversity and abundance followed closely by the gap release treatment (Fig. 7). This pattern is also evident in the Wellington East autumn data.

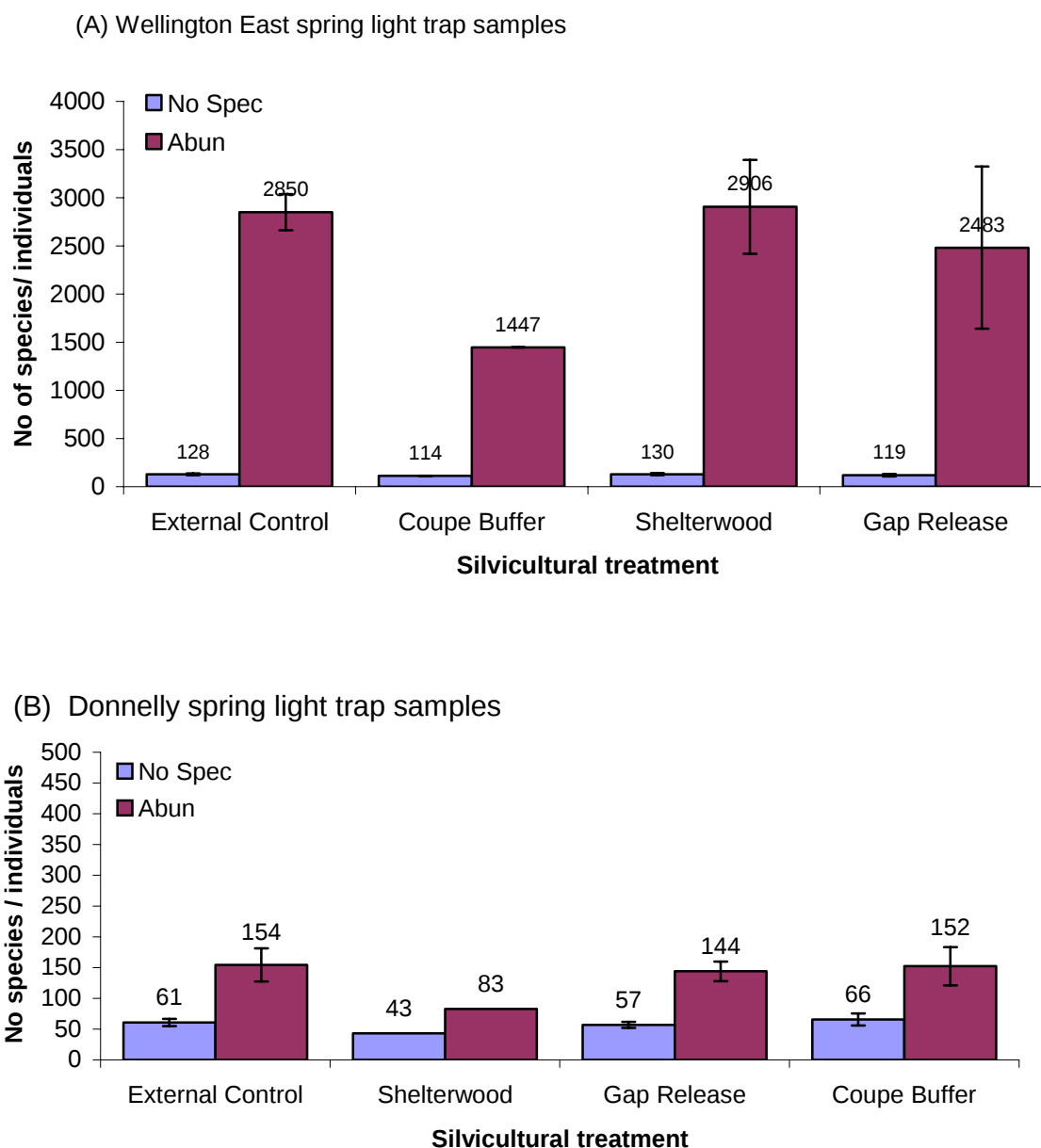


Figure 6. Mean ($\pm$ SE) spring light trap captures against treatment grids for (a) Wellington East and (b) Donnelly. (number of morphospecies = No Spec; number of individuals = abundance).

Species differences

Table 4 shows the most frequent species captured for Wellington East, Perth Hills, Wellington and Donnelly. Lepidopteran 436 was the most common species collected in Wellington East, Perth Hills and Wellington but does not feature in the 10 most common species captured in Donnelly (captured on 4 occasions only). Trichopteran 145 and species 52 (the introduced honey bee) were the only species present in the top 10 of all sample sites.

Table 3. Number of morphospecies and abundance (number of individuals captured at each grid) for active (beat, sweep, coarse woody debris, litter), light and pitfall trap capture techniques in spring and autumn for Wellington East.

Treatment	Site No	Location	Season	<u>Active capture</u>		<u>Light trap</u>		<u>Pitfall trap</u>		<u>All capture methods</u>	
				No Spec	Abund	No Spec	Abund	No Spec	Abund	No Spec	Abund
External Control	FC28	Nalyerin	AU	22	25	42	107	9	12	73	144
External Control	FC28	Nalyerin	SP	33	48	116	2746	17	54	166	2848
External Control	FC32	Stockyard	AU	14	15	42	128	6	17	62	160
External Control	FC32	Stockyard	SP	32	35	119	3213	17	26	168	3274
External Control	FC35	Bell	AU	13	15	47	93	6	8	66	116
External Control	FC35	Bell	SP	23	27	148	2591	18	37	189	2655
Coupe Buffer	FC29	Godfrey	AU	13	13	70	300	7	17	90	330
Coupe Buffer	FC29	Godfrey	SP	19	32	114	1447	5	10	138	1489
Shelterwood	FC30	Godfrey	AU	11	10	72	249	6	10	88	270
Shelterwood	FC30	Godfrey	SP	39	29	150	2441	12	13	191	2493
Shelterwood	FC33	Stockyard	AU	19	18	50	123	8	18	76	160
Shelterwood	FC33	Stockyard	SP	24	18	124	3880	29	54	171	3958
Shelterwood	FC36	Bell	AU	13	11	71	137	6	7	88	157
Shelterwood	FC36	Bell	SP	45	35	115	2397	7	7	157	2449
Gap Release	FC31	Godfrey	AU	12	10	47	134	4	5	61	151
Gap Release	FC31	Godfrey	SP	36	25	100	1128	5	5	130	1169
Gap Release	FC34	Stockyard	AU	11	9	49	137	12	17	70	165
Gap Release	FC34	Stockyard	SP	49	33	140	4029	12	24	185	4102
Gap Release	FC37	Bell	AU	9	8	51	128	9	16	68	153
Gap Release	FC37	Bell	SP	49	33	118	2291	17	32	168	2372

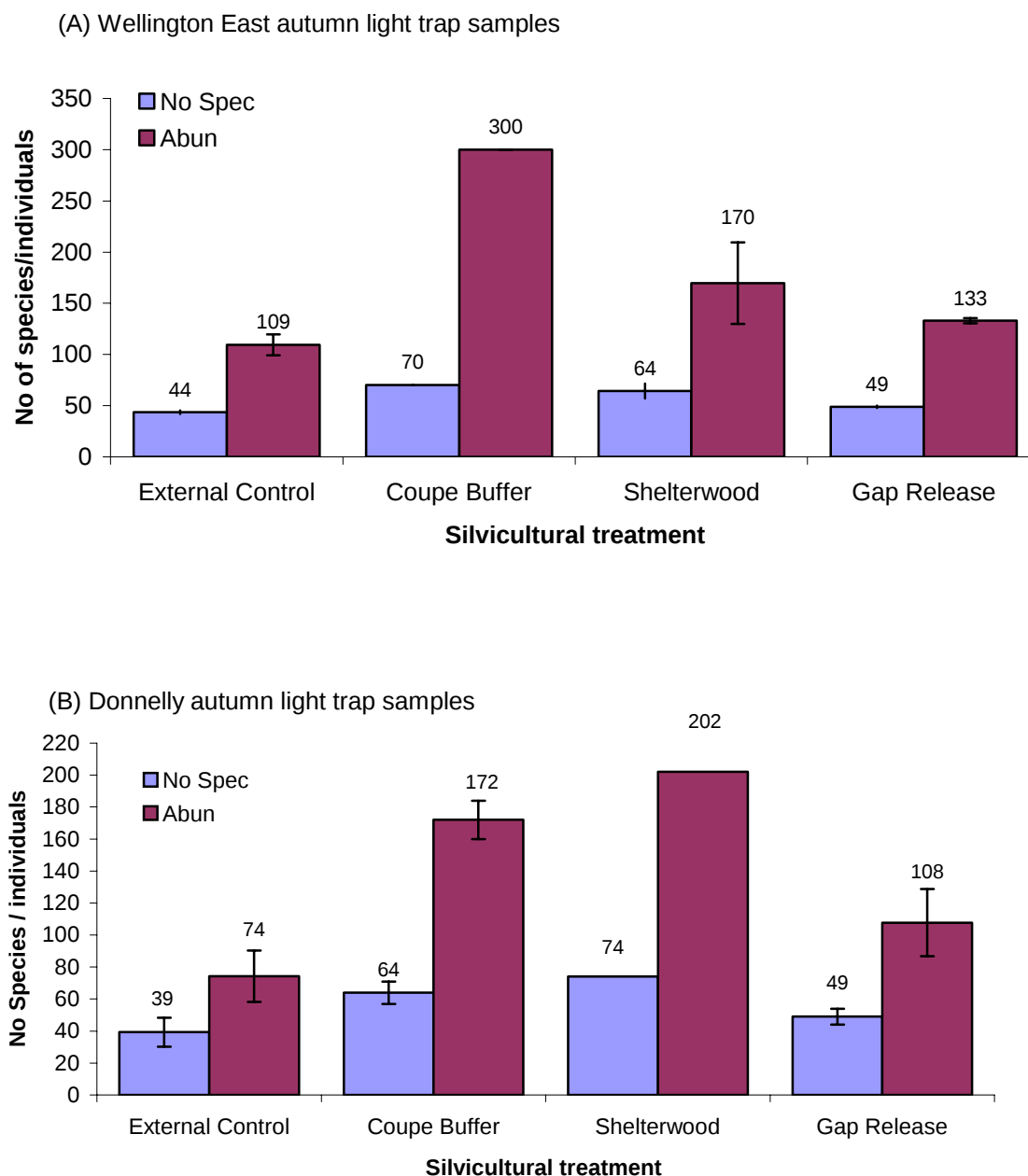


Figure 7. Mean ($n = 3$, $\pm$ SE) autumn light trap captures against treatment grids for (a) Wellington East and (b) Donnelly (note: Donnelly shelterwood is for $n=1$). (number of morphospecies = No Spec; number of individuals = abundance).

There were 137 morphospecies common to Perth Hills, Wellington and Donnelly (Fig. 8). Of the 960 total morphospecies, 202 were exclusive to Perth Hills (45% of the Perth Hills capture), 118 to Wellington (30% of the Wellington capture) and 299 were exclusive to Donnelly (51% of the Donnelly capture).

Table 4. Ten most frequent species captured for Perth Hills, Wellington and Donnelly (Capture frequency is the number of times a specimen is collected at its respective sample site and is not to be confused with abundance levels which refer to the total number of specimens collected; capture frequency ranks relate to Wellington East samples, eg. a rank of 6 for a Perth Hills specimen means this species was the 6th most frequent species at Wellington East, a rank >31 indicates single specimen only found in Wellington East, a rank = 0 indicates no specimen captured in Wellington East).

Sample Site	Species No.	Capture frequency	Wellington East capture frequency rank	Order	Family	Genus
Wellington East	436	132	1	Lepidoptera	Geometridae	
	14	49	2	Coleoptera	Hydrophilidae	
	4	45	3	Lepidoptera	Notodontidae	<i>Destolmia</i>
	16	40	4	Diptera	Tipulidae	
	145	38	5	Trichoptera		
	424	36	6	Lepidoptera	Geometridae	
	52	36	6	Hymenoptera	Apidae	<i>Apis</i>
	326	32	7	Lepidoptera	Geometridae	
	39	32	7	Lepidoptera	Noctuidae	
	423	30	8	Hymenoptera	Formicidae	<i>Camponotus</i>
Perth Hills	436	92	1	Lepidoptera	Geometridae	
	45	34	13	Lepidoptera	Zygaenidae	<i>Pollanisus</i>
	145	32	5	Trichoptera		
	14	26	2	Coleoptera	Hydrophilidae	
	144	23	9	Trichoptera		
	39	22	7	Lepidoptera	Noctuidae	
	424	21	6	Lepidoptera	Geometridae	
	52	20	6	Hymenoptera	Apidae	<i>Apis</i>
	235	20	18	Orthoptera	Acrididae	
	880	19	0	Scorpionida		
	634	19	30	Lepidoptera	Geometridae	
Wellington	436	84	1	Lepidoptera	Geometridae	
	52	72	6	Hymenoptera	Apidae	<i>Apis</i>
	145	52	5	Trichoptera		
	4	27	3	Lepidoptera	Notodontidae	<i>Destolmia</i>
	235	25	18	Orthoptera	Acrididae	
	258	22	14	Dermaptera		
	163	22	31	Hemiptera	Reduviidae	
	11	20	22	Lepidoptera	Thaumetopoeidae	<i>Ochrogaster</i>
	423	19	8	Hymenoptera	Formicidae	<i>Camponotus</i>
	1	19	21	Lepidoptera	Carthaeidae	<i>Carthaea</i>
	16	19	4	Diptera	Tipulidae	
Donnelly	52	64	6	Hymenoptera	Apidae	<i>Apis</i> (honey bee)
	6	54	30	Lepidoptera	Arctiidae	
	235	45	18	Orthoptera	Acrididae	
	373	28	0	Lepidoptera	Hepialidae	<i>Abantiades</i>
	39	28	7	Lepidoptera	Noctuidae	
	145	26	5	Trichoptera		
	18	26	13	Lepidoptera	Noctuidae	<i>Agrotis</i>
	45	26	13	Lepidoptera	Zygaenidae	
	376	26	0	Lepidoptera		
	423	24	8	Hymenoptera	Formicidae	<i>Iridomyrex</i>

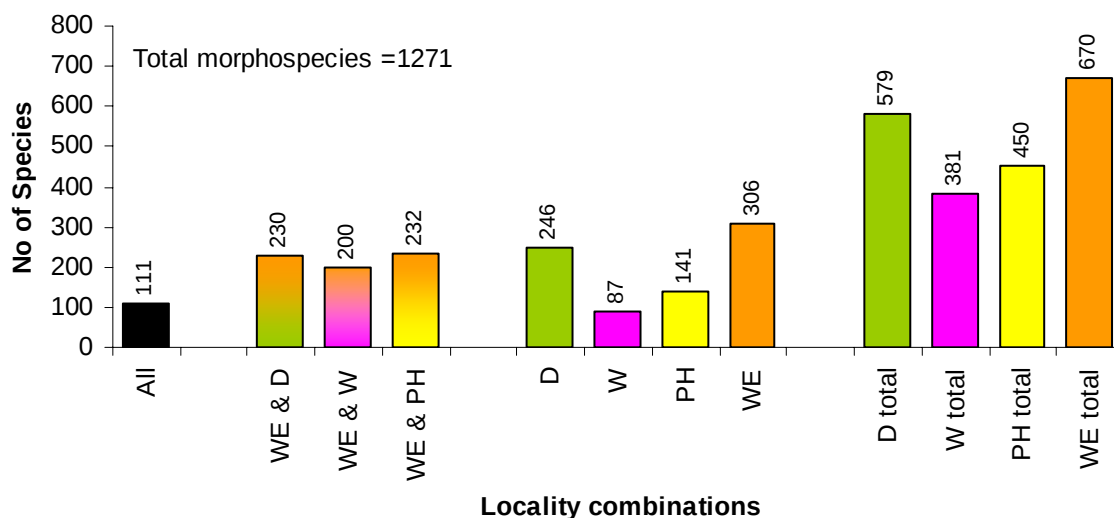


Figure 8. Number of species common and exclusive to Wellington East (WE), Perth Hills (PH), Wellington (W) and Donnelly (D). (All refers to number of species common to all localities; locality combinations such as WE & D refers to the number of species common to both; a single locality shows the number of species exclusive to that locality; location totals refer to total morphospecies for the respective locality and is included for comparison)

Pest presence

Gumleaf skeletonizer was absent from all Wellington east grids (Table 5). Bullseye borer was present at all sites and Jarrah leafminer was present at all grids, with high populations at the Nalyerin external control (FC28) and Godfrey shelterwood (FC30).

Table 5. Pest presence and abundance assessment at each site (JLM = jarrah leafminer; GLS = gumleaf skeletonizer; BEB = bullseye borer; 0 = absent, 1 = present, 2 = abundant).

Treatment	Site No	Location	JLM	GLS	BEB
External Control	FC28	Nalyerin	2	0	1
External Control	FC32	Stockyard	1	0	1
External Control	FC35	Bell	1	0	1
Coupe Buffer	FC29	Godfrey	1	0	1
Shelterwood	FC30	Godfrey	2	0	1
Shelterwood	FC33	Stockyard	1	0	1
Shelterwood	FC36	Bell	1	0	1
Gap Release	FC31	Godfrey	1	0	1
Gap Release	FC34	Stockyard	1	0	1
Gap Release	FC37	Bell	1	0	1

Conclusions

Species composition, richness and abundance

- After 4 years of collection, 1 271 morphospecies of macroinvertebrates (>10 mm length) have been collected. After an initial collection of 582 morphospecies in 2001, about 200 new morphospecies have been collected in 2002 and in 2003. Remarkably, 306 new species were collected from Wellington East indicating high invertebrate diversity in the eastern part of the Jarrah forest.
- Light traps from Wellington East were unusually productive in spring compared to collections in previous years. Thus greater diversity of Lepidoptera, the principal catch of light traps, was collected from Wellington East than from previous years.
- Wellington East also had greatest species diversity for the Araneomorpha (spiders), Dermaptera (earwigs) and Orthoptera (crickets and grasshoppers).

Effects of Silvicultural treatments

- Silvicultural treatment had no significant effect on overall (combined collection methods) macroinvertebrate abundance or morphospecies diversity at Wellington East.
- Further analysis is required to determine interaction between collection method and silvicultural effects.
- Further analysis is required to determine effect of silvicultural treatment on individual species.

Presence of pest species

- Jarrah leafminer was present on all sites sampled and abundant at 2 grids (FC28, Nalyerin external control and FC30, Godfrey shelterwood).
- Gumleaf skeletonizer was not detected on any site.
- Bullseye borer was present on all sites.

Appendix 1. Morphospecies list for FORESTCHECK and Walpole Fire Fine Grain Mosaic invertebrates for July 2005. Data sorted on Taxa (K = assigned indicator species; GA = species with Gondwanan affinities; GR = suspected Gondwanan relic species)

Spec #	Order	Family	Tax 3	Genus	Species	GR/K
1303	Amphipoda					GR
261	Amphipoda		Not found			GR
472	Araneomorphae		Degraded			
1221	Araneomorphae	Araneidae				
1746	Araneomorphae	Araneidae				
1726	Araneomorphae	Araneidae		<i>Arachnura</i>	<i>higginsii</i> ?	
1684	Araneomorphae	Araneidae		<i>Argiope</i>		
1206	Araneomorphae	Araneidae		<i>Eriophora</i>		
1319	Araneomorphae	Araneidae		<i>Eriophora</i>		
1680	Araneomorphae	Araneidae		<i>Eriophora</i>		K
1217	Araneomorphae	Araneidae		<i>Eriophora</i>	<i>transmarina</i> ?	K
1710	Araneomorphae	Araneidae		<i>Gasteracantha</i>		
1213	Araneomorphae	Araneidae		<i>Gasteracantha</i>	<i>minax</i> ?	
1551	Araneomorphae	Araneidae		<i>Nephila</i>	<i>edulis</i>	
1671	Araneomorphae	Araneidae		<i>Phonographa</i>	<i>graeffei</i> ?	K
1471	Araneomorphae	Araneidae		<i>Phonographa</i>	<i>graeffei</i>	K

Spec #	Order	Family	Tax 3	Genus	Species	GR/K
285	Araneomorphae	Araneidae		<i>Eriophora</i>		
536	Araneomorphae	Corinnidae		<i>Supunna</i>	<i>albopunctata</i>	K
537	Araneomorphae	Corinnidae		<i>Supunna</i>	<i>picta</i>	K
553	Araneomorphae	Ctenidae				
1682	Araneomorphae	Deinopidae		<i>Deinopis</i>		
1544	Araneomorphae	Deinopidae		<i>Deinopis ?</i>		K
941	Araneomorphae	Gnaphosidae				
975	Araneomorphae	Gnaphosidae				
1377	Araneomorphae	Gnaphosidae				
1413	Araneomorphae	Gnaphosidae				
1778	Araneomorphae	Gnaphosidae				
560	Araneomorphae	Gnaphosidae				
938	Araneomorphae	Gnaphosidae		<i>Rebilus</i>		
1427	Araneomorphae	Gnaphosidae		<i>Rebilus</i>		
1809	Araneomorphae	Gnaphosidae		<i>Rebilus ?</i>		
742	Araneomorphae	Gnaphosidae				
620	Araneomorphae	Gnaphosidae		<i>Rebilus</i>		
1793	Araneomorphae	Linophiidae				
561	Araneomorphae	Lycosidae				
1294	Araneomorphae	Lycosidae				
1373	Araneomorphae	Lycosidae				
1558	Araneomorphae	Lycosidae				
1581	Araneomorphae	Lycosidae				
1588	Araneomorphae	Lycosidae				
1589	Araneomorphae	Lycosidae				
1593	Araneomorphae	Lycosidae				
1595	Araneomorphae	Lycosidae				
1786	Araneomorphae	Lycosidae				
554	Araneomorphae	Lycosidae				
733	Araneomorphae	Lycosidae				
741	Araneomorphae	Lycosidae				
743	Araneomorphae	Lycosidae				
740	Araneomorphae	Miturgidae				
933	Araneomorphae	Miturgidae				
1428	Araneomorphae	Miturgidae				
1448	Araneomorphae	Miturgidae				
1449	Araneomorphae	Miturgidae				K
1476	Araneomorphae	Miturgidae				
1477	Araneomorphae	Miturgidae				
1564	Araneomorphae	Miturgidae				
1574	Araneomorphae	Miturgidae				
1579	Araneomorphae	Miturgidae				
1580	Araneomorphae	Miturgidae				
1737	Araneomorphae	Miturgidae				
597	Araneomorphae	Miturgidae				
271	Araneomorphae	Miturgidae				
740	Araneomorphae	Miturgidae				
794	Araneomorphae	Miturgidae				
812	Araneomorphae	Miturgidae				
1044	Araneomorphae	Sparassidae				
1282	Araneomorphae	Sparassidae				
1432	Araneomorphae	Sparassidae				
1446	Araneomorphae	Sparassidae				
286	Araneomorphae	Sparassidae				K

Spec #	Order	Family	Tax 3	Genus	Species	GR/K
939	Araneomorphae	Sparassidae		<i>Olios</i>		
1250	Araneomorphae	Sparassidae		<i>Olios</i>	<i>diana ?</i>	
724	Araneomorphae	Sparassidae				
1403	Araneomorphae	Stiphidiidae				
1570	Araneomorphae	Stiphidiidae				
1609	Araneomorphae	Stiphidiidae				
732	Araneomorphae	Stiphidiidae		<i>Balami</i>		
735	Araneomorphae	Stiphidiidae		<i>Balami</i>	<i>volucripes</i>	
725	Araneomorphae	Stiphidiidae		<i>Balami ?</i>	<i>volucripes</i>	
793	Araneomorphae	Theridiidae				
788	Araneomorphae	Theridiidae ?				
932	Araneomorphae	Zodariidae				
972	Araneomorphae	Zodariidae				
1007	Araneomorphae	Zodariidae				
1015	Araneomorphae	Zodariidae				
1244	Araneomorphae	Zodariidae				
1584	Araneomorphae	Zodariidae				
1610	Araneomorphae	Zodariidae				
468	Araneomorphae	Zodariidae		<i>Storena</i>		
731	Araneomorphae	Zodariidae				
783	Araneomorphae	Zodariidae				
468	Araneomorphae	Zodariidae		<i>Storena</i>		K
1115	Blattodea					
1118	Blattodea					
1462	Blattodea		not found			
269	Blattodea					
509	Blattodea		not found			
926	Blattodea	Blaberidae				
1101	Blattodea	Blaberidae				
570	Blattodea	Blaberidae				K
1578	Blattodea	Blaberidae		<i>Laxta ?</i>		
148	Blattodea	Blaberidae				
410	Blattodea	Blaberidae				K
479	Blattodea	Blaberidae				
483	Blattodea	Blaberidae				
27	Blattodea	Blaberidae		<i>Calolampra</i>		K
119	Blattodea	Blaberidae		<i>Calolampra</i>		K
147	Blattodea	Blaberidae		<i>Calolampra</i>		
292	Blattodea	Blaberidae		<i>Laxta</i>		K
483	Blattodea	Blaberidae		<i>Laxta</i>		K
781	Blattodea	Blaberidae		<i>Laxta</i>		K
120	Blattodea	Blattellidae		<i>Neotemnapteryx</i>		K
591	Blattodea	Blattellidae		<i>Neotemnapteryx</i>		
121	Blattodea	Blattellidae		<i>Platyzosteria</i>		K
122	Blattodea	Blattellidae		<i>Platyzosteria</i>		K
874	Blattodea	Blattidae				
878	Blattodea	Blattidae				
891	Blattodea	Blattidae				
899	Blattodea	Blattidae				
905	Blattodea	Blattidae				
936	Blattodea	Blattidae				
961	Blattodea	Blattidae				
967	Blattodea	Blattidae				
968	Blattodea	Blattidae				

Spec #	Order	Family	Tax 3	Genus	Species	GR/K
971	Blattodea	Blattidae				
1016	Blattodea	Blattidae				
1035	Blattodea	Blattidae				
1247	Blattodea	Blattidae				
1313	Blattodea	Blattidae				
1434	Blattodea	Blattidae				
1460	Blattodea	Blattidae				K
1474	Blattodea	Blattidae				
1555	Blattodea	Blattidae				
1559	Blattodea	Blattidae				
1573	Blattodea	Blattidae				
1587	Blattodea	Blattidae				
1741	Blattodea	Blattidae				
1766	Blattodea	Blattidae				
1773	Blattodea	Blattidae				
1780	Blattodea	Blattidae				
525	Blattodea	Blattidae				K
190	Blattodea	Blattidae				K
490	Blattodea	Blattidae				
507	Blattodea	Blattidae				
508	Blattodea	Blattidae				K
706	Blattodea	Blattidae				
254	Blattodea	Blattidae		<i>Platyzosteria</i>		
282	Blattodea	Blattidae		<i>Platyzosteria</i>		K
219	Blattodea	Blattidae		<i>Platyzosteria</i>		K
266	Blattodea	Blattidae		<i>Platyzosteria</i>		K
592	Blattodea	Blattidae		<i>Polyzosteria</i>		
777	Blattodea	Blattidae	Michells cocky	<i>Polyzosteria</i>	<i>mitchelli</i>	K
875	Chilopoda					
877	Chilopoda					
1200	Chilopoda					
1429	Chilopoda					
1531	Chilopoda					
1563	Chilopoda					
1767	Chilopoda					
223	Chilopoda					
224	Chilopoda					
225	Chilopoda					
226	Chilopoda					
227	Chilopoda					
228	Chilopoda					
229	Chilopoda					
267	Chilopoda					
277	Chilopoda					
623	Chilopoda					
716	Chilopoda					
815	Chilopoda					
1768	Chilopoda	Scolopendridae				
1783	Chilopoda	Scolopendridae				
1583	Chilopoda	Scolopendridae ?				
865	Coleoptera					
1050	Coleoptera					
1183	Coleoptera					
1187	Coleoptera					

Spec #	Order	Family	Tax 3	Genus	Species	GR/K
1208	Coleoptera					
1238	Coleoptera					
1242	Coleoptera					
1243	Coleoptera					
1245	Coleoptera					
1248	Coleoptera					
1290	Coleoptera					
1295	Coleoptera					
1307	Coleoptera					
116	Coleoptera	?				
158	Coleoptera	?				
159	Coleoptera	?				
173	Coleoptera	?				
194	Coleoptera	?				
215	Coleoptera	?				
1298	Coleoptera	Belidae				GR
100	Coleoptera	Belidae				GR
201	Coleoptera	Belidae		<i>Araiobelus</i>		GR
168	Coleoptera	Belidae		<i>Rhinotia</i>		GR
1300	Coleoptera	Buprestidae				K
1309	Coleoptera	Buprestidae				K
1435	Coleoptera	Buprestidae				K
1437	Coleoptera	Buprestidae				K
1443	Coleoptera	Buprestidae				K
1464	Coleoptera	Buprestidae				K
1708	Coleoptera	Buprestidae				K
299	Coleoptera	Buprestidae				K
701	Coleoptera	Buprestidae		<i>Melobasis</i>		K
1724	Coleoptera	Cantharidae		<i>Chauliognathus</i>		
198	Coleoptera	Cantharidae		<i>Heteromastix</i>		
851	Coleoptera	Carabidae				GA
992	Coleoptera	Carabidae				GA
1000	Coleoptera	Carabidae				GA
1059	Coleoptera	Carabidae				GA
1087	Coleoptera	Carabidae				GA
1100	Coleoptera	Carabidae				GA
1104	Coleoptera	Carabidae				GA
1107	Coleoptera	Carabidae				GA
1306	Coleoptera	Carabidae				GA
1347	Coleoptera	Carabidae				GA
1363	Coleoptera	Carabidae				GA
1364	Coleoptera	Carabidae				GA
1366	Coleoptera	Carabidae				GA
1387	Coleoptera	Carabidae				GA
1442	Coleoptera	Carabidae				GA
1522	Coleoptera	Carabidae				GA
1605	Coleoptera	Carabidae				GA
1642	Coleoptera	Carabidae				GA
1785	Coleoptera	Carabidae				
1791	Coleoptera	Carabidae				
1058	Coleoptera	Carabidae		<i>Calosoma</i>	<i>schayeri</i>	K
956	Coleoptera	Carabidae		<i>Philophloeus</i>	<i>eucalypti</i>	GA
93	Coleoptera	Carabidae				GA
253	Coleoptera	Carabidae				GA

Spec #	Order	Family	Tax 3	Genus	Species	GR/K
528	Coleoptera	Carabidae				K
529	Coleoptera	Carabidae				K
727	Coleoptera	Carabidae				GA
746	Coleoptera	Carabidae				GA
747	Coleoptera	Carabidae		<i>?Notonomus</i>		GA
280	Coleoptera	Carabidae		<i>Carenum</i>		GA
288	Coleoptera	Carabidae		<i>Chlaenius</i>		GA
557	Coleoptera	Carabidae				K
587	Coleoptera	Carabidae				K
839	Coleoptera	Carabidae				GA
842	Coleoptera	Carabidae				GA
439	Coleoptera	Carabidae	Carabinae			GA
340	Coleoptera	Carabidae	Chlaeniinae			GA
265	Coleoptera	Carabidae	Esydrinae			GA
264	Coleoptera	Carabidae	Harpalinae	<i>?Cenogmus</i>		GA
914	Coleoptera	Carabidae	Licininae	<i>Dicrochile</i>		GA
558	Coleoptera	Carabidae	Pentagonicinae	<i>Scapodes</i>	<i>boops</i>	GA
1033	Coleoptera	Cerambycidae				
1040	Coleoptera	Cerambycidae				
1041	Coleoptera	Cerambycidae				
1067	Coleoptera	Cerambycidae				
1082	Coleoptera	Cerambycidae				
1177	Coleoptera	Cerambycidae				
1283	Coleoptera	Cerambycidae				
1334	Coleoptera	Cerambycidae				
1418	Coleoptera	Cerambycidae				
1417	Coleoptera	Cerambycidae		<i>Sceleocantha ?</i>		
654	Coleoptera	Cerambycidae		<i>Coptocercus</i>	<i>rubripes</i>	
762	Coleoptera	Cerambycidae		<i>Phoracantha</i>	<i>semipunctata</i>	
673	Coleoptera	Cerambycidae		<i>Stenoderus</i>	<i>suturalis</i>	
351	Coleoptera	Cerambycidae		<i>Uracantha</i>	<i>triangularis</i>	K
476	Coleoptera	Cerambycidae	Laminae			
182	Coleoptera	Chrysomelidae	Chrysomelinae			
56	Coleoptera	Chrysomelidae	Chrysomelinae	<i>Calomela</i>		
101	Coleoptera	Chrysomelidae	Chrysomelinae	<i>Calomela</i>		
115	Coleoptera	Chrysomelidae	Chrysomelinae	<i>Calomela</i>		
155	Coleoptera	Chrysomelidae	Chrysomelinae	<i>Calomela</i>		
807	Coleoptera	Chrysomelidae	Chrysomelinae	<i>Chalcolampra</i>		K
112	Coleoptera	Chrysomelidae	Chrysomelinae	<i>Chrysophtharta</i>		
308	Coleoptera	Chrysomelidae	Chrysomelinae	<i>Chrysophtharta</i>		K
463	Coleoptera	Chrysomelidae	Chrysomelinae	<i>Chrysophtharta</i>		K
677	Coleoptera	Chrysomelidae	Chrysomelinae	<i>Chrysophtharta</i>		
786	Coleoptera	Chrysomelidae	Chrysomelinae	<i>Chrysophtharta</i>		
800	Coleoptera	Chrysomelidae	Chrysomelinae	<i>Chrysophtharta</i>		
803	Coleoptera	Chrysomelidae	Chrysomelinae	<i>Chrysophtharta</i>		
804	Coleoptera	Chrysomelidae	Chrysomelinae	<i>Chrysophtharta</i>		
805	Coleoptera	Chrysomelidae	Chrysomelinae	<i>Chrysophtharta</i>		
1205	Coleoptera	Chrysomelidae	Chrysomelinae	<i>Chrysophtharta</i>		
1254	Coleoptera	Chrysomelidae	Chrysomelinae	<i>Chrysophtharta</i>		
1263	Coleoptera	Chrysomelidae	Chrysomelinae	<i>Chrysophtharta</i>		
1279	Coleoptera	Chrysomelidae	Chrysomelinae	<i>Chrysophtharta</i>		
1322	Coleoptera	Chrysomelidae	Chrysomelinae	<i>Chrysophtharta</i>		
1329	Coleoptera	Chrysomelidae	Chrysomelinae	<i>Chrysophtharta</i>		
1444	Coleoptera	Chrysomelidae	Chrysomelinae	<i>Chrysophtharta</i>		

Spec #	Order	Family	Tax 3	Genus	Species	GR/K
1540	Coleoptera	Chrysomelidae	Chrysomelinae	<i>Chrysophtharta</i>		
1669	Coleoptera	Chrysomelidae	Chrysomelinae	<i>Chrysophtharta</i>		
1745	Coleoptera	Chrysomelidae	Chrysomelinae	<i>Chrysophtharta</i>		
1752	Coleoptera	Chrysomelidae	Chrysomelinae	<i>Chrysophtharta</i>		
1772	Coleoptera	Chrysomelidae	Chrysomelinae	<i>Chrysophtharta</i>		
175	Coleoptera	Chrysomelidae	Chrysomelinae	<i>Chrysophtharta</i>		
248	Coleoptera	Chrysomelidae	Chrysomelinae	<i>Chrysophtharta</i>		
471	Coleoptera	Chrysomelidae	Chrysomelinae	<i>Chrysophtharta</i>		
307	Coleoptera	Chrysomelidae	Chrysomelinae	<i>Paropsis</i>		K
665	Coleoptera	Chrysomelidae	Chrysomelinae	<i>Paropsis</i>		
667	Coleoptera	Chrysomelidae	Chrysomelinae	<i>Paropsis</i>		K
707	Coleoptera	Chrysomelidae	Chrysomelinae	<i>Paropsis</i>		
913	Coleoptera	Chrysomelidae	Chrysomelinae	<i>Paropsis</i>		
1554	Coleoptera	Chrysomelidae	Chrysomelinae	<i>Paropsis</i>		
1825	Coleoptera	Chrysomelidae	Chrysomelinae	<i>Paropsis</i>		
1826	Coleoptera	Chrysomelidae	Chrysomelinae	<i>Paropsis</i>		
1827	Coleoptera	Chrysomelidae	Chrysomelinae	<i>Paropsis</i>		
665	Coleoptera	Chrysomelidae	Chrysomelinae	<i>Paropsis</i>	<i>yilgarnensis ?</i>	K
1092	Coleoptera	Chrysomelidae	Chrysomelinae	<i>Paropsisterna</i>		
1253	Coleoptera	Chrysomelidae	Cryptocephalinae	<i>Cadmus</i>	<i>excrementarius</i>	
1103	Coleoptera	Cleridae				
684	Coleoptera	Cleridae				
695	Coleoptera	Cleridae		<i>Eleale</i>		
1310	Coleoptera	Coccinellidae				
912	Coleoptera	Coccinellidae		<i>Paraprius</i>		
193	Coleoptera	Coccinellidae		<i>Coccinella</i>	<i>repanda</i>	
852	Coleoptera	Curculionidae				
911	Coleoptera	Curculionidae				
993	Coleoptera	Curculionidae				
1110	Coleoptera	Curculionidae				
1175	Coleoptera	Curculionidae				
1182	Coleoptera	Curculionidae				
1216	Coleoptera	Curculionidae				
1225	Coleoptera	Curculionidae				
1270	Coleoptera	Curculionidae				
1292	Coleoptera	Curculionidae				
1304	Coleoptera	Curculionidae				
1331	Coleoptera	Curculionidae				
1414	Coleoptera	Curculionidae				
1438	Coleoptera	Curculionidae				
1505	Coleoptera	Curculionidae				
1716	Coleoptera	Curculionidae				
368	Coleoptera	Curculionidae				K
1219	Coleoptera	Curculionidae		<i>Catasarcus</i>		
898	Coleoptera	Curculionidae		<i>Oxyops</i>		
1278	Coleoptera	Curculionidae		<i>Oxyops</i>		
102	Coleoptera	Curculionidae				
114	Coleoptera	Curculionidae				
156	Coleoptera	Curculionidae				
169	Coleoptera	Curculionidae				
199	Coleoptera	Curculionidae				
209	Coleoptera	Curculionidae				K
214	Coleoptera	Curculionidae				
244	Coleoptera	Curculionidae				K

Spec #	Order	Family	Tax 3	Genus	Species	GR/K
349	Coleoptera	Curculionidae				K
514	Coleoptera	Curculionidae				
709	Coleoptera	Curculionidae				
736	Coleoptera	Curculionidae				
702	Coleoptera	Curculionidae		<i>Gonipterus</i>		
841	Coleoptera	Curculionidae				
843	Coleoptera	Curculionidae				
906	Coleoptera	Curculionidae	Amycterinae			GR
910	Coleoptera	Curculionidae	Amycterinae			GR
934	Coleoptera	Curculionidae	Amycterinae			GR
1014	Coleoptera	Curculionidae	Amycterinae			GR
1251	Coleoptera	Curculionidae	Amycterinae			GR
1318	Coleoptera	Curculionidae	Amycterinae			GR
1352	Coleoptera	Curculionidae	Amycterinae			GR
1361	Coleoptera	Curculionidae	Amycterinae			GR
1396	Coleoptera	Curculionidae	Amycterinae			GR
1397	Coleoptera	Curculionidae	Amycterinae			GR
1409	Coleoptera	Curculionidae	Amycterinae			GR
1461	Coleoptera	Curculionidae	Amycterinae			GR
1486	Coleoptera	Curculionidae	Amycterinae			GR
1543	Coleoptera	Curculionidae	Amycterinae			GR
1596	Coleoptera	Curculionidae	Amycterinae			GR
1597	Coleoptera	Curculionidae	Amycterinae			GR
1624	Coleoptera	Curculionidae	Amycterinae			GR
1672	Coleoptera	Curculionidae	Amycterinae			GR
300	Coleoptera	Curculionidae	Amycterinae			GR
744	Coleoptera	Curculionidae	Amycterinae			GR
748	Coleoptera	Curculionidae	Amycterinae			GR
814	Coleoptera	Curculionidae	Amycterinae			GR
817	Coleoptera	Curculionidae	Amycterinae			GR
496	Coleoptera	Curculionidae	Amycterinae	<i>Acantholophus</i>		GR
869	Coleoptera	Curculionidae	Amycterinae	<i>Acantholophus</i>		GR
970	Coleoptera	Curculionidae	Amycterinae	<i>Acantholophus</i>		GR
1215	Coleoptera	Curculionidae	Amycterinae	<i>Acantholophus</i>		GR
1262	Coleoptera	Curculionidae	Amycterinae	<i>Acantholophus</i>		GR
1523	Coleoptera	Curculionidae	Amycterinae	<i>Acantholophus</i>		GR
1774	Coleoptera	Curculionidae	Amycterinae	<i>Acantholophus</i>		GR
1775	Coleoptera	Curculionidae	Amycterinae	<i>Acantholophus</i>		GR
1803	Coleoptera	Curculionidae	Amycterinae	<i>Acantholophus</i>		GR
1571	Coleoptera	Curculionidae	Amycterinae	<i>Cucullothorax</i>	<i>horridus</i>	GR
1764	Coleoptera	Curculionidae	Amycterinae	<i>Euomus</i>		GR
1747	Coleoptera	Curculionidae	Amycterinae	<i>Euomus</i>	<i>sp nova</i>	GR
1376	Coleoptera	Curculionidae	Amycterinae	<i>Sclerorinus</i>		GR
1763	Coleoptera	Curculionidae	Amycterinae	<i>Sclerorinus</i>		GR
1810	Coleoptera	Curculionidae	Amycterinae	<i>Sclerorinus</i>		GR
1789	Coleoptera	Curculionidae	Amycterinae	<i>Sclerorinus ?</i>		GR
1801	Coleoptera	Curculionidae	Amycterinae	<i>Sclerorinus ?</i>		GR
157	Coleoptera	Curculionidae	Aterpinae	<i>Rhadinosomus</i>	<i>lacordairei</i>	K
103	Coleoptera	Curculionidae	Aterpinae	<i>Rhinaria</i>	<i>aberrans (?)</i>	
710	Coleoptera	Curculionidae	Entiminae			
210	Coleoptera	Curculionidae	Entiminae	<i>Aesolithna</i>		
113	Coleoptera	Curculionidae	Entiminae	<i>Polyphrades</i>	<i>aesalon (?)</i>	
160	Coleoptera	Curculionidae	Gonipterinae	<i>Gonipterus</i>		
488	Coleoptera	Curculionidae	Gonipterinae	<i>Gonipterus</i>		

Spec #	Order	Family	Tax 3	Genus	Species	GR/K
462	Coleoptera	Curculionidae	Gonipterinae	<i>Oxyops</i>		
470	Coleoptera	Curculionidae	Gonipterinae	<i>Oxyops</i>		
161	Coleoptera	Curculionidae	Gonipterinae	<i>Oxyops</i>	<i>fasciata</i>	K
98	Coleoptera	Curculionidae	Gonipterinae	<i>Oxyops</i>	<i>pictipennis</i>	
290	Coleoptera	Curculionidae	Molytinae	<i>Helanotranes</i>	<i>roei</i>	K
291	Coleoptera	Curculionidae	Molytinae	<i>Tranes</i>	<i>vigorsii</i>	K
1412	Coleoptera	Dytiscidae				
1123	Coleoptera	Dytiscidae		<i>Cybister</i>		K
850	Coleoptera	Dytiscidae		<i>Lancetes</i>		
13	Coleoptera	Dytiscidae				
440	Coleoptera	Dytiscidae				
774	Coleoptera	Dytiscidae		<i>Eretes</i>		
651	Coleoptera	Dytiscidae		<i>Lancetes</i>		
989	Coleoptera	Elateridae				K
1083	Coleoptera	Elateridae				
1817	Coleoptera	Elateridae				
909	Coleoptera	Elateridae		<i>Conoderus</i>		
1816	Coleoptera	Elateridae		<i>Conoderus</i>		
1818	Coleoptera	Elateridae		<i>Conoderus</i>		
1819	Coleoptera	Elateridae		<i>Conoderus</i>		
997	Coleoptera	Elateridae		<i>Conoderus ?</i>		
1062	Coleoptera	Elateridae		<i>Conoderus ?</i>		
1109	Coleoptera	Elateridae		<i>Conoderus ?</i>		
1120	Coleoptera	Elateridae		<i>Conoderus ?</i>		
1121	Coleoptera	Elateridae		<i>Conoderus ?</i>		
15	Coleoptera	Elateridae				
220	Coleoptera	Elateridae				
444	Coleoptera	Elateridae				
571	Coleoptera	Elateridae				
621	Coleoptera	Elateridae				
636	Coleoptera	Elateridae				
26	Coleoptera	Elateridae		<i>Conoderus</i>		
135	Coleoptera	Elateridae		<i>Conoderus ?</i>		
831	Coleoptera	Geotrupidae		<i>Blackbolbus</i>		K
959	Coleoptera	Histeridae				
14	Coleoptera	Hydrophilidae				
1338	Coleoptera	Lucanidae				
1629	Coleoptera	Lucanidae				
1640	Coleoptera	Lucanidae				
1797	Coleoptera	Lucanidae				
437	Coleoptera	Lucanidae		<i>Syndesus</i>		K
795	Coleoptera	Lycidae				
99	Coleoptera	Lycidae		<i>Metriorrhynchus</i>		K
208	Coleoptera	Lycidae		<i>Metriorrhynchus</i>		K
802	Coleoptera	Lycidae		<i>Metriorrhynchus</i>		K
191	Coleoptera	Phycosecidae	Phycosecis			
824	Coleoptera	Scarabaeidae	Dynastinae			
189	Coleoptera	Scarabaeidae	Dynastinae	<i>Cryptodus</i>		K
1160	Coleoptera	Scarabaeidae	Dynastinae	<i>Cryptodus</i>		
824	Coleoptera	Scarabaeidae	Dynastinae	<i>Semanopterus</i>		
1562	Coleoptera	Scarabaeidae	Dynastinae	<i>Trissodon</i>		
1133	Coleoptera	Scarabaeidae	Melolonthinae			
550	Coleoptera	Scarabaeidae	Melolonthinae			
212	Coleoptera	Scarabaeidae	Melolonthinae	<i>Automolus ?</i>		

Spec #	Order	Family	Tax 3	Genus	Species	GR/K
1823	Coleoptera	Scarabaeidae	Melolonthinae	<i>Colpochila</i>		
1138	Coleoptera	Scarabaeidae	Melolonthinae	<i>Colpochila</i>	<i>andersoni</i>	
846	Coleoptera	Scarabaeidae	Melolonthinae	<i>Colpochila</i>	<i>antennalis</i> ?	K
1063	Coleoptera	Scarabaeidae	Melolonthinae	<i>Colpochila</i>	<i>bogaria</i> ?	
353	Coleoptera	Scarabaeidae	Melolonthinae	<i>Colpochila</i>	<i>major</i>	K
55	Coleoptera	Scarabaeidae	Melolonthinae	<i>Colymbomorpha</i>	<i>vittata</i>	
28	Coleoptera	Scarabaeidae	Melolonthinae	<i>Heteronyx</i>		
29	Coleoptera	Scarabaeidae	Melolonthinae	<i>Heteronyx</i>		
70	Coleoptera	Scarabaeidae	Melolonthinae	<i>Heteronyx</i>		
94	Coleoptera	Scarabaeidae	Melolonthinae	<i>Heteronyx</i>		
154	Coleoptera	Scarabaeidae	Melolonthinae	<i>Heteronyx</i>		
171	Coleoptera	Scarabaeidae	Melolonthinae	<i>Heteronyx</i>		
172	Coleoptera	Scarabaeidae	Melolonthinae	<i>Heteronyx</i>		
289	Coleoptera	Scarabaeidae	Melolonthinae	<i>Heteronyx</i>		
363	Coleoptera	Scarabaeidae	Melolonthinae	<i>Heteronyx</i>		
562	Coleoptera	Scarabaeidae	Melolonthinae	<i>Heteronyx</i>		
823	Coleoptera	Scarabaeidae	Melolonthinae	<i>Heteronyx</i>		
951	Coleoptera	Scarabaeidae	Melolonthinae	<i>Heteronyx</i>		
991	Coleoptera	Scarabaeidae	Melolonthinae	<i>Heteronyx</i>		
1073	Coleoptera	Scarabaeidae	Melolonthinae	<i>Heteronyx</i>		
1116	Coleoptera	Scarabaeidae	Melolonthinae	<i>Heteronyx</i>		
1133	Coleoptera	Scarabaeidae	Melolonthinae	<i>Heteronyx</i>		
1154	Coleoptera	Scarabaeidae	Melolonthinae	<i>Heteronyx</i>		
1192	Coleoptera	Scarabaeidae	Melolonthinae	<i>Heteronyx</i>		
1566	Coleoptera	Scarabaeidae	Melolonthinae	<i>Heteronyx</i>		
1612	Coleoptera	Scarabaeidae	Melolonthinae	<i>Heteronyx</i>		
1660	Coleoptera	Scarabaeidae	Melolonthinae	<i>Heteronyx</i>		
1820	Coleoptera	Scarabaeidae	Melolonthinae	<i>Heteronyx</i>		
1821	Coleoptera	Scarabaeidae	Melolonthinae	<i>Heteronyx</i>		
1822	Coleoptera	Scarabaeidae	Melolonthinae	<i>Heteronyx</i>		
347	Coleoptera	Scarabaeidae	Melolonthinae	<i>Heteronyx</i>		
359	Coleoptera	Scarabaeidae	Melolonthinae	<i>Heteronyx</i>		
363	Coleoptera	Scarabaeidae	Melolonthinae	<i>Heteronyx</i>		
427	Coleoptera	Scarabaeidae	Melolonthinae	<i>Heteronyx</i>		
162	Coleoptera	Scarabaeidae	Melolonthinae	<i>Liparetrus</i>	<i>jenkinsi</i>	
287	Coleoptera	Scarabaeidae	Melolonthinae	<i>Maechidius</i>		
287	Coleoptera	Scarabaeidae	Melolonthinae	<i>Maechidius</i>		
1388	Coleoptera	Scarabaeidae	Melolonthinae	<i>Maechidius</i> ?		
668	Coleoptera	Scarabaeidae	Melolonthinae	<i>Phyllotocus</i>	<i>ustulatus</i>	
1813	Coleoptera	Scarabaeidae	Melolonthinae	<i>Scitalini</i>		
1405	Coleoptera	Scarabaeidae	Scarabaeinae	<i>Onthophagus</i>		
1608	Coleoptera	Scarabaeidae	Scarabaeinae	<i>Onthophagus</i>		
1824	Coleoptera	Scarabaeidae	Scarabaeinae	<i>Onthophagus</i>		
511	Coleoptera	Scarabaeidae	Scarabaeinae	<i>Onthophagus</i>		
17	Coleoptera	Scarabaeidae	Scarabaeinae	<i>Onthophagus</i>	<i>ferox</i>	K
1656	Coleoptera	Silphidae				
924	Coleoptera	Silphidae		<i>Ptomophila</i>	<i>lacrymosa</i>	GR
628	Coleoptera	Staphylinidae				K
904	Coleoptera	Tenebrionidae				GA
995	Coleoptera	Tenebrionidae				GA
1076	Coleoptera	Tenebrionidae				GA
1117	Coleoptera	Tenebrionidae				GA
1246	Coleoptera	Tenebrionidae				GA
1392	Coleoptera	Tenebrionidae				GA

Spec #	Order	Family	Tax 3	Genus	Species	GR/K
1400	Coleoptera	Tenebrionidae				GA
1536	Coleoptera	Tenebrionidae				GA
1614	Coleoptera	Tenebrionidae				GA
1717	Coleoptera	Tenebrionidae				GA
1721	Coleoptera	Tenebrionidae				GA
1722	Coleoptera	Tenebrionidae				GA
1734	Coleoptera	Tenebrionidae				GA
930	Coleoptera	Tenebrionidae		<i>Chalcopteroides</i>		K
1012	Coleoptera	Tenebrionidae		<i>Helea</i>		K
1389	Coleoptera	Tenebrionidae		<i>Pterohelaeus</i>		GA
711	Coleoptera	Tenebrionidae		<i>Oectosis</i>		K
778	Coleoptera	Tenebrionidae		<i>Oectosis</i>		GA
192	Coleoptera	Tenebrionidae	Lagriinae	<i>Lagria</i>	<i>aneouiobcea</i>	GA
1061	Coleoptera	Trogidae				
1086	Coleoptera	Trogidae				
1097	Coleoptera	Trogidae				
1136	Coleoptera	Trogidae				
1189	Coleoptera	Trogidae				
825	Coleoptera	Trogidae		<i>Omorgus</i>		K
848	Coleoptera	Trogidae		<i>Omorgus</i>		
935	Coleoptera	Trogidae		<i>Omorgus</i>		K
1112	Dermaptera					
1113	Dermaptera					
1390	Dermaptera					
1433	Dermaptera					
1538	Dermaptera					
1678	Dermaptera					
1723	Dermaptera					
1790	Dermaptera					
123	Dermaptera					K
257	Dermaptera					K
258	Dermaptera					K
484	Dermaptera					K
491	Dermaptera					
492	Dermaptera					
522	Dermaptera					
734	Dermaptera					K
682	Dermaptera	Anisolabididae	Isolabellinae			
999	Dermaptera	Spongiphoridae				
876	Diplopoda					
966	Diplopoda					
1526	Diplopoda					
1546	Diplopoda					
259	Diplopoda					
260	Diplopoda					K
717	Diplopoda					
1336	Diptera	Acroceridae				K
1472	Diptera	Acroceridae				K
127	Diptera	Anthomyiidae				
530	Diptera	Anthomyiidae				
1343	Diptera	Apioceridae				K
1393	Diptera	Apioceridae				
1812	Diptera	Apioceridae				K
1232	Diptera	Asilidae				GA

Spec #	Order	Family	Tax 3	Genus	Species	GR/K
1255	Diptera	Asilidae				GA
1273	Diptera	Asilidae				GA
1369	Diptera	Asilidae				GA
1370	Diptera	Asilidae				GA
1411	Diptera	Asilidae				GA
1424	Diptera	Asilidae				GA
1478	Diptera	Asilidae				GA
1529	Diptera	Asilidae				GA
1690	Diptera	Asilidae				GA
1703	Diptera	Asilidae				GA
1732	Diptera	Asilidae				GA
1755	Diptera	Asilidae				GA
1756	Diptera	Asilidae				GA
1760	Diptera	Asilidae				GA
1784	Diptera	Asilidae				GA
1811	Diptera	Asilidae				GA
564	Diptera	Asilidae				GA
165	Diptera	Asilidae				GA
204	Diptera	Asilidae				GA
217	Diptera	Asilidae				GA
312	Diptera	Asilidae				GA
313	Diptera	Asilidae				GA
532	Diptera	Asilidae				GA
541	Diptera	Asilidae				GA
751	Diptera	Asilidae				GA
775	Diptera	Asilidae				GA
810	Diptera	Asilidae				GA
907	Diptera	Bombyliidae				K
1053	Diptera	Bombyliidae				K
1202	Diptera	Bombyliidae				
1542	Diptera	Bombyliidae				K
245	Diptera	Bombyliidae				K
506	Diptera	Bombyliidae				K
683	Diptera	Bombyliidae				K
719	Diptera	Bombyliidae				K
745	Diptera	Bombyliidae				K
1190	Diptera	Calliphoridae				
1603	Diptera	Calliphoridae				
1634	Diptera	Calliphoridae				
53	Diptera	Calliphoridae		<i>Calliphora</i>		
480	Diptera	Calliphoridae		<i>Calliphora</i>		
1419	Diptera	Calliphoridae ?				
1561	Diptera	Calliphoridae ?				
676	Diptera	Conopoidea	Conopidae			
1291	Diptera	Dolichopodidae				
125	Diptera	Drosophilidae				
179	Diptera	Drosophilidae				
1230	Diptera	Empididae				
1725	Diptera	Helosciomyzidae				
68	Diptera	Lauxaniidae				
1358	Diptera	Muscidae				
51	Diptera	Muscidae				
128	Diptera	Muscidae				
134	Diptera	Muscidae				

Spec #	Order	Family	Tax 3	Genus	Species	GR/K
205	Diptera	Muscidae				
498	Diptera	Muscidae				
675	Diptera	Muscidae				
818	Diptera	Muscidae				
88	Diptera	Pyrgotidae				K
1151	Diptera	Sarcophagidae				
1191	Diptera	Sarcophagidae				
1229	Diptera	Sarcophagidae				
1267	Diptera	Sarcophagidae				K
1274	Diptera	Sarcophagidae				
1704	Diptera	Sarcophagidae				
579	Diptera	Sarcophagidae				
1203	Diptera	Syrphidae				
1268	Diptera	Syrphidae				K
1421	Diptera	Syrphidae				
1422	Diptera	Syrphidae				
1425	Diptera	Syrphidae				
1439	Diptera	Syrphidae				
1455	Diptera	Syrphidae				
1480	Diptera	Syrphidae				
54	Diptera	Syrphidae				
129	Diptera	Syrphidae				
130	Diptera	Syrphidae				
143	Diptera	Syrphidae				
206	Diptera	Syrphidae				
242	Diptera	Syrphidae				
565	Diptera	Syrphidae				K
884	Diptera	Tabanidae				GA
901	Diptera	Tabanidae				GA
1297	Diptera	Tabanidae				GA
1321	Diptera	Tabanidae				
1440	Diptera	Tabanidae				GA
1548	Diptera	Tabanidae				GA
1630	Diptera	Tabanidae				GA
603	Diptera	Tabanidae				GA
126	Diptera	Tabanidae				GA
178	Diptera	Tabanidae				GA
466	Diptera	Tabanidae				GA
467	Diptera	Tabanidae				GA
473	Diptera	Tabanidae				GA
495	Diptera	Tabanidae				GA
531	Diptera	Tabanidae				GA
929	Diptera	Tachinidae				
1398	Diptera	Tachinidae				
136	Diptera	Tachinidae				K
464	Diptera	Tachinidae				K
1382	Diptera	Therevidae				K
1395	Diptera	Therevidae				K
1769	Diptera	Therevidae				K
1802	Diptera	Therevidae				K
142	Diptera	Therevidae				K
1233	Diptera	Tipulidae				
1324	Diptera	Tipulidae				
1615	Diptera	Tipulidae				

Spec #	Order	Family	Tax 3	Genus	Species	GR/K
577	Diptera	Tipulidae				
16	Diptera	Tipulidae				K
588	Diptera	Tipulidae				
792	Diptera	Tipulidae				K
195	Diptera	UNKNOWN				
969	Gastropoda					
1330	Gastropoda					
1431	Gastropoda					
1527	Gastropoda					
1565	Gastropoda					
1590	Gastropoda					
1692	Gastropoda					
1714	Gastropoda					
963	Hemiptera					
964	Hemiptera					
1024	Hemiptera					
1049	Hemiptera					
1072	Hemiptera					
1090	Hemiptera					
1122	Hemiptera					
1195	Hemiptera					
1196	Hemiptera					
1227	Hemiptera					
1239	Hemiptera					
1240	Hemiptera					
1257	Hemiptera					
1260	Hemiptera					
1281	Hemiptera					
1299	Hemiptera					
1302	Hemiptera					
1320	Hemiptera					
1524	Hemiptera					
1697	Hemiptera					
1751	Hemiptera					
107	Hemiptera					
109	Hemiptera					
110	Hemiptera					
164	Hemiptera					
166	Hemiptera					
170	Hemiptera					
177	Hemiptera					
187	Hemiptera					
188	Hemiptera					
200	Hemiptera					
239	Hemiptera					
241	Hemiptera					
249	Hemiptera					
489	Hemiptera					K
1753	Hemiptera	Alydidae				
1748	Hemiptera	Alydidae		<i>Riptortus</i>		
787	Hemiptera	Cicadellidae				GA
1288	Hemiptera	Cicadidae				
1394	Hemiptera	Cicadidae				
49	Hemiptera	Cicadidae		<i>Cicadetta</i>		

Spec #	Order	Family	Tax 3	Genus	Species	GR/K
207	Hemiptera	Cicadidae		<i>Cicadetta</i>		K
916	Hemiptera	Cicadidae				
1667	Hemiptera	Coreidae		<i>Amorbus</i> ?		
700	Hemiptera	Coreidae		<i>Amorbus</i>	<i>bispinus</i>	
503	Hemiptera	Eurymelidae		<i>Pogonoscopus</i>		K
1415	Hemiptera	Flatidae				
764	Hemiptera	Fulgoridae				
1351	Hemiptera	Gelastocoridae				
1383	Hemiptera	Gelastocoridae				
1567	Hemiptera	Gelastocoridae				
1611	Hemiptera	Gelastocoridae				
527	Hemiptera	Gelastocoridae		<i>Nerthra</i>		
1445	Hemiptera	Lygaeidae				K
679	Hemiptera	Lygaeidae				GA
108	Hemiptera	Membracidae				K
301	Hemiptera	Membracidae				K
302	Hemiptera	Membracidae				K
1683	Hemiptera	Nabidae				
838	Hemiptera	Pentatomidae				
960	Hemiptera	Pentatomidae				
962	Hemiptera	Pentatomidae				
990	Hemiptera	Pentatomidae				
1466	Hemiptera	Pentatomidae				
105	Hemiptera	Pentatomidae				K
117	Hemiptera	Pentatomidae				
153	Hemiptera	Pentatomidae				
176	Hemiptera	Pentatomidae				
221	Hemiptera	Pentatomidae				
240	Hemiptera	Pentatomidae				K
251	Hemiptera	Pentatomidae				
475	Hemiptera	Pentatomidae				K
513	Hemiptera	Pentatomidae				K
669	Hemiptera	Pentatomidae				
670	Hemiptera	Pentatomidae				
680	Hemiptera	Pentatomidae				
678	Hemiptera	Pentatomidae	(green)			
779	Hemiptera	Pentatomidae	(nymph)			
230	Hemiptera	Pseudococcidae				
433	Hemiptera	Reduviidae				
433	Hemiptera	Reduviidae				
863	Hemiptera	Reduviidae				
885	Hemiptera	Reduviidae				
886	Hemiptera	Reduviidae				
1379	Hemiptera	Reduviidae				
1576	Hemiptera	Reduviidae				
1647	Hemiptera	Reduviidae				
150	Hemiptera	Reduviidae				K
163	Hemiptera	Reduviidae				
196	Hemiptera	Reduviidae				
270	Hemiptera	Reduviidae				
284	Hemiptera	Reduviidae				K
311	Hemiptera	Reduviidae				K
482	Hemiptera	Reduviidae				K
512	Hemiptera	Reduviidae				K

Spec #	Order	Family	Tax 3	Genus	Species	GR/K
714	Hemiptera	Reduviidae				
573	Hemiptera	Reduviidae				
504	Hymenoptera					
1264	Hymenoptera	Apidae				K
1271	Hymenoptera	Apidae				
1328	Hymenoptera	Apidae				
52	Hymenoptera	Apidae		<i>Apis</i>	<i>melifera</i>	K
1093	Hymenoptera	Braconidae				
1258	Hymenoptera	Braconidae				
1259	Hymenoptera	Braconidae				
1467	Hymenoptera	Braconidae				
1525	Hymenoptera	Braconidae				
1528	Hymenoptera	Braconidae				
184	Hymenoptera	Braconidae				
493	Hymenoptera	Braconidae				K
1552	Hymenoptera	Chrysidae				K
893	Hymenoptera	Colletidae				K
1272	Hymenoptera	Colletidae				K
1289	Hymenoptera	Colletidae				K
1410	Hymenoptera	Colletidae				K
1794	Hymenoptera	Colletidae				K
1808	Hymenoptera	Colletidae				K
545	Hymenoptera	Colletidae				K
596	Hymenoptera	Colletidae				K
183	Hymenoptera	Colletidae				K
186	Hymenoptera	Colletidae				K
203	Hymenoptera	Colletidae				K
546	Hymenoptera	Colletidae				K
696	Hymenoptera	Colletidae				K
704	Hymenoptera	Colletidae				K
730	Hymenoptera	Colletidae				K
1362	Hymenoptera	Evaniidae				K
1553	Hymenoptera	Evaniidae				K
243	Hymenoptera	Evaniidae				K
500	Hymenoptera	Evaniidae				K
888	Hymenoptera	Formicidae				
889	Hymenoptera	Formicidae				
952	Hymenoptera	Formicidae				
1006	Hymenoptera	Formicidae				
1011	Hymenoptera	Formicidae				
1495	Hymenoptera	Formicidae				
1496	Hymenoptera	Formicidae				
1497	Hymenoptera	Formicidae				
1507	Hymenoptera	Formicidae				
1537	Hymenoptera	Formicidae				
1569	Hymenoptera	Formicidae				
1664	Hymenoptera	Formicidae				
1700	Hymenoptera	Formicidae				
1776	Hymenoptera	Formicidae				
423	Hymenoptera	Formicidae		<i>Camponotus</i>		K
1661	Hymenoptera	Formicidae		<i>Camponotus</i>		
1770	Hymenoptera	Formicidae		<i>Camponotus</i>		
1585	Hymenoptera	Formicidae		<i>Camponotus</i>		
1594	Hymenoptera	Formicidae		<i>Camponotus</i>		

Spec #	Order	Family	Tax 3	Genus	Species	GR/K
1602	Hymenoptera	Formicidae		<i>Camponotus</i>		
1575	Hymenoptera	Formicidae		<i>Rhytidoponera</i>		
1618	Hymenoptera	Formicidae		<i>Rhytidoponera</i>		
510	Hymenoptera	Formicidae				
275	Hymenoptera	Formicidae		<i>Iridomyrmex</i>		
737	Hymenoptera	Formicidae		<i>Pachycondyla</i>		11
535	Hymenoptera	Formicidae	Dolichoderinae	<i>Iridomyrmex</i>		
1676	Hymenoptera	Formicidae	Formicinae			
222	Hymenoptera	Formicidae	Myrmeciinae	<i>Myrmecia</i>		GA
252	Hymenoptera	Formicidae	Myrmeciinae	<i>Myrmecia</i>		GA
279	Hymenoptera	Formicidae	Myrmeciinae	<i>Myrmecia</i>		GA
281	Hymenoptera	Formicidae	Myrmeciinae	<i>Myrmecia</i>		GA
343	Hymenoptera	Formicidae	Myrmeciinae	<i>Myrmecia</i>		GA
478	Hymenoptera	Formicidae	Myrmeciinae	<i>Myrmecia</i>		GA
486	Hymenoptera	Formicidae	Myrmeciinae	<i>Myrmecia</i>		GA
487	Hymenoptera	Formicidae	Myrmeciinae	<i>Myrmecia</i>		GA
552	Hymenoptera	Formicidae	Myrmeciinae	<i>Myrmecia</i>		GA
712	Hymenoptera	Formicidae	Myrmeciinae	<i>Myrmecia</i>		GA
945	Hymenoptera	Formicidae	Myrmeciinae	<i>Myrmecia</i>		GA
974	Hymenoptera	Formicidae	Myrmeciinae	<i>Myrmecia</i>		GA
998	Hymenoptera	Formicidae	Myrmeciinae	<i>Myrmecia</i>		GA
1209	Hymenoptera	Formicidae	Myrmeciinae	<i>Myrmecia</i>		GA
1226	Hymenoptera	Formicidae	Myrmeciinae	<i>Myrmecia</i>		GA
1266	Hymenoptera	Formicidae	Myrmeciinae	<i>Myrmecia</i>		GA
1269	Hymenoptera	Formicidae	Myrmeciinae	<i>Myrmecia</i>		GA
1280	Hymenoptera	Formicidae	Myrmeciinae	<i>Myrmecia</i>		GA
1287	Hymenoptera	Formicidae	Myrmeciinae	<i>Myrmecia</i>		GA
1293	Hymenoptera	Formicidae	Myrmeciinae	<i>Myrmecia</i>		GA
1317	Hymenoptera	Formicidae	Myrmeciinae	<i>Myrmecia</i>		GA
1374	Hymenoptera	Formicidae	Myrmeciinae	<i>Myrmecia</i>		GA
1384	Hymenoptera	Formicidae	Myrmeciinae	<i>Myrmecia</i>		GA
1391	Hymenoptera	Formicidae	Myrmeciinae	<i>Myrmecia</i>		GA
1406	Hymenoptera	Formicidae	Myrmeciinae	<i>Myrmecia</i>		GA
1457	Hymenoptera	Formicidae	Myrmeciinae	<i>Myrmecia</i>		GA
1473	Hymenoptera	Formicidae	Myrmeciinae	<i>Myrmecia</i>		GA
1534	Hymenoptera	Formicidae	Myrmeciinae	<i>Myrmecia</i>		GA
1535	Hymenoptera	Formicidae	Myrmeciinae	<i>Myrmecia</i>		GA
1577	Hymenoptera	Formicidae	Myrmeciinae	<i>Myrmecia</i>		GA
1649	Hymenoptera	Formicidae	Myrmeciinae	<i>Myrmecia</i>		GA
1668	Hymenoptera	Formicidae	Myrmeciinae	<i>Myrmecia</i>		GA
1673	Hymenoptera	Formicidae	Myrmeciinae	<i>Myrmecia</i>		GA
1686	Hymenoptera	Formicidae	Myrmeciinae	<i>Myrmecia</i>		GA
1687	Hymenoptera	Formicidae	Myrmeciinae	<i>Myrmecia</i>		GA
1689	Hymenoptera	Formicidae	Myrmeciinae	<i>Myrmecia</i>		GA
1718	Hymenoptera	Formicidae	Myrmeciinae	<i>Myrmecia</i>		GA
1757	Hymenoptera	Formicidae	Myrmeciinae	<i>Myrmecia</i>		GA
1759	Hymenoptera	Formicidae	Myrmeciinae	<i>Myrmecia</i>		GA
1782	Hymenoptera	Formicidae	Myrmeciinae	<i>Myrmecia</i>		GA
1787	Hymenoptera	Formicidae	Myrmeciinae	<i>Myrmecia</i>		GA
1795	Hymenoptera	Formicidae	Myrmeciinae	<i>Myrmecia</i>		GA
1805	Hymenoptera	Formicidae	Myrmeciinae	<i>Myrmecia</i>		GA
1807	Hymenoptera	Formicidae	Myrmeciinae	<i>Myrmecia</i>		GA
408	Hymenoptera	Formicidae	Myrmeciinae	<i>Myrmecia</i>		GA
409	Hymenoptera	Formicidae	Myrmeciinae	<i>Myrmecia</i>		GA

Spec #	Order	Family	Tax 3	Genus	Species	GR/K
487	Hymenoptera	Formicidae	Myrmeciinae	<i>Myrmecia</i>		GA
664	Hymenoptera	Formicidae	Myrmeciinae	<i>Myrmecia</i>		GA
477	Hymenoptera	Formicidae	Myrmeciinae	<i>Myrmecia</i>	<i>callima</i>	GA
543	Hymenoptera	Formicidae	Poneri	<i>Rhytidoponera</i>		
542	Hymenoptera	Formicidae	Ponerinae	<i>Prionopella</i>		
697	Hymenoptera	Gasteruptiidae				
1771	Hymenoptera	Ichneumonidae				GA
1037	Hymenoptera	Ichneumonidae				GA
1038	Hymenoptera	Ichneumonidae				GA
1055	Hymenoptera	Ichneumonidae				GA
1077	Hymenoptera	Ichneumonidae				GA
1079	Hymenoptera	Ichneumonidae				GA
1089	Hymenoptera	Ichneumonidae				GA
1105	Hymenoptera	Ichneumonidae				GA
1146	Hymenoptera	Ichneumonidae				GA
1153	Hymenoptera	Ichneumonidae				GA
1156	Hymenoptera	Ichneumonidae				GA
1164	Hymenoptera	Ichneumonidae				GA
1165	Hymenoptera	Ichneumonidae				GA
1436	Hymenoptera	Ichneumonidae				GA
1606	Hymenoptera	Ichneumonidae				GA
1637	Hymenoptera	Ichneumonidae				GA
1648	Hymenoptera	Ichneumonidae				GA
1662	Hymenoptera	Ichneumonidae				GA
1685	Hymenoptera	Ichneumonidae				GA
1688	Hymenoptera	Ichneumonidae				GA
1720	Hymenoptera	Ichneumonidae				GA
1740	Hymenoptera	Ichneumonidae				GA
1743	Hymenoptera	Ichneumonidae				GA
515	Hymenoptera	Ichneumonidae				GA
698	Hymenoptera	Ichneumonidae				GA
87	Hymenoptera	Ichneumonidae		<i>Ophion</i>		GA
533	Hymenoptera	Ichneumonidae	Branchinae	<i>Australoglypta</i>		GA
1359	Hymenoptera	Mutillidae				K
1613	Hymenoptera	Mutillidae				K
1623	Hymenoptera	Mutillidae				K
534	Hymenoptera	Mutillidae				
580	Hymenoptera	Mutillidae				K
1380	Hymenoptera	Mutillidae				
1804	Hymenoptera	Mutillidae				K
1694	Hymenoptera	Pergidae				GA
1550	Hymenoptera	Pergidae		<i>Perga</i>		GA
1002	Hymenoptera	Pompilidae				GA
1017	Hymenoptera	Pompilidae				GA
1204	Hymenoptera	Pompilidae				GA
1356	Hymenoptera	Pompilidae				GA
1591	Hymenoptera	Pompilidae				GA
1598	Hymenoptera	Pompilidae				GA
1601	Hymenoptera	Pompilidae				GA
1777	Hymenoptera	Pompilidae				GA
1781	Hymenoptera	Pompilidae				GA
1788	Hymenoptera	Pompilidae				GA
584	Hymenoptera	Pompilidae				GA
607	Hymenoptera	Pompilidae				GA

Spec #	Order	Family	Tax 3	Genus	Species	GR/K
611	Hymenoptera	Pompilidae				GA
616	Hymenoptera	Pompilidae				GA
295	Hymenoptera	Pompilidae				GA
494	Hymenoptera	Pompilidae				GA
612	Hymenoptera	Pompilidae				GA
617	Hymenoptera	Pompilidae				GA
619	Hymenoptera	Pompilidae				GA
699	Hymenoptera	Pompilidae				GA
790	Hymenoptera	Pompilidae				GA
813	Hymenoptera	Pompilidae				GA
622	Hymenoptera	Pompilidae		<i>Cryptocheilus</i>	<i>fabricolor</i>	GA
1742	Hymenoptera	Scoliidae				
1212	Hymenoptera	Sphecidae				K
1344	Hymenoptera	Sphecidae				K
1402	Hymenoptera	Sphecidae				
1408	Hymenoptera	Sphecidae				K
1727	Hymenoptera	Sphecidae				
505	Hymenoptera	Sphecidae				K
720	Hymenoptera	Sphecidae				
723	Hymenoptera	Sphecidae				
1806	Hymenoptera	Sphecidae	Larrinae			
1071	Hymenoptera	Tiphiidae				GA
1234	Hymenoptera	Tiphiidae				GA
1335	Hymenoptera	Tiphiidae				GA
1458	Hymenoptera	Tiphiidae				GA
1479	Hymenoptera	Tiphiidae				GA
1706	Hymenoptera	Tiphiidae				GA
1707	Hymenoptera	Tiphiidae				GA
604	Hymenoptera	Tiphiidae				GA
614	Hymenoptera	Tiphiidae				GA
796	Hymenoptera	Tiphiidae				GA
801	Hymenoptera	Tiphiidae				GA
806	Hymenoptera	Tiphiidae				GA
1712	Hymenoptera	Tiphiidae	Thynninae			GA
1713	Hymenoptera	Tiphiidae	Thynninae			GA
1730	Hymenoptera	Tiphiidae	Thynninae			GA
1731	Hymenoptera	Tiphiidae	Thynninae			GA
1286	Hymenoptera	Tiphiidae	Thynninae			GA
1325	Hymenoptera	Tiphiidae	Thynninae			GA
1530	Hymenoptera	Tiphiidae	Thynninae			GA
1663	Hymenoptera	Tiphiidae	Thynninae			GA
1709	Hymenoptera	Tiphiidae	Thynninae			GA
481	Hymenoptera	Tiphiidae	Thynninae			GA
516	Hymenoptera	Tiphiidae	Thynninae			GA
685	Hymenoptera	Tiphiidae	Thynninae			GA
894	Hymenoptera	Vespidae				
1198	Hymenoptera	Vespidae				K
1371	Hymenoptera	Vespidae				
1701	Hymenoptera	Vespidae				K
1339	Hymenoptera	Vespidae	Eumeninae			K
544	Isopoda					
549	Isopoda					
1305	Isopoda					GR
1430	Isopoda					

Spec #	Order	Family	Tax 3	Genus	Species	GR/K
1586	Isopoda					
1735	Isopoda					
519	Isopoda		collective sample			
262	Isopoda					GR
539	Isopoda					
540	Isopoda					
671	Isopoda					
73	Lepidoptera					
345	Lepidoptera					
556	Lepidoptera					
627	Lepidoptera					
797	Lepidoptera					
1023	Lepidoptera					
1027	Lepidoptera					
1028	Lepidoptera					
1029	Lepidoptera					
1030	Lepidoptera					
1031	Lepidoptera					
1032	Lepidoptera					
1034	Lepidoptera					
1036	Lepidoptera					
1045	Lepidoptera					
1046	Lepidoptera					
1047	Lepidoptera					
1048	Lepidoptera					
1051	Lepidoptera					
1054	Lepidoptera					
1056	Lepidoptera					
1060	Lepidoptera					
1066	Lepidoptera					
1068	Lepidoptera					
1070	Lepidoptera					
1075	Lepidoptera					
1078	Lepidoptera					
1081	Lepidoptera					
1084	Lepidoptera					
1085	Lepidoptera					
1088	Lepidoptera					
1091	Lepidoptera					
1094	Lepidoptera					
1095	Lepidoptera					
1096	Lepidoptera					
1098	Lepidoptera					
1099	Lepidoptera					
1106	Lepidoptera					
1119	Lepidoptera					
1127	Lepidoptera					
1128	Lepidoptera					
1129	Lepidoptera					
1130	Lepidoptera					
1131	Lepidoptera					
1132	Lepidoptera					
1134	Lepidoptera					
1135	Lepidoptera					

Spec #	Order	Family	Tax 3	Genus	Species	GR/K
1137	Lepidoptera					
1139	Lepidoptera					
1140	Lepidoptera					
1141	Lepidoptera					
1142	Lepidoptera					
1144	Lepidoptera					
1145	Lepidoptera					
1147	Lepidoptera					
1148	Lepidoptera					
1149	Lepidoptera					
1150	Lepidoptera					
1152	Lepidoptera					
1155	Lepidoptera					
1157	Lepidoptera					
1158	Lepidoptera					
1161	Lepidoptera					
1162	Lepidoptera					
1163	Lepidoptera					
1166	Lepidoptera					
1167	Lepidoptera					
1168	Lepidoptera					
1169	Lepidoptera					
1170	Lepidoptera					
1171	Lepidoptera					
1173	Lepidoptera					
1174	Lepidoptera					
1178	Lepidoptera					
1179	Lepidoptera					
1180	Lepidoptera					
1181	Lepidoptera					
1184	Lepidoptera					
1193	Lepidoptera					
1194	Lepidoptera					
1201	Lepidoptera					
1222	Lepidoptera					
1256	Lepidoptera					
1296	Lepidoptera					K
1301	Lepidoptera					
1454	Lepidoptera					
1488	Lepidoptera					
1489	Lepidoptera					
1490	Lepidoptera					
1491	Lepidoptera					
1492	Lepidoptera					
1493	Lepidoptera					
1494	Lepidoptera					
1499	Lepidoptera					
1501	Lepidoptera					
1502	Lepidoptera					
1503	Lepidoptera					
1504	Lepidoptera					
1506	Lepidoptera					
1509	Lepidoptera					
1510	Lepidoptera					

Spec #	Order	Family	Tax 3	Genus	Species	GR/K
1512	Lepidoptera					
1513	Lepidoptera					
1514	Lepidoptera					
1515	Lepidoptera					
1516	Lepidoptera					
1517	Lepidoptera					
1518	Lepidoptera					
1519	Lepidoptera					
1521	Lepidoptera					
1626	Lepidoptera					
1627	Lepidoptera					
1628	Lepidoptera					
1631	Lepidoptera					
1632	Lepidoptera					
1633	Lepidoptera					
1635	Lepidoptera					
1638	Lepidoptera					
1641	Lepidoptera					
1643	Lepidoptera					
1645	Lepidoptera					
1646	Lepidoptera					
1650	Lepidoptera					
1651	Lepidoptera					
1652	Lepidoptera					
1654	Lepidoptera					
1655	Lepidoptera					
1657	Lepidoptera					
1658	Lepidoptera					
1659	Lepidoptera					
1665	Lepidoptera					
1674	Lepidoptera					
1729	Lepidoptera					
1815	Lepidoptera					
322	Lepidoptera					
752	Lepidoptera					
20	Lepidoptera					
21	Lepidoptera					
25	Lepidoptera					
32	Lepidoptera					
33	Lepidoptera					
38	Lepidoptera					
48	Lepidoptera					
60	Lepidoptera					
61	Lepidoptera					
62	Lepidoptera					K
63	Lepidoptera					
65	Lepidoptera					
67	Lepidoptera					
73	Lepidoptera					
76	Lepidoptera					
77	Lepidoptera					
80	Lepidoptera					K
104	Lepidoptera					
111	Lepidoptera					

Spec #	Order	Family	Tax 3	Genus	Species	GR/K
138	Lepidoptera					
197	Lepidoptera					
236	Lepidoptera					
238	Lepidoptera					
315	Lepidoptera					
316	Lepidoptera					
337	Lepidoptera					
350	Lepidoptera					
362	Lepidoptera					
366	Lepidoptera					
367	Lepidoptera					
369	Lepidoptera					
376	Lepidoptera					
380	Lepidoptera					
382	Lepidoptera					
383	Lepidoptera					
387	Lepidoptera					
394	Lepidoptera					
399	Lepidoptera					
406	Lepidoptera					
407	Lepidoptera					
411	Lepidoptera					
413	Lepidoptera					
414	Lepidoptera					
419	Lepidoptera					
420	Lepidoptera					
421	Lepidoptera					
422	Lepidoptera					
428	Lepidoptera					
429	Lepidoptera					
430	Lepidoptera					
431	Lepidoptera					
434	Lepidoptera					
438	Lepidoptera					
441	Lepidoptera					
442	Lepidoptera					
443	Lepidoptera					
446	Lepidoptera					K
452	Lepidoptera					K
453	Lepidoptera					
454	Lepidoptera					
456	Lepidoptera					
459	Lepidoptera					
657	Lepidoptera					K
660	Lepidoptera					
661	Lepidoptera					
662	Lepidoptera					
750	Lepidoptera					
754	Lepidoptera					
760	Lepidoptera					
773	Lepidoptera					
797	Lepidoptera					
798	Lepidoptera					
5	Lepidoptera			<i>Agrotis ?</i>		

Spec #	Order	Family	Tax 3	Genus	Species	GR/K
341	Lepidoptera	?				
821	Lepidoptera	?				
828	Lepidoptera	?				
829	Lepidoptera	?				
849	Lepidoptera	?				
855	Lepidoptera	?				
856	Lepidoptera	?				
861	Lepidoptera	?				
864	Lepidoptera	?				
866	Lepidoptera	?				
896	Lepidoptera	?				
900	Lepidoptera	?				
915	Lepidoptera	?				
917	Lepidoptera	?				
918	Lepidoptera	?				
920	Lepidoptera	?				
921	Lepidoptera	?				
944	Lepidoptera	?				
946	Lepidoptera	?				
947	Lepidoptera	?				
950	Lepidoptera	?				
954	Lepidoptera	?				
958	Lepidoptera	?				
978	Lepidoptera	?				
983	Lepidoptera	?				
984	Lepidoptera	?				
985	Lepidoptera	?				
986	Lepidoptera	?				
1018	Lepidoptera	?				
1019	Lepidoptera	?				
1020	Lepidoptera	?				
840	Lepidoptera	?				
381	Lepidoptera	Anthelidae				K
457	Lepidoptera	Anthelidae				K
352	Lepidoptera	Anthelidae		<i>Anthela</i>		K
91	Lepidoptera	Anthelidae		<i>Chenuala</i>		K
987	Lepidoptera	Arctiidae		<i>Utetheisa</i>	<i>pulchelloides</i>	
6	Lepidoptera	Arctiidae				K
445	Lepidoptera	Arctiidae	Arctiinae	<i>Spilosoma</i>		K
749	Lepidoptera	Bombycidae				
1	Lepidoptera	Carthaeidae		<i>Carthaea</i>	<i>saturnioides</i>	K
658	Lepidoptera	Gelechiodea				
820	Lepidoptera	Geometridae				
827	Lepidoptera	Geometridae				
835	Lepidoptera	Geometridae				
836	Lepidoptera	Geometridae				
858	Lepidoptera	Geometridae				
862	Lepidoptera	Geometridae				
919	Lepidoptera	Geometridae				
923	Lepidoptera	Geometridae				
925	Lepidoptera	Geometridae				
927	Lepidoptera	Geometridae				
942	Lepidoptera	Geometridae				
977	Lepidoptera	Geometridae				

Spec #	Order	Family	Tax 3	Genus	Species	GR/K
955	Lepidoptera	Geometridae		<i>Hypobapta</i>	<i>percomptaria</i>	K
832	Lepidoptera	Geometridae		<i>Lissomma</i>		
12	Lepidoptera	Geometridae				K
23	Lepidoptera	Geometridae				K
24	Lepidoptera	Geometridae				K
41	Lepidoptera	Geometridae				K
46	Lepidoptera	Geometridae				
47	Lepidoptera	Geometridae				
50	Lepidoptera	Geometridae				K
59	Lepidoptera	Geometridae				K
66	Lepidoptera	Geometridae				
72	Lepidoptera	Geometridae				
82	Lepidoptera	Geometridae				
83	Lepidoptera	Geometridae				
85	Lepidoptera	Geometridae				
86	Lepidoptera	Geometridae				
95	Lepidoptera	Geometridae				
96	Lepidoptera	Geometridae				
97	Lepidoptera	Geometridae				
317	Lepidoptera	Geometridae				
318	Lepidoptera	Geometridae				
321	Lepidoptera	Geometridae				K
323	Lepidoptera	Geometridae				
326	Lepidoptera	Geometridae				
327	Lepidoptera	Geometridae				
338	Lepidoptera	Geometridae				
339	Lepidoptera	Geometridae				
355	Lepidoptera	Geometridae				
358	Lepidoptera	Geometridae				
375	Lepidoptera	Geometridae				K
389	Lepidoptera	Geometridae				K
392	Lepidoptera	Geometridae				K
393	Lepidoptera	Geometridae				K
395	Lepidoptera	Geometridae				
402	Lepidoptera	Geometridae				
403	Lepidoptera	Geometridae				K
415	Lepidoptera	Geometridae				K
424	Lepidoptera	Geometridae				K
425	Lepidoptera	Geometridae				K
436	Lepidoptera	Geometridae				K
436	Lepidoptera	Geometridae				
451	Lepidoptera	Geometridae				K
517	Lepidoptera	Geometridae				
634	Lepidoptera	Geometridae				
637	Lepidoptera	Geometridae				
638	Lepidoptera	Geometridae				
639	Lepidoptera	Geometridae				
644	Lepidoptera	Geometridae				
655	Lepidoptera	Geometridae				
691	Lepidoptera	Geometridae				
694	Lepidoptera	Geometridae				
756	Lepidoptera	Geometridae				
757	Lepidoptera	Geometridae				
758	Lepidoptera	Geometridae				

Spec #	Order	Family	Tax 3	Genus	Species	GR/K
759	Lepidoptera	Geometridae				
765	Lepidoptera	Geometridae				
776	Lepidoptera	Geometridae				
320	Lepidoptera	Geometridae		<i>Arhodia</i>		K
22	Lepidoptera	Geometridae		<i>Chlorocoma</i>		K
19	Lepidoptera	Geometridae		<i>Chlorocoma</i>	<i>dicloraria</i>	K
330	Lepidoptera	Geometridae		<i>Crypsiphona</i>	<i>ocultaria</i>	K
357	Lepidoptera	Geometridae		<i>Eucyclodes</i>	<i>buprestaria</i>	K
417	Lepidoptera	Geometridae		<i>Gastrina</i>		K
42	Lepidoptera	Geometridae		<i>Gastrina</i>	<i>cristaria</i>	K
455	Lepidoptera	Geometridae		<i>Gastrina</i>	<i>cristaria</i>	
334	Lepidoptera	Geometridae		<i>Gastrina</i>	<i>cristarina</i>	K
663	Lepidoptera	Geometridae		<i>Heliomystis</i>		
31	Lepidoptera	Geometridae		<i>Parepisparis</i>	<i>excusata</i>	K
377	Lepidoptera	Geometridae		<i>Phallaria</i>	<i>ophiusaria</i>	K
384	Lepidoptera	Geometridae		<i>Pholodes</i>		K
385	Lepidoptera	Geometridae		<i>Pholodes</i>		K
450	Lepidoptera	Geometridae		<i>Thalaina</i>	<i>clara</i>	K
976	Lepidoptera	Geometridae	Oenochrominae			K
2	Lepidoptera	Geometridae	Oenochrominae	<i>Arhodia</i>		K
79	Lepidoptera	Geometridae	Oenochrominae	<i>Arhodia</i>		K
625	Lepidoptera	Geometridae ?				
830	Lepidoptera	Geometridae ?				
1022	Lepidoptera	Geometridae ?				
41	Lepidoptera	Geometridae ?				
753	Lepidoptera	Geometridae ?				
772	Lepidoptera	Geometridae ?				
761	Lepidoptera	Hepialidae		<i>Abantiades</i>		K
372	Lepidoptera	Hepialidae		<i>Abantiades</i>	<i>hydrographis</i>	GA
373	Lepidoptera	Hepialidae		<i>Abantiades</i>	<i>ocellatus</i>	GA
1228	Lepidoptera	Hesperiidae		<i>Exometoeca</i>	<i>nycteris</i>	K
593	Lepidoptera	Hesperiidae		<i>Hesperilla</i>	<i>chrysotricha</i>	K
1340	Lepidoptera	Hesperiidae	Trapezitinae	<i>Mesodina</i>	<i>cyanophracta</i>	
693	Lepidoptera	Lasiocampidae				
755	Lepidoptera	Lasiocampidae				
426	Lepidoptera	Lasiocampidae		<i>Entometa</i>		K
371	Lepidoptera	Lasiocampidae		<i>Entometa</i>	<i>fervens</i>	K
895	Lepidoptera	Limacodidae		<i>Doratifera</i>		
1625	Lepidoptera	Limacodidae		<i>Doratifera</i>		
551	Lepidoptera	Limacodidae		<i>Doratifera</i>	<i>quadriguttata</i>	
90	Lepidoptera	Limacodidae				K
81	Lepidoptera	Limacodidae		<i>Doratifera</i>		K
332	Lepidoptera	Limacodidae		<i>Doratifera</i>		K
398	Lepidoptera	Limacodidae		<i>Doratifera</i>		K
870	Lepidoptera	Lycaenidae				
1341	Lepidoptera	Lycaenidae				
1249	Lepidoptera	Lycaenidae		<i>Candalides</i>		
296	Lepidoptera	Lycaenidae				K
34	Lepidoptera	Lymantriidae		<i>Teia</i>	<i>athlophora</i>	K
345	Lepidoptera	Noctuidae				K
523	Lepidoptera	Noctuidae				
626	Lepidoptera	Noctuidae				
833	Lepidoptera	Noctuidae				
847	Lepidoptera	Noctuidae				

Spec #	Order	Family	Tax 3	Genus	Species	GR/K
853	Lepidoptera	Noctuidae				
996	Lepidoptera	Noctuidae				
1003	Lepidoptera	Noctuidae				
1511	Lepidoptera	Noctuidae				
345	Lepidoptera	Noctuidae				K
844	Lepidoptera	Noctuidae		<i>Agrotis</i>		
39	Lepidoptera	Noctuidae				K
74	Lepidoptera	Noctuidae				
75	Lepidoptera	Noctuidae				
133	Lepidoptera	Noctuidae				
137	Lepidoptera	Noctuidae				
139	Lepidoptera	Noctuidae				
140	Lepidoptera	Noctuidae				
329	Lepidoptera	Noctuidae				K
344	Lepidoptera	Noctuidae				K
346	Lepidoptera	Noctuidae				K
364	Lepidoptera	Noctuidae				
386	Lepidoptera	Noctuidae				
391	Lepidoptera	Noctuidae				
405	Lepidoptera	Noctuidae				
412	Lepidoptera	Noctuidae				K
435	Lepidoptera	Noctuidae				K
449	Lepidoptera	Noctuidae				K
518	Lepidoptera	Noctuidae				
589	Lepidoptera	Noctuidae				
598	Lepidoptera	Noctuidae				
642	Lepidoptera	Noctuidae				
648	Lepidoptera	Noctuidae				
649	Lepidoptera	Noctuidae				
650	Lepidoptera	Noctuidae				
656	Lepidoptera	Noctuidae				
769	Lepidoptera	Noctuidae				
770	Lepidoptera	Noctuidae				
771	Lepidoptera	Noctuidae				
799	Lepidoptera	Noctuidae				
18	Lepidoptera	Noctuidae		<i>Agrotis</i>	<i>munda</i>	K
659	Lepidoptera	Noctuidae		<i>Chrysodeixis</i>		
336	Lepidoptera	Noctuidae		<i>Chrysodeixis</i>	<i>argentifera</i>	K
30	Lepidoptera	Noctuidae		<i>Dasypodia</i>	<i>selenophora</i>	
388	Lepidoptera	Noctuidae		<i>Pantylia</i>		
379	Lepidoptera	Noctuidae		<i>Peripyra</i>	<i>sanguinipuncta</i>	K
185	Lepidoptera	Noctuidae		<i>Periscepta</i>	<i>polystieta</i>	K
40	Lepidoptera	Noctuidae		<i>Persectania</i>		K
686	Lepidoptera	Noctuidae		<i>Uraba</i>	<i>lugens</i>	K
859	Lepidoptera	Noctuidae	Mythimna (?)			
643	Lepidoptera	Noctuidae ?				
766	Lepidoptera	Noctuidae ?				
1738	Lepidoptera	Notodontidae		<i>Danima</i>		
4	Lepidoptera	Notodontidae		<i>Destolmia</i>	<i>lineata</i>	K
58	Lepidoptera	Notodontidae				K
374	Lepidoptera	Notodontidae				K
57	Lepidoptera	Notodontidae		<i>Danima</i>	<i>banksiae</i>	K
58	Lepidoptera	Notodontidae		<i>Hylaeora</i>	<i>dilucida</i>	K
306	Lepidoptera	Nymphalidae		<i>Geitoneura</i>	<i>klugii</i>	K

Spec #	Order	Family	Tax 3	Genus	Species	GR/K
594	Lepidoptera	Nymphalidae	Heteronympha	<i>Vanessa</i>	<i>kershawi</i>	K
298	Lepidoptera	Nymphalidae		<i>Merope</i>	<i>duboulayi</i>	K
64	Lepidoptera	Oecophoridae				K
396	Lepidoptera	Oecophoridae				
331	Lepidoptera	Oecophoridae		<i>Wingia</i>	<i>aurata</i>	K
325	Lepidoptera	Psychidae		<i>Iphierga</i>	<i>euphragma</i>	K
948	Lepidoptera	Pyralidae				
953	Lepidoptera	Pyralidae				
957	Lepidoptera	Pyralidae				
979	Lepidoptera	Pyralidae				
982	Lepidoptera	Pyralidae				
922	Lepidoptera	Pyralidae		<i>Hedonota</i>	<i>recurvella</i>	
333	Lepidoptera	Pyralidae				
341	Lepidoptera	Pyralidae				
342	Lepidoptera	Pyralidae				
356	Lepidoptera	Pyralidae				
365	Lepidoptera	Pyralidae				
397	Lepidoptera	Pyralidae				
401	Lepidoptera	Pyralidae				
432	Lepidoptera	Pyralidae				K
631	Lepidoptera	Pyralidae				K
635	Lepidoptera	Pyralidae	Epipaschinae			
641	Lepidoptera	Pyralidae				
645	Lepidoptera	Pyralidae				
646	Lepidoptera	Pyralidae				
84	Lepidoptera	Pyralidae		<i>Uresiphita</i>	<i>ornithopteralis</i>	K
460	Lepidoptera	Pyralidae				
837	Lepidoptera	Pyralidae ?				
928	Lepidoptera	Pyralidae ?				
1126	Lepidoptera	Pyralidae ?				
43	Lepidoptera	Pyralidae ?				K
768	Lepidoptera	Pyralidae ?				
328	Lepidoptera	Saturniidae		<i>Opodiphthera</i>	<i>helena</i>	K
819	Lepidoptera	Thaumetopoeidae		<i>Ochrogaster</i>		K
32	Lepidoptera	Thaumetopoeidae				K
692	Lepidoptera	Thaumetopoeidae				K
3	Lepidoptera	Thaumetopoeidae		<i>Epicoma</i>	<i>melanosticta</i>	K
7	Lepidoptera	Thaumetopoeidae		<i>Ochrogaster</i>		K
8	Lepidoptera	Thaumetopoeidae		<i>Ochrogaster</i>		K
9	Lepidoptera	Thaumetopoeidae		<i>Ochrogaster</i>		K
11	Lepidoptera	Thaumetopoeidae		<i>Ochrogaster</i>		K
35	Lepidoptera	Thaumetopoeidae		<i>Ochrogaster</i>		K
36	Lepidoptera	Thaumetopoeidae		<i>Ochrogaster</i>		K
10	Lepidoptera	Thaumetopoeidae		<i>Ochrogaster</i>	<i>lunifer</i>	K
404	Lepidoptera	Thaumetopoeidae		<i>Oenosandra</i>		K
324	Lepidoptera	Tineidae		<i>Moerarchis</i>		K
141	Lepidoptera	Tineidae		<i>Moerarchis</i>	<i>australasiella</i>	K
319	Lepidoptera	Tineidae		<i>Moerarchis</i>	<i>clathrella</i>	K
943	Lepidoptera	Tortricidae				
1814	Lepidoptera	Tortricidae ?				
92	Lepidoptera	Tortricidae ?				
1172	Lepidoptera	UNIDENTIFIABLE		unidentifiable	unidentifiable	
45	Lepidoptera	Zygaenidae		<i>Pollanisus</i>	<i>viridipulverulenta</i>	K
1459	Mantodea					

Spec #	Order	Family	Tax 3	Genus	Species	GR/K
132	Mantodea					
309	Mantodea					
674	Mantodea					
718	Mantodea					
784	Mantodea					
789	Mantodea					
739	Mantodea	Amorphoscelidae		<i>Paroxypilus</i>	<i>tasmaniensis?</i>	
854	Mantodea	Amorphoscelidae	Paraoxypilinae			
1124	Mantodea	Mantidae				K
1541	Mantodea	Mantidae				
767	Mantodea	Mantidae				
981	Mantodea	Mantidae	Mantinae			
1001	Mantodea	Mantidae	Mantinae			
1277	Mecoptera	Bittacidae				GA
1453	Mecoptera	Bittacidae				GA
1456	Mecoptera	Bittacidae				GA
1463	Mecoptera	Bittacidae				GA
908	Mecoptera	Bittacidae		<i>Harpobittacus</i>		GA
250	Mecoptera	Bittacidae		<i>Harpobittacus</i>		GA
89	Mecoptera	Meropidae		<i>Austromerope</i>	<i>poultoni</i>	GR
1636	Megaloptera	Corydalidae	Chauliodinae			GR
1532	Mygalomorphae	Barychelidae				GR
497	Mygalomorphae	Barychelidae ?				GR
1468	Mygalomorphae	Idiopidae				GR
887	Mygalomorphae	Nemesiidae				GR
1367	Mygalomorphae	Nemesiidae				GR
1401	Mygalomorphae	Nemesiidae				GR
1560	Mygalomorphae	Nemesiidae				GR
1792	Mygalomorphae	Nemesiidae				GR
567	Mygalomorphae	Nemesiidae		<i>Chenistonia</i>		GR
581	Mygalomorphae	Nemesiidae		<i>Chenistonia</i>		GR
1350	Mygalomorphae	Nemesiidae		<i>Chenistonia</i>		GR
590	Mygalomorphae	Nemesiidae			juvenile	GR
283	Mygalomorphae	Nemesiidae				GR
502	Mygalomorphae	Nemesiidae				GR
538	Mygalomorphae	Nemesiidae			juvenile	GR
585	Mygalomorphae	Nemesiidae			juvenile	GR
721	Mygalomorphae	Nemesiidae		<i>Chenistonia</i>		GR
949	Neuroptera	Chrysopidae				
1057	Neuroptera	Chrysopidae				
1186	Neuroptera	Chrysopidae				
1639	Neuroptera	Chrysopidae				
822	Neuroptera	Chrysopidae		<i>Chrysopa</i>		
361	Neuroptera	Chrysopidae		<i>Chrysopa</i>		
131	Neuroptera	Hemerobiidae				
360	Neuroptera	Hemerobiidae				
653	Neuroptera	Hemerobiidae				
1315	Neuroptera	Mantispidae				
687	Neuroptera	Mantispidae				
305	Neuroptera	Myrmeleontidae				
400	Neuroptera	Myrmeleontidae				
1005	Odonata	Zygoptera				GR
1758	Odonata	Zygoptera				GR
1004	Odonata	Zygoptera	Lestoidea			GR

Spec #	Order	Family	Tax 3	Genus	Species	GR/K
1237	Odonata	Zygoptera	Lestoidea			GR
237	Odonata	Zygoptera	Lestoidea			GR
1381	Oligochaeta	Megascolecidae				
1407	Oligochaeta	Megascolecidae				
1475	Oligochaeta	Megascolecidae				
1482	Oligochaeta	Megascolecidae				
1484	Oligochaeta	Megascolecidae				
1556	Oligochaeta	Megascolecidae				
1798	Oligochaeta	Megascolecidae				
520	Oligochaeta	Megascolecidae	collective sp			
937	Onychophora					GR
1481	Onychophora					GR
1365	Opiliones					GR
1447	Opiliones					GR
1616	Opiliones					GR
1799	Opiliones					
1125	Orthoptera					K
1176	Orthoptera					
1199	Orthoptera					
1210	Orthoptera					
1214	Orthoptera					
1218	Orthoptera					
1224	Orthoptera					
1235	Orthoptera					
1236	Orthoptera					
1241	Orthoptera					
1252	Orthoptera					
1261	Orthoptera					
1275	Orthoptera					
1284	Orthoptera					
1285	Orthoptera					
1308	Orthoptera					
1311	Orthoptera					
1312	Orthoptera					
1316	Orthoptera					
1323	Orthoptera					
1353	Orthoptera					
1372	Orthoptera					
1420	Orthoptera					
1426	Orthoptera					
1452	Orthoptera					
1465	Orthoptera					
1679	Orthoptera					
1711	Orthoptera					
294	Orthoptera					
276	Orthoptera					
278	Orthoptera					
310	Orthoptera					
314	Orthoptera					
485	Orthoptera					
688	Orthoptera					
705	Orthoptera					
708	Orthoptera					
715	Orthoptera					

Spec #	Order	Family	Tax 3	Genus	Species	GR/K
791	Orthoptera					
1010	Orthoptera	Acrididae				
1265	Orthoptera	Acrididae				GR
1337	Orthoptera	Acrididae				
1348	Orthoptera	Acrididae				
1354	Orthoptera	Acrididae				
1360	Orthoptera	Acrididae				
1368	Orthoptera	Acrididae				
1399	Orthoptera	Acrididae				
1441	Orthoptera	Acrididae				
1498	Orthoptera	Acrididae				
1533	Orthoptera	Acrididae				
1545	Orthoptera	Acrididae				
1547	Orthoptera	Acrididae				
1599	Orthoptera	Acrididae				
1619	Orthoptera	Acrididae				
1620	Orthoptera	Acrididae				
1621	Orthoptera	Acrididae				
1666	Orthoptera	Acrididae				
1677	Orthoptera	Acrididae				
1699	Orthoptera	Acrididae				
1762	Orthoptera	Acrididae				
1800	Orthoptera	Acrididae				
890	Orthoptera	Acrididae		<i>Cedarinia</i>		
892	Orthoptera	Acrididae		<i>Cedarinia</i>		
1572	Orthoptera	Acrididae		<i>Cedarinia</i>		
1728	Orthoptera	Acrididae		<i>Gonia</i>		
871	Orthoptera	Acrididae		<i>Goniaea</i>		K
872	Orthoptera	Acrididae		<i>Goniaea</i>		K
940	Orthoptera	Acrididae		<i>Goniaea</i>		
1470	Orthoptera	Acrididae		<i>Goniaea</i>		
174	Orthoptera	Acrididae				K
231	Orthoptera	Acrididae				K
235	Orthoptera	Acrididae				K
501	Orthoptera	Acrididae				K
548	Orthoptera	Acrididae				
613	Orthoptera	Acrididae				
304	Orthoptera	Acrididae		<i>Goniae</i>		K
232	Orthoptera	Acrididae		<i>Goniaea</i>		K
233	Orthoptera	Acrididae		<i>Goniaea</i>		K
293	Orthoptera	Acrididae		<i>Phaulacridium</i>	<i>vitatum</i>	K
681	Orthoptera	Acrididae	Acridinae			
703	Orthoptera	Acrididae	Acridinae			
576	Orthoptera	Acrididae	Catantopinae	<i>Cedarinia</i>		
690	Orthoptera	Acrididae	Catantopinae	<i>Cedarinia</i>		
722	Orthoptera	Acrididae	Catantopinae	<i>Cedarinia</i>		
726	Orthoptera	Acrididae	Catantopinae	<i>Cedarinia</i>		
868	Orthoptera	Acrididae	Oxyinae			
883	Orthoptera	Eumastacidae				
1345	Orthoptera	Eumastacidae				
1469	Orthoptera	Eumastacidae				
1549	Orthoptera	Eumastacidae				
738	Orthoptera	Eumastacidae				
816	Orthoptera	Eumastacidae				

Spec #	Order	Family	Tax 3	Genus	Species	GR/K
834	Orthoptera	Gryllidae				
857	Orthoptera	Gryllidae				
1333	Orthoptera	Gryllidae				
1349	Orthoptera	Gryllidae				
1508	Orthoptera	Gryllidae				
1604	Orthoptera	Gryllidae				
1617	Orthoptera	Gryllidae				
1653	Orthoptera	Gryllidae				
1702	Orthoptera	Gryllidae				
180	Orthoptera	Gryllidae				K
216	Orthoptera	Gryllidae				
608	Orthoptera	Gryllidae				
609	Orthoptera	Gryllidae				
618	Orthoptera	Gryllidae				
672	Orthoptera	Gryllidae				
729	Orthoptera	Gryllidae				
809	Orthoptera	Gryllidae				
811	Orthoptera	Gryllidae		<i>Apterogryllus</i>		
555	Orthoptera	Gryllidae				
782	Orthoptera	Pyrgomorphidae				
524	Orthoptera	Stenopelmatidae				
1008	Orthoptera	Stenopelmatidae				
1043	Orthoptera	Stenopelmatidae				
1557	Orthoptera	Stenopelmatidae				
1582	Orthoptera	Stenopelmatidae				
526	Orthoptera	Stenopelmatidae		<i>Onosandrus</i>		K
881	Orthoptera	Tetrigidae				
713	Orthoptera	Tetrigidae				
873	Orthoptera	Tettigoniidae				
882	Orthoptera	Tettigoniidae				
897	Orthoptera	Tettigoniidae				
902	Orthoptera	Tettigoniidae				
903	Orthoptera	Tettigoniidae				K
931	Orthoptera	Tettigoniidae				
980	Orthoptera	Tettigoniidae				
988	Orthoptera	Tettigoniidae				
994	Orthoptera	Tettigoniidae				
1009	Orthoptera	Tettigoniidae				
1013	Orthoptera	Tettigoniidae				
1026	Orthoptera	Tettigoniidae				
1039	Orthoptera	Tettigoniidae				
1052	Orthoptera	Tettigoniidae				
1326	Orthoptera	Tettigoniidae				
1327	Orthoptera	Tettigoniidae				
1332	Orthoptera	Tettigoniidae				
1342	Orthoptera	Tettigoniidae				
1346	Orthoptera	Tettigoniidae				
1386	Orthoptera	Tettigoniidae				
1483	Orthoptera	Tettigoniidae				
1485	Orthoptera	Tettigoniidae				
1487	Orthoptera	Tettigoniidae				
1539	Orthoptera	Tettigoniidae				
1622	Orthoptera	Tettigoniidae				
1644	Orthoptera	Tettigoniidae				

Spec #	Order	Family	Tax 3	Genus	Species	GR/K
1675	Orthoptera	Tettigoniidae				
1695	Orthoptera	Tettigoniidae				
1736	Orthoptera	Tettigoniidae				
1739	Orthoptera	Tettigoniidae				
1749	Orthoptera	Tettigoniidae				
1750	Orthoptera	Tettigoniidae				
1779	Orthoptera	Tettigoniidae				
106	Orthoptera	Tettigoniidae				K
118	Orthoptera	Tettigoniidae				
149	Orthoptera	Tettigoniidae				K
167	Orthoptera	Tettigoniidae				
202	Orthoptera	Tettigoniidae				
218	Orthoptera	Tettigoniidae				K
246	Orthoptera	Tettigoniidae				K
268	Orthoptera	Tettigoniidae				
1754	Orthoptera	Tettigoniidae	Phasmatidinae			
1207	Orthoptera	Tettigoniidae	Phasmodinae			
973	Phasmatodea					
1080	Phasmatodea					
1220	Phasmatodea					
1223	Phasmatodea					
1314	Phasmatodea					
1520	Phasmatodea					
1705	Phasmatodea					
1715	Phasmatodea					
1733	Phasmatodea					
785	Phasmatodea					
303	Phasmatodea					
458	Phasmatodea					
1385	Platyhelminthes					
1404	Platyhelminthes					
1568	Platyhelminthes					
521	Platyhelminthes					
879	Scorpionida					
880	Scorpionida					
965	Scorpionida					
1451	Scorpionida					
1592	Scorpionida					
1600	Scorpionida					
1796	Scorpionida					
568	Scorpionida				Scorpion sp1	K
469	Scorpionida				Scorpion sp2	K
629	Scorpionida				Scorpion sp3	K
1042	Trichoptera					GA
69	Trichoptera					GA
144	Trichoptera					GA
145	Trichoptera					GA
151	Trichoptera					GA

BIRDS

Graeme Liddelow and Chris Vellios

Introduction

The object of recording birds in FORESTCHECK is to monitor the impacts of logging and associated burning on bird species composition and abundance. This is achieved by:

- Recording species richness and abundance within each treatment (coupe buffer, shelterwood and gap release) and uncut controls.
- Comparing species richness and abundance between each treatment
- Analysing trends within species between treatments

Monitoring

All Wellington East FORESTCHECK grids were monitored for diurnal birds in the spring of 2004. Science Division personnel were able to undertake all the bird census work in 2004. Both sight and sound were used to identify the birds, and the census method is described in the FORESTCHECK Operating Plan.

Preliminary Results and Discussion

Diurnal Birds

A total of 29 species of birds, comprising 487 individuals, were recorded in the ten grids. Ten or more individuals were recorded in 14 species (Table 1). For each of the 3 treatment grids, there were 25 species and 155 individuals in the controls, 21 species and 178 individuals in the shelterwood treatment and 20 species and 118 individuals in the gap release treatment. The single coupe buffer had 14 species and 36 individuals.

The overall density of birds in this year's census was 9.7 birds/ha. In previous years it was 5.3 at Donnelly (2001), 15.4 at Wellington (2002) and 9.7 at Perth Hills (2003). The 9.7 birds/ha at Wellington East is similar to the ongoing Kingston Study in an "average" year with 10.4 birds/ha.

Seven species of birds occurred in only one treatment (either control, coupe buffer, shelterwood or gap release, see Table 1), however, none of these would be expected to be restricted to that treatment.

Table 1. Bird species and number of individuals recorded by treatment in the Wellington East FORESTCHECK grids.

RAOU No.	Common Name	Scientific Name	Total	Ext Control	Coupe Buffer	Shelter-wood	Gap Release
264	Forest Red-tailed Black Cockatoo	<i>Calyptorhynchus banksii naso</i>	3	3			
290	Red-capped Parrot	<i>Platycerus spurius</i>	13			13	
294	Australian Ringneck	<i>Platycerus zonarius</i>	10	5		5	
322	Laughing Kookaburra	<i>Dacelo novaeguineae</i>	1		1		
337	Pallid Cuckoo	<i>Cuculus pallidus</i>	3	1		1	1
344	Shining Bronze-Cuckoo	<i>Chrysococcyx lucidus</i>	9	3		4	2
361	Grey Fantail	<i>Rhipidura fuliginosa</i>	21	11	2	6	2
380	Scarlet Robin	<i>Petroica multicolor</i>	30	9	2	6	13
394	Western Yellow Robin	<i>Eopsaltria griseogularis</i>	11	2	1	5	3
398	Golden Whistler	<i>Pachycephala pectoralis</i>	26	7	7	8	4
408	Grey Shrike-thrush	<i>Colluricincla harmonica</i>	11	3	1	6	1
424	Black-faced Cuckoo-shrike	<i>Coracina novaehollandiae</i>	4	2		1	1
463	Western Gerygone	<i>Gerygone fusca</i>	61	20	4	21	16
465	Weebill	<i>Smicornis brevirostris</i>	1		1		
472	Western Thornbill	<i>Acanthiza inornata</i>	32	7		6	19
476	Broad-tailed (Inland) Thornbill	<i>Acanthiza apicalis</i>	94	28	6	41	19
488	White-browed Scrubwren	<i>Sericornis frontalis</i>	1	1			
549	Varied Sitella	<i>Daphoenositta chrysoptera</i>	25	7		12	6
556	Rufous Treecreeper	<i>Climacteris rufa</i>	10	2	2	6	
565	Spotted Pardalote	<i>Pardalotus punctatus</i>	5	1		3	1
574	Silvereye	<i>Zosterops lateralis</i>	7	1		4	2
578	Western White-naped Honeyeater	<i>Melithreptus chloropsis</i>	5	1	1	2	1
592	Western Spinebill	<i>Acanthorhynchus superciliosus</i>	17	4		6	7
608	Singing Honeyeater	<i>Lichenostomus virescens</i>	1	1			
638	Red Wattlebird	<i>Anthochaera carunculata</i>	1				1
697	Grey Currawong	<i>Sterpera versicolor</i>	6	1	2	2	1
710	Western Little Wattlerbird	<i>Anthochaera lunulata</i>	6	4			2
930	Australian Raven	<i>Corvus coronoides</i>	2	1	1		
976	Striated Pardalote	<i>Pardalotus striatus</i>	71	30	5	20	16
Total Individuals			487	155	36	178	118
Total Species			29	25	14	21	20

Table 2. Bird species and number of individuals recorded by forest block in the Wellington East FORESTCHECK grids.

RAOU No.	Common Name	Scientific Name	Total	Nalyerin	Godfrey	Stockyard	Bell
264	Forest Red-tailed Black Cockatoo	<i>Calyptrorhynchus banksii naso</i>	3				3
290	Red-capped Parrot	<i>Platycerus spurius</i>	13				13
294	Port Lincoln Ringneck	<i>Platycerus zonarius</i>	10		2	4	4
322	Laughing Kookaburra	<i>Dacelo novaeguineae</i>	1		1		
337	Pallid Cuckoo	<i>Cuculus pallidus</i>	3	1		2	
344	Shining Bronze-Cuckoo	<i>Chrysococcyx lucidus</i>	9	2	3	2	2
361	Grey Fantail	<i>Rhipidura fuliginosa</i>	21	4	2	5	10
380	Scarlet Robin	<i>Petroica multicolor</i>	30	2	10	8	10
394	Western Yellow Robin	<i>Eopsaltria griseogularis</i>	11		5	2	4
398	Golden Whistler	<i>Pachycephala pectoralis</i>	26	4	11	4	7
408	Grey Shrike-thrush	<i>Colluricincla harmonica</i>	11	1	3	3	4
424	Black-faced Cuckoo-shrike	<i>Coracina novaehollandiae</i>	4				4
463	Western Gerygone	<i>Gerygone fusca</i>	61	7	16	18	20
465	Weebill	<i>Smicornis brevirostris</i>	1		1		
472	Western Thornbill	<i>Acanthiza inornata</i>	32	2	4	15	11
476	Broad-tailed (Inland) Thornbill	<i>Acanthiza apicalis</i>	94	13	37	24	20
488	White-browed Scrubwren	<i>Sericornis frontalis</i>	1			1	
549	Varied Sitella	<i>Daphoenositta chrysoptera</i>	25		4	1	20
556	Rufous Treecreeper	<i>Climacteris rufa</i>	10		2	2	6
565	Spotted Pardalote	<i>Pardalotus punctatus</i>	5		2		3
574	Silvereye	<i>Zosterops lateralis</i>	7	1	2	2	2
578	Western White-naped Honeyeater	<i>Melithreptus chloropsis</i>	5		1	1	3
592	Western Spinebill	<i>Acanthorhynchus superciliosus</i>	17	2		6	9
608	Singing Honeyeater	<i>Lichenostomus virescens</i>	1	1			
638	Red Wattlebird	<i>Anthochaera carunculata</i>	1				1
697	Grey Currawong	<i>Sterpera versicolor</i>	6	1	3	1	1
710	Western Little Wattlerbird	<i>Anthochaera lunulata</i>	6	1			5
930	Australian Raven	<i>Corvus coronoides</i>	2		1		1
976	Striated Pardalote	<i>Pardalotus striatus</i>	71	8	18	23	22
Total Species			29	15	20	19	24
Total Individuals			487	50	128	124	185

All 13 records of the Red-capped Parrot (*Platycerus spurius*) were recorded in the shelterwood at Bell forest block, but there appears to be no obvious reason as to why. Of the small scrub birds only one individual, a White-browed Scrubwren (*Sericornis frontalis*), was recorded in Stockyard external control. There were no other small species, such as Fairy-wrens, recorded. This is to be expected, as the majority of the scrub in the grids is only about 20 cm or lower.

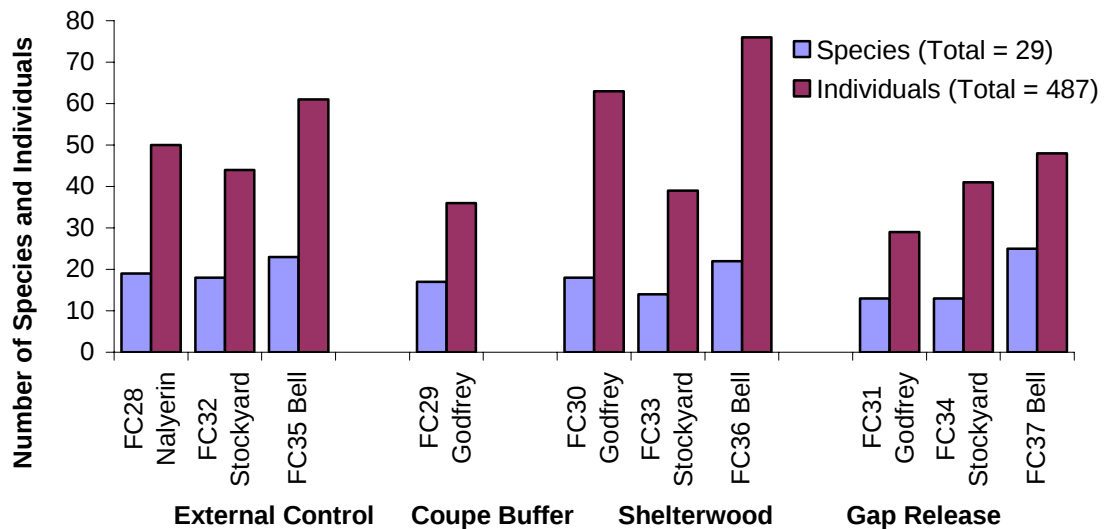


Figure 1. The number of bird species and individuals recorded in each FORESTCHECK grid in Wellington East.

In previous years we have stated the importance of the Kingston jarrah bird study for monitoring the annual changes that occur following logging and regeneration. In 2005, The Kingston study will have been going for 10 years, and as the FORESTCHECK sites are monitored on a 5-6 year rotation it will be important to keep the Kingston study going to document any changes that could occur, especially after crown separation occurs.

Nocturnal Birds

Systematic survey of the nocturnal birds was not carried in this round of monitoring due to unsuitable weather, and unavailability of personnel. However, one night's survey was conducted (Fig. 2) and opportunistic records were kept. A past survey of owls in the southwest by Liddelow *et.al* (2002) suggests that most owls are common in this area.



Figure 2. Results of a single night's survey for nocturnal birds and bats in Wellington East.

During the course of other monitoring, such as the mammal spotlighting, owls were heard calling or were seen. Southern Boobook Owls (*Ninox novaeseelandiae*) were heard at all sites. One Masked Owl (*Tyto novaehollandiae*) was heard calling in Bell forest block in jarrah-wandoo semi-woodland between the shelterwood and external control sites. A number of Masked Owls were seen on Boundary Rd. One individual was seen twice, 1km south of Alamo Road, along the private property boundary (which is a preferred hunting zone for Masked owls) and another was heard in a wandoo gully 1 km north of Mistle Road. Australian Owlet-nightjars (*Aegotheles cristatus*) were commonly seen on the roads between sites at night when travelling for the spotlight surveys and Tawny Frogmouths (*Podargus strigoides*) were also seen at all sites. These species all appear to be common in this forest habitat.

Reference

Liddel, G.L., Wheeler, I.B. and Kavanagh, R.P. 2002. Owls in the southwest forest of Western Australia. Pp. 233-241 In, Newton, I., Kavanagh, R., Olsen, J., Tayler, I. (eds). *Ecology and Conservation of Owls*. CSIRO, Collingwood.

MAMMALS AND HERPETOFAUNA

Graeme Liddelow

Introduction

The object of recording mammals and herpetofauna in FORESTCHECK is to monitor the impacts of logging and associated burning on species status and abundance. This is achieved by:

- Trapping and recording the suite of medium and small sized mammals, reptiles and amphibians on each FORESTCHECK grid
- Recording the presence of small mammals in nest boxes placed within each grid
- Comparing species richness, abundance, sex ratios and trap percentages between treatments within and between FORESTCHECK sites
- Recording the presence of the larger mammals along set transects that cover all treatments of the FORESTCHECK sites on a landscape basis
- Recording the presence of nocturnal mammals by spotlighting along set transects that cover all the treatments of the FORESTCHECK sites

The species and abundance of feral animals is also recorded with the use of sand pads placed at regular intervals along pre-determined tracks and roads within each FORESTCHECK site.

Monitoring

Trapping and spotlighting was carried out on all grids in spring 2004 and autumn 2005. Spring monitoring went according to plan, with only one day of rain that disrupted the road and spotlight surveys on that day but had little impact on the following day's trapping. However, autumn monitoring was disrupted by constant rain, which resulted in the pit traps being opened for only 2 of the 4 nights. The sand pad survey had to be abandoned in autumn as the pads were hard and wet and could not be interpreted.

Voucher Specimens

Thirty individual specimens were lodged with the WA Museum. This number would have been higher if there was not the inclement weather in autumn.

The specimens included 24 skinks with 7 species, 1 snake, 1 dragon and 4 species of amphibian (Table 1).

Results

Trapping

A total of 173 individuals were trapped in the spring and autumn sessions (Table 2) with 107 individuals trapped in spring and 66 in autumn.

There were 28 mammals, 71 reptiles and 8 amphibians trapped in spring and 63 mammals, 2 reptiles and 1 amphibian trapped in autumn (Table 2 and Fig. 1). The results clearly show the lack of reptile and amphibian captures due to the inclement weather conditions in autumn.

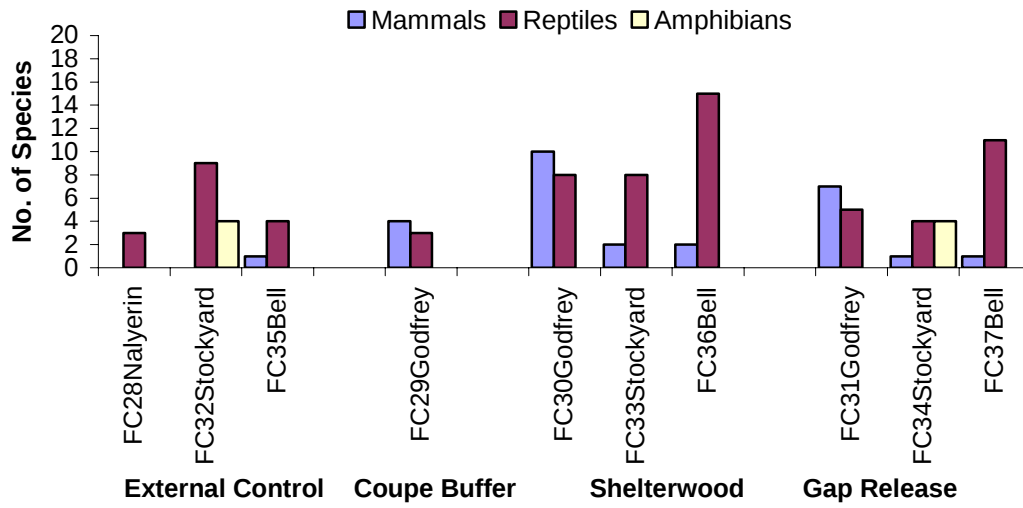
Table 1. Specimens from the Wellington East FORESTCHECK grids that were lodged with the W.A. Museum.

Field #	Museum #	Museum Name
FC145	156646	<i>Lerista distinguenda</i>
FC146	Discard	<i>Morethia obscura</i>
FC147	156647	<i>Lerista distinguenda</i>
FC148	156651	<i>Morethia obscura</i>
FC149	156650	<i>Lerista distinguenda</i>
FC150	156649	<i>Egernia napoleonis</i>
FC151	156648	<i>Ctenotus labillardieri</i>
FC152	Discard	<i>Lerista distinguenda</i>
FC153	156652	<i>Morethia obscura</i>
FC154	156653	<i>Morethia obscura</i>
FC155	156654	<i>Morethia obscura</i>
FC156	156659	<i>Limnodynastes dorsalis</i>
FC157	156658	<i>Pseudophryne guentheri</i>
FC158	156656	<i>Hemiergis initialis initialis</i>
FC159	156655	<i>Morethia obscura</i>
FC160	156657	<i>Crinia georgiana</i>
FC161	156661	<i>Pogona minor minor</i>
FC162	Discard	<i>Hemiergis initialis initialis</i>
FC163	156660	<i>Morethia obscura</i>
FC164	156662	<i>Lerista distinguenda</i>
FC165	156644	<i>Egernia napoleonis</i>
FC166	156645	<i>Morethia obscura</i>
FC167	156643	<i>Ctenotus delli</i>
FC168	156642	<i>Heleioporus barycragus</i>
FC169	156663	<i>Morethia obscura</i>
FC170	156664	<i>Menetia greyii</i>
FC171	156665	<i>Ramphotyphlops australis</i>
FC172	Discard	<i>Hemiergis initialis initialis</i>
FC173	156666	<i>Lerista distinguenda</i>
FC174	156667	<i>Lerista distinguenda</i>

Table 2. The species and abundance of mammals, reptiles and amphibians recorded in traps within treatments in Wellington East.

Species	External Control				Coupe Buffer				Shelterwood				Gap release			
	Spring		Autumn		Spring		Autumn		Spring		Autumn		Spring		Autumn	
	Pit	Wire	Pit	Wire	Pit	Wire	Pit	Wire	Pit	Wire	Pit	Wire	Pit	Wire	Pit	Wire
MAMMALS																
<i>Antechinus flavipes</i>				3								1		1		2
<i>Bettongia penicillata ogilbyi</i>						3		4		8		12		7		12
<i>Cercartetus concinnus</i>												1				
<i>Dasyurus geoffroii</i>								4		1		4				4
<i>Sminthopsis gilbertii</i>	1								3				1			
<i>Tachyglossus aculeatus</i>												1				1
<i>Trichosurus vulpecula</i>						1		8		2		6				
REPTILES																
<i>Ctenotus catenifer</i>														1		
<i>Ctenotus labillardieri</i>									1							
<i>Egernia napoleonis</i>									3					2		
<i>Hemiergis initialis</i>	1								3							
<i>Hemiergis peroni</i>												2				
<i>Lerista distinguenda</i>	6					2			12					11		
<i>Menetia greyii</i>	1								1					2		
<i>Morethia linocellata</i>	3								1					3		
<i>Morethia obscura</i>	5								6					1		
<i>Pogona minima</i>						1			1							
<i>Ramphotyphlops australis</i>									1							
<i>Tiliqua rugosa</i>										2					1	
AMPHIBIANS																
<i>Crinia georgiana</i>	1															
<i>Heleioporus eyrei</i>														4		1
<i>Limnodynastes dorsalis</i>	2															
<i>Pseudophryne guentheri</i>	1															
TOTAL (173)	21	0	0	3	3	4	0	16	32	13	3	24	25	9	1	19
TREATMENT TOTALS		24				23				72				54		

(a) Spring – all traps



(b) Autumn – all traps

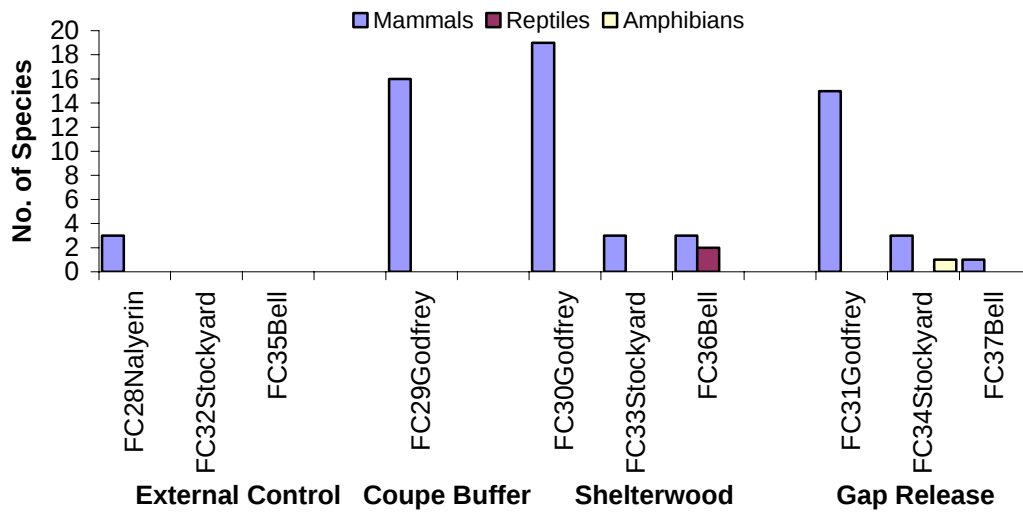


Figure 1. The number of individual animals, reptiles and amphibians recorded in traps in spring (a) and autumn (b) in the Wellington East FORESTCHECK grids.

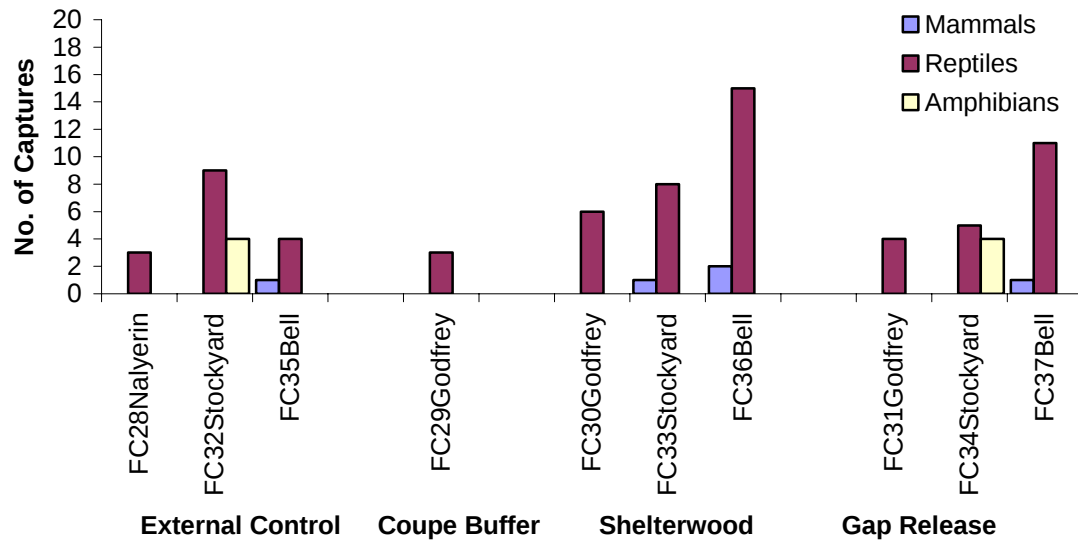
The highest total numbers of captures occurred in the shelterwood treatment followed by the gap release treatment and the lowest numbers occurred in the external controls (Table 3). There was only one coupe buffer treatment (compared to 3 each for the other treatments) therefore no comparisons are made to this treatment.

Table 3. The species and abundance of mammals, reptiles and amphibians recorded in traps by block on the Wellington East FORESTCHECK grids in 2004-05. NB. The shaded area highlights the higher number of mammals recorded in the Godfrey forest block, which is baited for foxes in the Western Shield program.

Species	Nalyerin/Godfrey				Stockyard			Bell			TOTAL
	Ext. Control	Coupe Buffer	Shelterwood	Gap Release	Ext. Control	Shelterwood	Gap Release	Ext. Control	Shelterwood	Gap Release	
MAMMALS											
<i>Antechinus flavipes</i>	3					1	3				7
<i>Bettongia penicillata</i>		7	18	18		2	1				46
<i>Cercartetus concinnus</i>					1						1
<i>Dasyurus geoffroii</i>		4	3	3					2	1	13
<i>Sminthopsis gilbertii</i>						1		1	2	1	5
<i>Tachyglossus aculeatus</i>				1					1		2
<i>Trichosurus vulpecula</i>		9	8								17
REPTILES											
<i>Ctenotus catenifer</i>							1				1
<i>Ctenotus labillardieri</i>			1								1
<i>Egernia napoleonis</i>			2			1	2				5
<i>Hemiergis initialis</i>					1	3					4
<i>Hemiergis peroni</i>									2		2
<i>Lerista distinguenda</i>	2	2	2	3	3	1		1	9	8	31
<i>Menetia greyii</i>							2	1	1		4
<i>Morethia linocellata</i>					2			1	1	3	7
<i>Morethia obscura</i>	1		1	1	3	2		1	3		12
<i>Pogona minima</i>		1				1					2
<i>Ramphotyphlops australis</i>									1		1
<i>Tiliqua rugosa</i>			2	1							3
AMPHIBIANS											
<i>Crinia georgiana</i>					1						1
<i>Heleioporus eyrei</i>							5				5
<i>Limnodynastes dorsalis</i>					2						2
<i>Pseudophryne guentheri</i>					1						1
Total	6	23	37	27	13	13	14	5	22	13	173

Pit trapping was very successful in the spring, but was interrupted by inclement weather in autumn. A total of 81 individuals were trapped in spring and only 3 in the autumn. The shelterwood treatment had the highest numbers followed by the gap release then the external controls (Fig. 2). All the reptiles, except the bobtailed lizard (*Tiliqua rugosa*), and all the amphibians were caught in pit traps.

(a) Spring – pit traps



(b) Autumn – pit traps

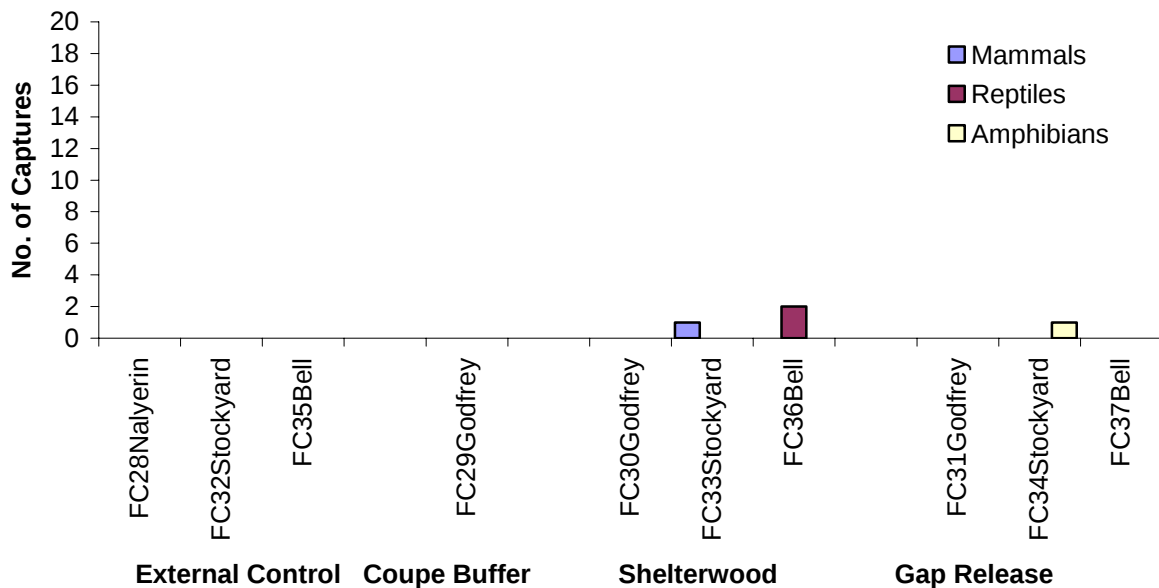
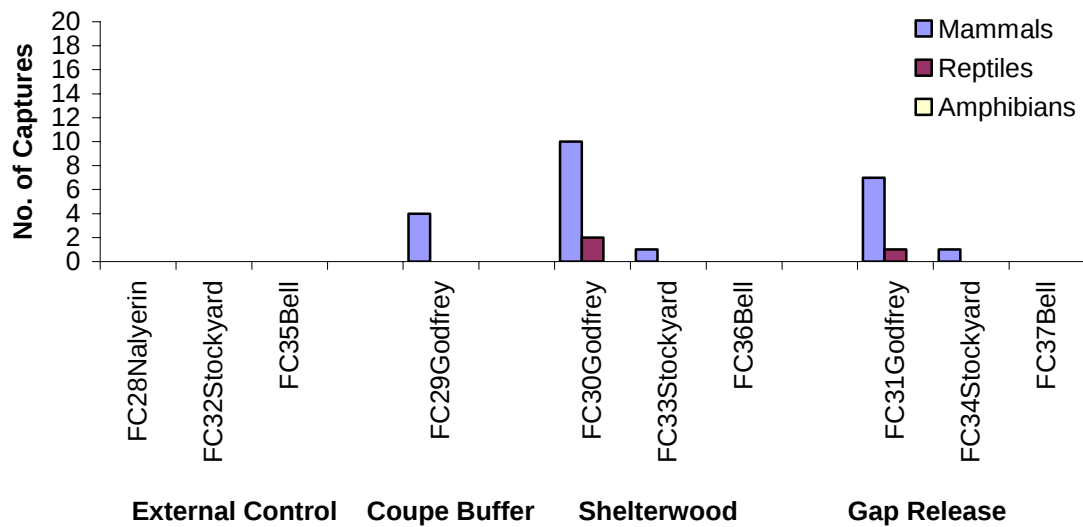


Figure 2. The number of individual mammals, reptiles and amphibians recorded in pit traps in the spring (a) and autumn (b) on the Wellington East FORESTCHECK grids.

Wire traps were very successful, especially in the Godfrey forest block treatments where excellent captures of Woylies (*Bettongia penicillata*), Brush-tailed Possums (*Trichosurus vulpecula*) and a Chuditch (*Dasyurus geoffroii*) were recorded. The autumn trapping session was more successful than spring (Fig. 3). The woylie captures at Stockyard forest block resulted from one individual being caught 3 times. This was a survivor from a woylie release in Stene forest block, to the east of Stockyard, some years before. The results support the success of the Western Shield fox-baiting program. Godfrey forest block is baited under the Western Shield program, but the sites north of the Williams-Collie Rd in Nalyerin, Stockyard and Bell forest blocks are not.

(a) Spring – wire traps



(b) Autumn – wire traps

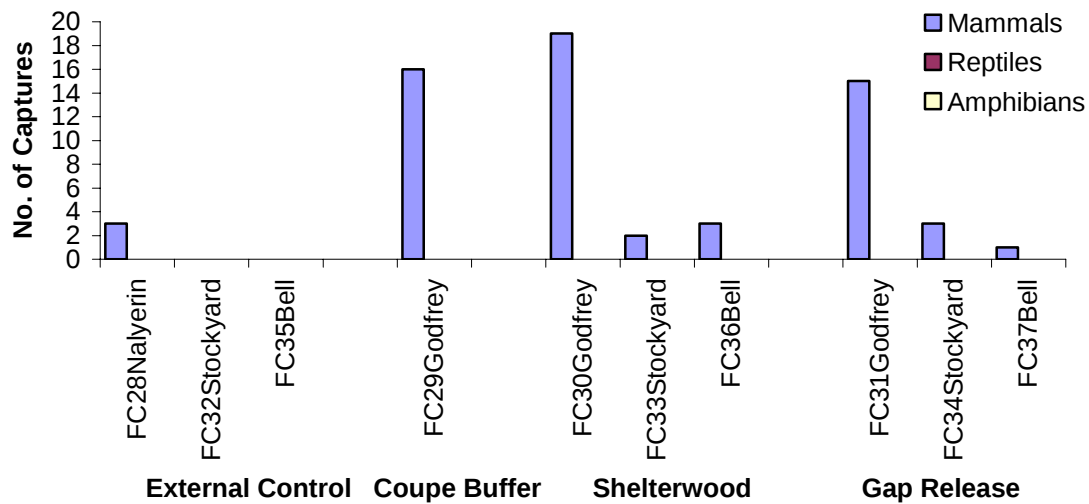


Figure 3. The number of individual mammals, reptiles and amphibians recorded in wire traps in the spring (a) and autumn (b) on the Wellington East FORESTCHECK grids.

Sandpads

Sand pads are installed and monitored on a landscape scale with pads at 1 km intervals over a distance of 50 km. Monitoring was carried out in spring 2004 but was not able to be undertaken in autumn 2005 due to inclement weather.

Recording the presence of feral animals is the main objective when monitoring sand pads, so it was gratifying not to record any fox activity. Native mammals, including kangaroos, wallabies, woylies and chuditch, accounted for 62% of the animals recorded on the sand pads. The other mammals were 2 rabbits and one pig (8%). Birds accounted for 18% of the records and the remaining 2% were reptiles (Fig. 4).

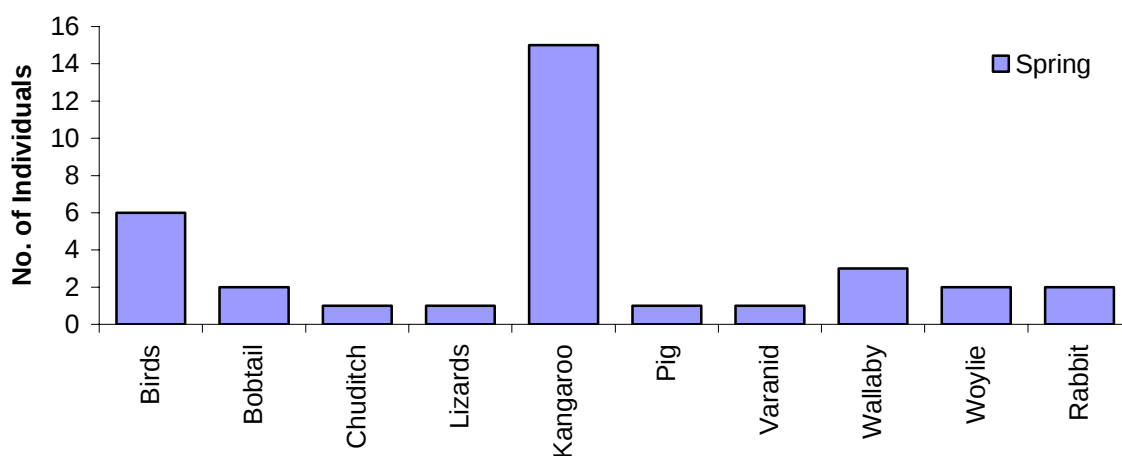


Figure 4. The number of species recording contacts on sand pads in the spring at the Wellington East FORESTCHECK location.

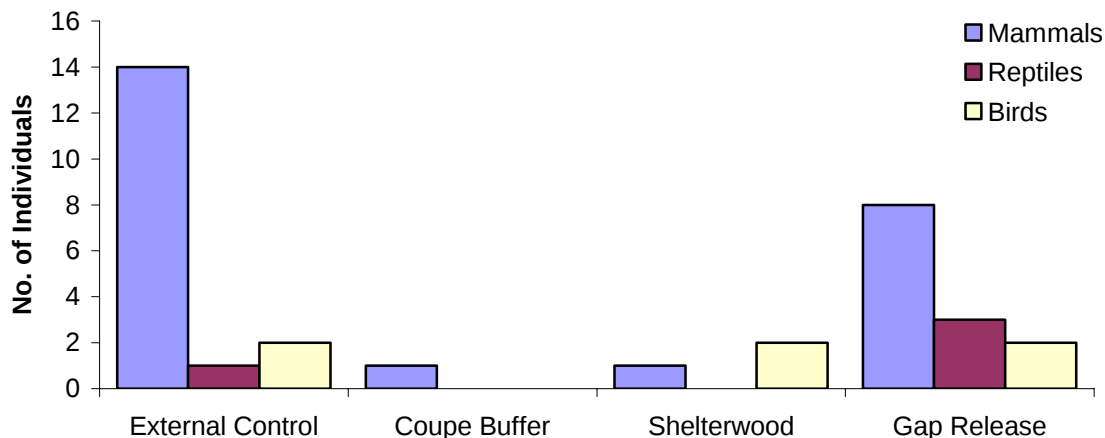


Figure 5. The number of individual animals recorded by treatment on sand pads in the spring at the Wellington East FORESTCHECK location.

Most activity was recorded in the vicinity of external controls followed by the gap release treatments and then the shelterwoods (Fig. 5).

Road Survey

Road surveys were conducted in spring 2004 and autumn 2005, and are also undertaken on a landscape basis, with the emphasis on differences between blocks as the treatments within the blocks are close together. The distance travelled this year was 42 km.

Only 2 species were recorded on these surveys, the Western Grey Kangaroo (*Macropus fuliginosus*) and the Western Brush Wallaby (*Macropus irma*) with the kangaroo dominating the counts (Fig. 6). The higher numbers of kangaroos in Stockyard forest block in spring is unexplained, as it was assumed they would be higher in the Nalyerin/Godfrey area with a younger fire successional stage and with more succulent regrowth and understorey plants for food.

No Western Brush Wallabies were recorded in Bell forest block on either spring or autumn surveys however one was seen during the spotlighting surveys (Fig. 7b). The country is similar to both Stockyard and Nalyerin/Godfrey forest blocks and it would be expected to see more of them there. The numbers of brush wallabies are similar to the eastern Donnelly FORESTCHECK sites (which were monitored in 2001-02).

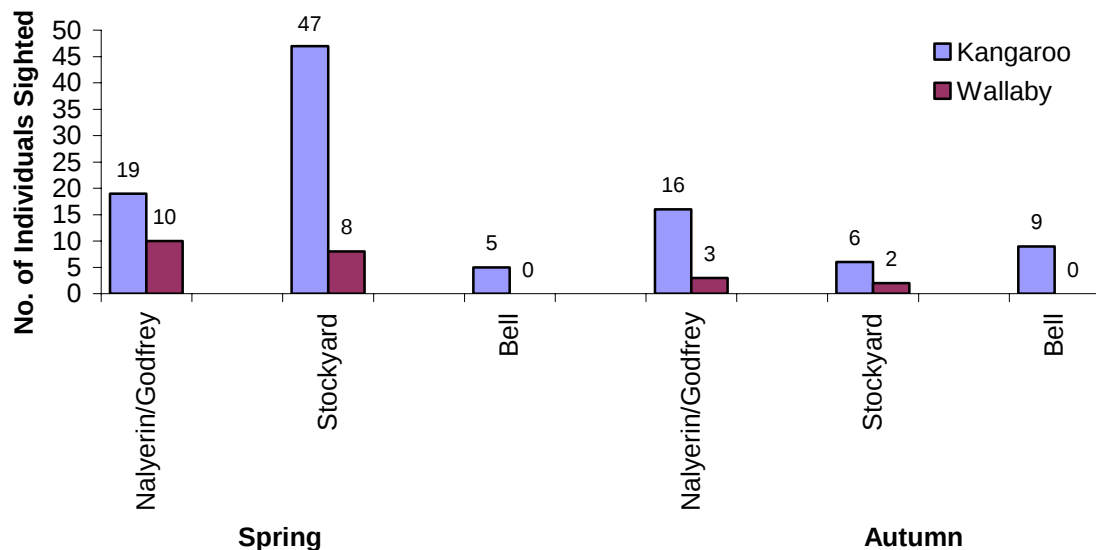
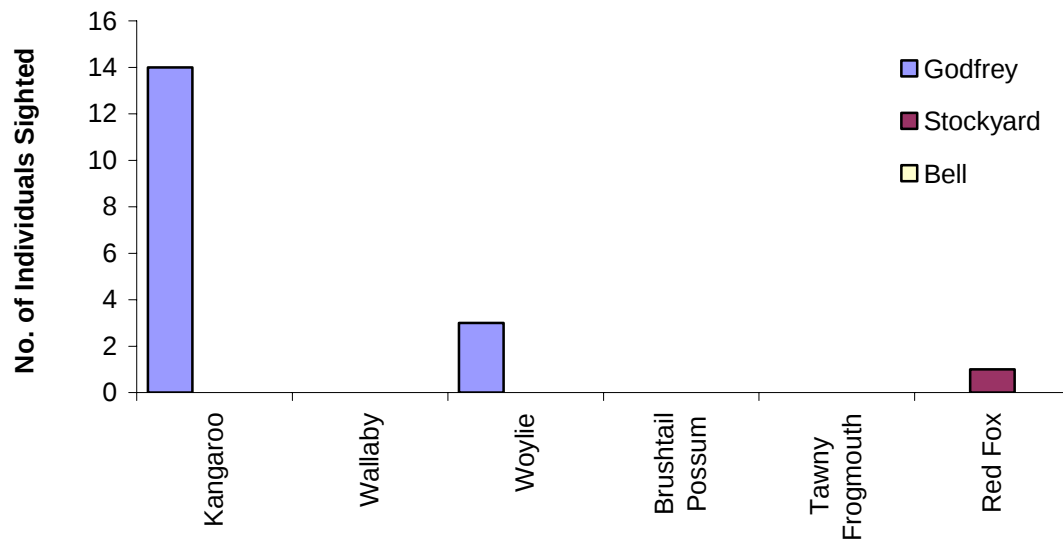


Figure 6. The number of individual animals recorded by block during road surveys in the spring and autumn within the Wellington East FORESTCHECK location.

Spotlighting

A total of 19 kangaroos, 10 wallabies, 4 woylies, 1 brushtailed possum, 2 Tawny Frogmouths and 1 red fox were recorded during spotlighting (fig 7). All the animals except 1 kangaroo and 2 brush wallabies and the fox were recorded in Godfrey forest block. The red fox was recorded in Stockyard forest block.

(a) Spring - Spotlighting



(b) Autumn - Spotlighting

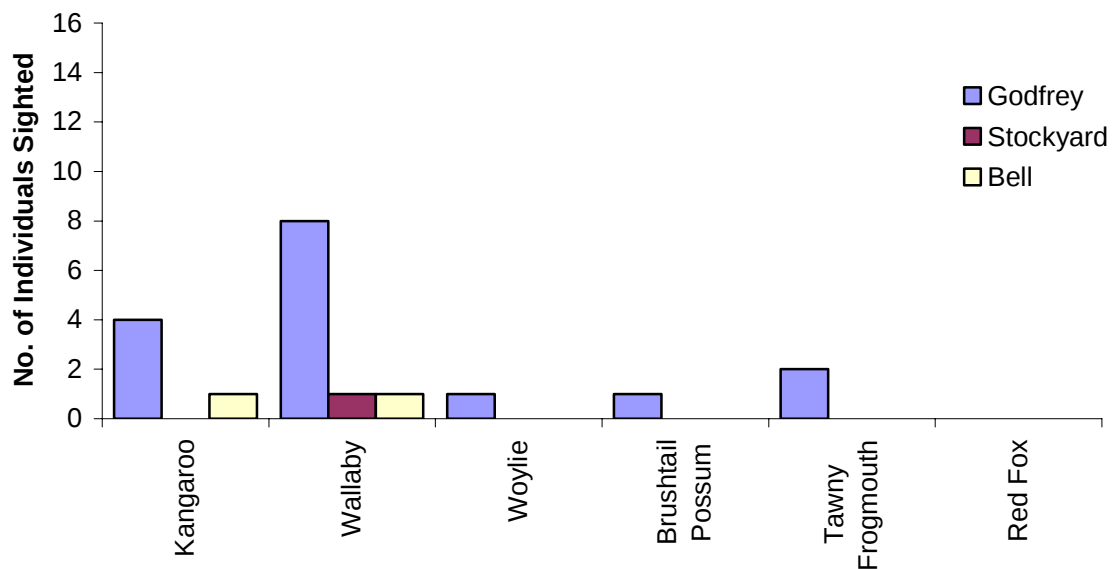


Figure 7. The number of individual animals recorded during spotlighting in the spring (a) and autumn (b) within the Wellington East FORESTCHECK location.

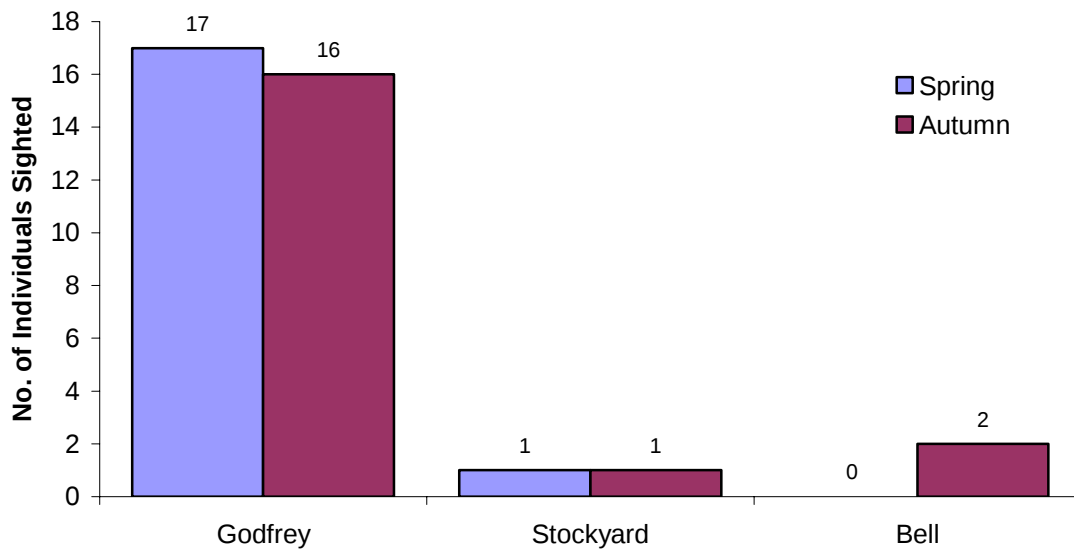


Figure 8. The number of animals recorded by block during spotlighting in the spring and autumn within the Wellington East FORESTCHECK location.

Conclusions

- The results support those from elsewhere (e.g. Perup, Kingston and Batalling) that fox baiting has a significant impact on the number of native animals recorded.
- The wet weather had a significant impact on small mammal and reptile trapping in the autumn.
- High numbers of animals were trapped on the logged treatments (shelterwood and gap release).

DATA MANAGEMENT AND STORAGE

Amanda Mellican and Verna Tunsell

Introduction

This group is responsible for entering the collected data into electronic format for Macro Vertebrates, Diurnal Birds, Nocturnals, Mammals and Herpetofauna, Vascular Plants, Macrofungi and Cryptogams, and the collected voucher specimens (Flora, Cryptogams and Macrofungi), and obtaining the electronic data from the remaining groups.

Data Entry

An excel program applying Visual Basic was developed for each of the survey sheets. There are 2 parts in the program: Data entry and Data correction. The aim is to save time and to reduce typing errors during the data entry process.

As an example, in the Trapping Field Data Sheet, there are 12 fields for the data entry (Location, Treatment, Personnel, Date, Trap point, Species, Weight, Tag #1, Tag #2, Sex, Breeding Condition and Comments). Places of location, names of treatment, names of personnel and gender are known and they are created as drop-down lists. As for the species, common species name was pre-listed in the program. Date as in Day, Month and Year (from 2001 to 2010) was also created as drop down lists. Thus, location, treatment and personnel are only entered once for all the records for a particular data sheet. If there was nothing to comment on, then the program will automatically record in the Comments section as “No comment”. If any one of the fields is missing or left as blank, an error message is given and the data would not be inserted into data file until all the fields are selected or filled. Species code, scientific name and common name are also automatically recorded into the data file depending upon the selected common species. Record number is automatically written into the data file.

To date, all the survey sheets have been completed by Verna Tunsell. A metadata form as shown in Appendix A is also completed.

Data Validation

Amanda Mellican validates the data for all the groups that we are responsible for. The validation date will be recorded in the metadata form. Then a DESCRIPTIONS file (which indicates the lists of an individual field, and codes and descriptions of an individual field), and the validated DATA file will be sent to the leader of the individual group.

Data Storage

All the individual sampling data will be saved and backed up as individual files on the network drive. The data are saved and secured since the network drive is backed up at COB everyday. The final version of the validated data will be printed and kept in a filing cabinet and eventually archived with the library at the completion of the project.

Collected Specimens

The 166 Vascular Plants and 413 Cryptogam specimens collected during the period, have been identified (as far as possible), prepared, and lodged at the WA Herbarium. The fungi season was late this year due to the weather and the 138 collected voucher specimens are almost ready to be processed. The fungi collection is housed at the Tony Annels Herbarium in Manjimup to enable further work to be carried out.

Each specimen is allocated its own unique barcode so that each specimen is readily located by electronic means or by physical means as required.

Vascular plant specimens are pressed and dried, then mounted, with specialised herbarium tape, on card, and placed in separate folders.

Cryptogams are dried (friable specimens are stabilised with emulsion), placed on a card with adhesive to keep the specimen together (Mosses are washed prior to drying to remove debris). The specimens are then secured in cardboard boxes to prevent damage.

Fungi specimens are also dried and placed into greaseproof paper, then into ziplock bags, those small enough then go into similar boxes to cryptogams, the larger remain unboxed.

The specimens that have been lodged at the WA Herbarium have also been databased on the Max system and submitted electronically. Max was developed by Simon Woodman and Paul Gioia and is used as the primary means of submitting specimen information to the Herbarium. There are many facets to Max, the sections used for FORESTCHECK are the collecting book, specimen tables and reporting facilities. (Examples attached).

Appendix A. Example of Metadata Form.

Group Name →

Leader →

Contact Officer →

No	File Name	File Size (KB)	File Type	Date (completed)	Name of Data Entry Person	Validated Date

Appendix B. Example of Flora Labels generated in Max.

WESTERN AUSTRALIAN HERBARIUM, PERTH Flora of Western Australia

Stackhousia monogyna Labill.

Stackhousiaceae

Identified by:

Erect open perennial shrub, height to 40 cm, width to 40 cm; flowers white. Population flowering 30%. State forest; hill slope; gravelly brown sandy clay. frequent. Forest with associated vegetation of *Eucalyptus marginata*, *Allocasuarina fraseriana*, *Macrozamia riedlei*, *Hakea lissocarpa*, *Hibbertia commutata* and a few *Corymbia calophylla*.

Loc.: Forest monitoring plot FC30, Godfrey forest block, 2.1 km along Steed Road from Ernie Road then left on shunt,

Lat. 33° 16' 2"S **Long.** 116° 25' 25"E (WGS84)

Coll. B.G. Ward FC670 **Date:** 05/10/2004

Voucher: Forestcheck Monitoring Program.

PERTH 6643337

WESTERN AUSTRALIAN HERBARIUM, PERTH Flora of Western Australia

Trymalium ledifolium Fenzl

Rhamnaceae

Identified by:

Erect compact perennial shrub, height to 1 m, width to 90 cm. Population flowering 100%. State forest; hill slope; gravelly brown clayey sand. occasional. Forest with associated vegetation of *Eucalyptus marginata*, *Allocasuarina fraseriana*, *Macrozamia riedlei*, *Hakea lissocarpa*, *Hibbertia commutata* and a few *Corymbia calophylla*.

Loc.: Forest monitoring plot FC30, Godfrey forest block, 2.1 km along Steed Road from Ernie Road, then left along shunt,

Lat. 33° 16' 2"S **Long.** 116° 25' 25"E (WGS84)

Coll. B.G. Ward FC671 **Date:** 05/10/2004

Voucher: Forestcheck Monitoring Program.

PERTH 6643329

WESTERN AUSTRALIAN HERBARIUM, PERTH Flora of Western Australia

Trymalium ledifolium Fenzl

Rhamnaceae

Identified by:

Erect compact perennial shrub, height to 1 m, width to 90 cm. Population flowering 100%. State forest; hill slope; gravelly brown clayey sand. occasional. Forest with associated vegetation of *Eucalyptus marginata*, *Allocasuarina fraseriana*, *Macrozamia riedlei*, *Hakea lissocarpa*, *Hibbertia commutata* and a few *Corymbia calophylla*.

Loc.: Forest monitoring plot FC30, Godfrey forest block, 2.1 km along Steed Road from Ernie Road, then left along shunt,

Lat. 33° 16' 2"S **Long.** 116° 25' 25"E (WGS84)

Coll. B.G. Ward FC671 **Date:** 05/10/2004

Voucher: Forestcheck Monitoring Program.

PERTH 6643329

WESTERN AUSTRALIAN HERBARIUM, PERTH Flora of Western Australia

Acacia pulchella R.Br.

Mimosaceae

Identified by:

Erect compact pungent perennial shrub, height to 70 cm, width to 40 cm; flowers yellow. Population flowering 10%. State forest; hill slope; gravelly brown clayey sand. frequent. Forest with associated vegetation of *Eucalyptus marginata*, *Allocasuarina fraseriana*, *Macrozamia riedlei*, *Hakea lissocarpa*, *Hibbertia commutata* and a few *Corymbia calophylla*.

Loc.: Forest monitoring plot FC30, Godfrey forest block, 2.1 km along Steed Road from Ernie Road, then left along shunt,

Lat. 33° 16' 2"S **Long.** 116° 25' 25"E (WGS84)

Coll. B.G. Ward FC672 **Date:** 05/10/2004

Voucher: Forestcheck Monitoring Program.

PERTH 6643310

Appendix C. Example of Cryptogam Label generated in Max.

WESTERN AUSTRALIAN HERBARIUM, PERTH
Flora of Western Australia

Rhynchohostegium tenuifolium var. *tenuifolium*

Brachytheciaceae

Identified by: R.J. Cranfield 25/02/2005

Moss: growing in sheltered wet positions on stone in ground layer. Littered brown gravelly clay soil with 5% or less outcropping of laterite, pH7. Occasional on isolated sites. Forest with associated vegetation of *Eucalyptus marginata*, *Corymbia calophylla*, *Allocasuarina fraseriana*, *Trymalium ledifolium*, *Hakea lissocapha*.

Loc.: Forest monitoring plot 36, 800 m on shunt of Belmont Road, 1km S of intersection with Alamo Road, Bell Forest Block

Lat. 32° 59' 53.2"S **Long.** 116° 21' 18.3"E (WGS84)

Coll. R.J. Cranfield 20970 **Date:** 12/10/2004

Voucher: Forest monitoring program.

WESTERN AUSTRALIAN HERBARIUM, PERTH
Flora of Western Australia

Cladonia rigida

Cladoniaceae

Identified by: R.J. Cranfield 23/02/2005

Lichen: active growth phase; fruticose; growing in sheltered wet positions on organic material in ground layer. Chemistry: cortex K+ yellow. Littered brown gravelly clay soil with 5% or less outcropping of laterite, pH7. Frequent on occasional sites. Forest with associated vegetation of *Eucalyptus marginata*, *Corymbia calophylla*, *Allocasuarina fraseriana*, *Trymalium ledifolium*, *Hakea lissocapha*.

Loc.: Forest monitoring plot 36, 800 m on shunt of Belmont Road, 1km S of intersection with Alamo Road, Bell Forest Block

Lat. 32° 59' 53.2"S **Long.** 116° 21' 18.3"E (WGS84)

Coll. R.J. Cranfield 20971 **Date:** 12/10/2004

Voucher: Forest monitoring program.

WESTERN AUSTRALIAN HERBARIUM, PERTH
Flora of Western Australia

Campylopus introflexus

Dicranaceae

Identified by: R.J. Cranfield 28/02/2005

Lichen: active growth phase; growing in sheltered wet positions on soil in ground layer. Littered brown gravelly clay soil with 5% or less outcropping of laterite, pH7. Frequent on frequent sites. Forest with associated vegetation of *Eucalyptus marginata*, *Corymbia calophylla*, *Allocasuarina fraseriana*, *Trymalium ledifolium*, *Hakea lissocapha*.

Loc.: Forest monitoring plot 36, 800 m on shunt of Belmont Road, 1km S of intersection with Alamo Road, Bell Forest Block

Lat. 32° 59' 53.2"S **Long.** 116° 21' 18.3"E (WGS84)

Coll. R.J. Cranfield 20972 **Date:** 12/10/2004

Voucher: Forest monitoring program.

WESTERN AUSTRALIAN HERBARIUM, PERTH
Flora of Western Australia

Usnea subciliata

Usneaceae

Identified by: R.J. Cranfield 25/02/2005

Lichen: active growth phase; filamentose; growing in sheltered wet positions on live and dead bark in tree layer. Littered brown gravelly clay soil with 5% or less outcropping of laterite, pH7. Occasional on occasional sites. Forest with associated vegetation of *Eucalyptus marginata*, *Corymbia calophylla*, *Allocasuarina fraseriana*, *Trymalium ledifolium*, *Hakea lissocapha*.

Loc.: Forest monitoring plot 36, 800 m on shunt of Belmont Road, 1km S of intersection with Alamo Road, Bell Forest Block

Lat. 32° 59' 53.2"S **Long.** 116° 21' 18.3"E (WGS84)

Coll. R.J. Cranfield 20973 **Date:** 12/10/2004

Voucher: Forest monitoring program.

Appendix D. Example of File Report generated in Max showing unique bar code allocated to each specimen.

10/08/2005		Example of Max Report showing unique barcode number			1	
COLLECTSHEETNO	FAMILY	GENUS	SPECIES	INFSP_TYP	INFSP_NAME	
FC670	6643337	Stackhousiaceae	Stackhousia	monogyna		
FC671	6643329	Rhamnaceae	Trymalium	ledifolium		
FC671	6643329	Rhamnaceae	Trymalium	ledifolium		
FC672	6643310	Mimosaceae	Acacia	pulchella		
FC674	6643299	Proteaceae	Dryandra	lindleyana		
FC675	6643280	Droseraceae	Drosera	huegelii		
FC676	6643272	Droseraceae	Drosera	menziesii	subsp. menziesii	
FC677	6643264	Proteaceae	Conospermum	caeruleum		
FC678	6643434	Casuarinaceae	Allocasuarina	fraseriana		
FC679	6643426	Papilionaceae	Daviesia	preissii		
FC680	6643418	Goodeniaceae	Lechenaultia	biloba		
FC682	6643388	Epacridaceae	Leucopogon	capitellatus		
FC684	6643353	Dilleniaceae	Hibbertia	amplexicaulis		
FC685	6643345	Proteaceae	Dryandra	bipinnatifida		
FC686	6643515	Tremandraceae	Tetratheca	hirsuta		
FC687	6643507	Epacridaceae	Astroloma	pallidum		
FC688	6643493	Thymelaeaceae	Pimelea	suaveolens	subsp. suaveolens	
FC689	6643485	Dasyopogonaceae	Lomandra	caespitosa		
FC690	6643477	Epacridaceae	Styphelia	tenuiflora		
FC691	6643469	Proteaceae	Petrophile	striata		
FC692	6643450	Loganiaceae	Logania	serpyllifolia		
FC693	6643442	Mimosaceae	Acacia	browniana		
FC694	6643604	Proteaceae	Synaphea	petiolaris		
FC695	6643590	Asteraceae	Senecio	hispidulus		
FC696	6643582	Orchidaceae	Cyanicula	sericea		
FC697	6643574	Orchidaceae	Caladenia	flava		
FC698	6643566	Orchidaceae	Elythranthera	brunonis		
FC699	6643558	Goodeniaceae	Dampiera	linearis		
FC700	6643531	Iridaceae	Patersonia	babianoides		
FC701	6643523	Anthericaceae	Chamaescilla	corymbosa		
FC702	6643698	Iridaceae	Patersonia	juncea		
FC703	6643671	Dilleniaceae	Hibbertia	commutata		
FC704	6643663	Asteraceae	Rhodanthe	citrina		
FC705	6643655	Goodeniaceae	Velleia	trinervis		
FC706	6643647	Anthericaceae	Sowerbaea	laxiflora		
FC707	6643639	Euphorbiaceae	Phyllanthus	calycinus		
FC708	6643620	Papilionaceae	Bossiaea	ornata		
FC709	6643612	Apiaceae	Xanthosia	huegelii		
FC710	6643787	Rutaceae	Boronia	crenulata		
FC711	6643779	Restionaceae	Desmocladius	fasciculatus		
FC712	6643760	Droseraceae	Drosera	stolonifera		
FC712A	6643752	Dilleniaceae	Hibbertia	spicata		
FC713	6643744	Olacaceae	Olax	benthamiana		
FC714	6643736	Asteraceae	Craspedia	variabilis		
FC715	6643728	Asteraceae	Hyalosperma	cotula		
FC716	6643701	Haemodoraceae	Conostylis	setigera		
FC717	6643876	Haemodoraceae	Conostylis	aculeata		
FC718	6643868	Haemodoraceae	Conostylis	serrulata		
FC719	6643841	Zamiaceae	Macrozamia	riedlei		

* = Alien species

Appendix E. Example of Specimen Table generated in Max.

11/08/2005

Taxon Name	NameID	SpeciesCode	Lifeform	Lifestyle	Fire Resp
<i>Acacia alata</i>	3207	ACAALA	S	P	A1
<i>Acacia browniana</i>	3247	ACABRO	S	P	A1
<i>Acacia celastrifolia</i>	3254	ACACEL	S	P	A1
<i>Acacia drummondii</i>	3311	ACADRU	S	P	A1
<i>Acacia drummondii</i> subsp. <i>drummondii</i>	11661	ACADRUDR	S	P	A1
<i>Acaena echinata</i>	3184	ACAECH	DS	P	A1
<i>Acacia extensa</i>	3331	ACAEXT	S	P	A1
<i>Acacia myrtifolia</i>	3453	ACAMYR	S	P	A1
<i>Acacia preissiana</i>	3496	ACAPRE	DS	P	A1
<i>Acacia pulchella</i>	3502	ACAPUL	S	P	A1
<i>Acacia saligna</i>	3527	ACASAL	S	P	A1
<i>Actinotus glomeratus</i>	6203	ACTGLO	DS	P	A1
<i>Agrostocrinum scabrum</i>	1261	AGRSCA	G	P	B3
<i>Aira cupaniana</i>	185	AIRCUP	GR	A	A1
<i>Allocastrum fraseriana</i>	1728	ALLFRA	T	P	B1
<i>Anigozanthos manglesii</i>	1411	ANIMAN	S	P	B3
<i>Arctotheca calendula</i>	7838	ARCCAL	H	A	A1
<i>Astroloma ciliatum</i>	6323	ASTCIL	DS	P	B2
<i>Astroloma drummondii</i>	6325	ASTDRU	DS	P	B2
<i>Astroloma pallidum</i>	6334	ASTPAL	DS	P	B2
<i>Austrodanthonia caespitosa</i>	17950	AUSCAE	GR	P	B3
<i>Austrostipa campylachne</i>	17233	AUSCAM	GR	P	B3
<i>Banksia grandis</i>	1819	BANGRA	T	P	A2
<i>Banksia sphaerocarpa</i>	1851	BANSPH	S	P	A2
<i>Billardiera variifolia</i>	3165	BILVAR	V	P	A1
<i>Boronia crenulata</i>	4413	BORCRE	DS	P	A1
<i>Boronia gracilipes</i>	4422	BORGRA	DS	P	A1
<i>Boronia ramosa</i>	4438	BORRAM	S	P	A1
<i>Boronia spathulata</i>	4441	BORSPA	S	P	B2
<i>Bossiaea eriocarpa</i>	3710	BOSERI	S	P	A1
<i>Bossiaea ornata</i>	3714	BOSORN	S	P	A1
<i>Briza minor</i>	245	BRIMIN	GR	A	A1
<i>Burchardia umbellata</i>	1387	BURUMB	G	P	B3
<i>Caesia micrantha</i>	1276	CAEMIC	G	P	B3
<i>Caladenia flava</i>	1592	CALFLA	G	P	B3
<i>Caladenia macrostylis</i>	1604	CALMAC	G	P	B3
<i>Caladenia reptans</i>	1613	CALREP	G	P	B3
<i>Caladenia splendens</i>	15380	CALSPL	G	P	B3
<i>Cassytha racemosa</i>	2957	CASRAC	P	P	A1
<i>Centaurium erythraea</i>	6539	CENERY	H	A	A1
<i>Centrolepis polygyna</i>	1134	CENPOL	H	A	A1
<i>Chamaescilla corymbosa</i>	1280	CHACOR	G	P	B3
<i>Clematis pubescens</i>	2929	CLEPUB	V	P	A1
<i>Comesperma calymega</i>	4550	COMCAL	DS	P	B2
<i>Conostylis aculeata</i>	1418	CONACU	DS	P	B3
<i>Conospermum caeruleum</i>	1862	CONCAE	S	P	A1
<i>Conostylis serrulata</i>	1453	CONSER	DS	P	B2
<i>Conostylis setigera</i>	1454	CONSET	DS	P	B3
<i>Corymbia calophylla</i>	17104	CORCAL	T	P	A2
<i>Craspedia variabilis</i>	13354	CRAVAR	G	P	B3