

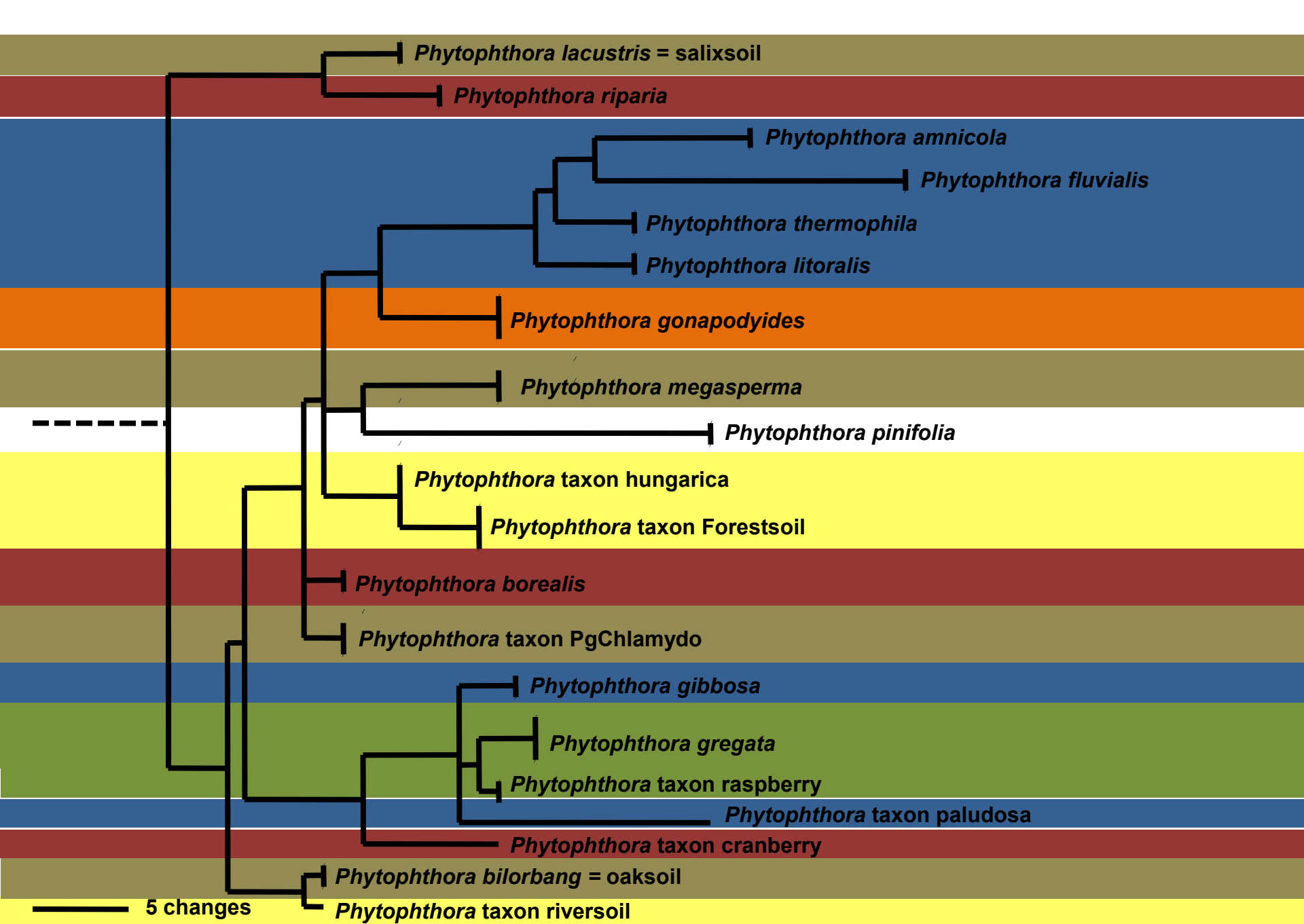
# Characterization of *Phytophthora* hybrids from ITS clade 6 associated with riparian ecosystems in South Africa and Australia

Treena Burgess, Jan Nagel, Giles Hardy, Mike Stukely and Mike Wingfield

## Promiscuity, fertility and survival of ITS clade 6 hybrids associated with riparian ecosystems in Western Australia

Treena Burgess, Daniel Hüberli, Diane White, Mike Stukely and Giles Hardy





- in 2008 when conducting molecular characterisation of the VHS collection of the Department of Conservation we had several isolates that could not be identified using conventional sequencing of the ITS region because of numerous sites exhibiting additivity (double peaks) and severely truncated sequence data.
- at the time we thought they may be hybrids
- in 2009, Daniel Hüberli conducted his 'Fishing for Phytophthora' survey in 40 natural rivers and waterways in the south-west of Western Australia. Over half the ITS sequences failed
- at the same time Bill Dunstan conducted a survey along a proposed pipeline within natural forest in Victoria and Tim Rudman sampled highland rivers in Tasmania.
- from this we ended up with a large set of isolates that could not be identified by ITS sequence, were fast growing, had high temperature optima and maxima and were sterile.



- the next step was then to commence molecular characterisation.
- the ITS region was cloned and 10-50 amplicons sequenced for each isolate
- the *coxI* mitochondrial gene region was amplified and if any evidence of additivity it was cloned before sequencing
- the single copy genes used in the *P. alni* study of Ioos et al (2006) were all tested using a panel of known species from ITS clade 6, some primers were redesigned and finally we had reliable amplification using ASF-like (anti-silencing factor) and GPA1 (G-protein alpha subunit). Both regions were cloned and 6-12 amplicons sequenced



- during research collaboration in South Africa I was asked to assist a Jan Nagel who was working on a 'Fishing for Phytophthora' project
- he had many isolated exhibiting additivity in the ITS region and he had cloned and sequenced ITS amplicons and also *coxI*
- I realised that he had some of the same 'hybrid' types as I had found in Australia.
- this discovery actually simplified the story as I split the Australian hybrid isolates into two groups;
  1. those predominately from eastern Australia (and RSA)
  2. those exclusively from Western Australia









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THERMOPHILA	T	A	C	T	G	G	C	-	A	A	A	C	G	T	C	T	A	G	A	C	C	T	
A-T	Y	W	Y	Y				N		W		S	K			Y		R		Y	Y		12

	n	41	44	59	71	110	148	162	171	180	184	457	476	477	518	519	555	699	746	748	753	788	815
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PG-T

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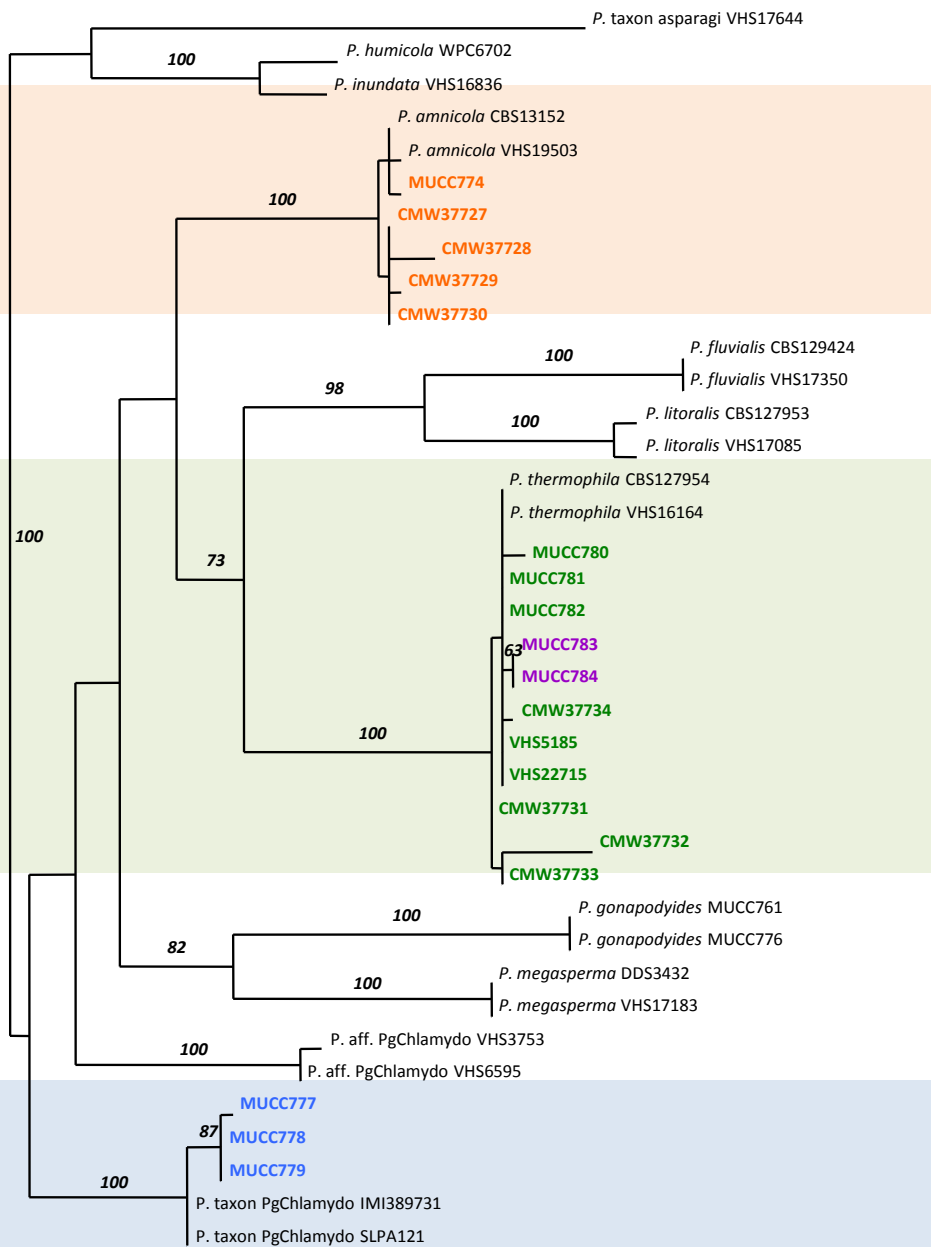
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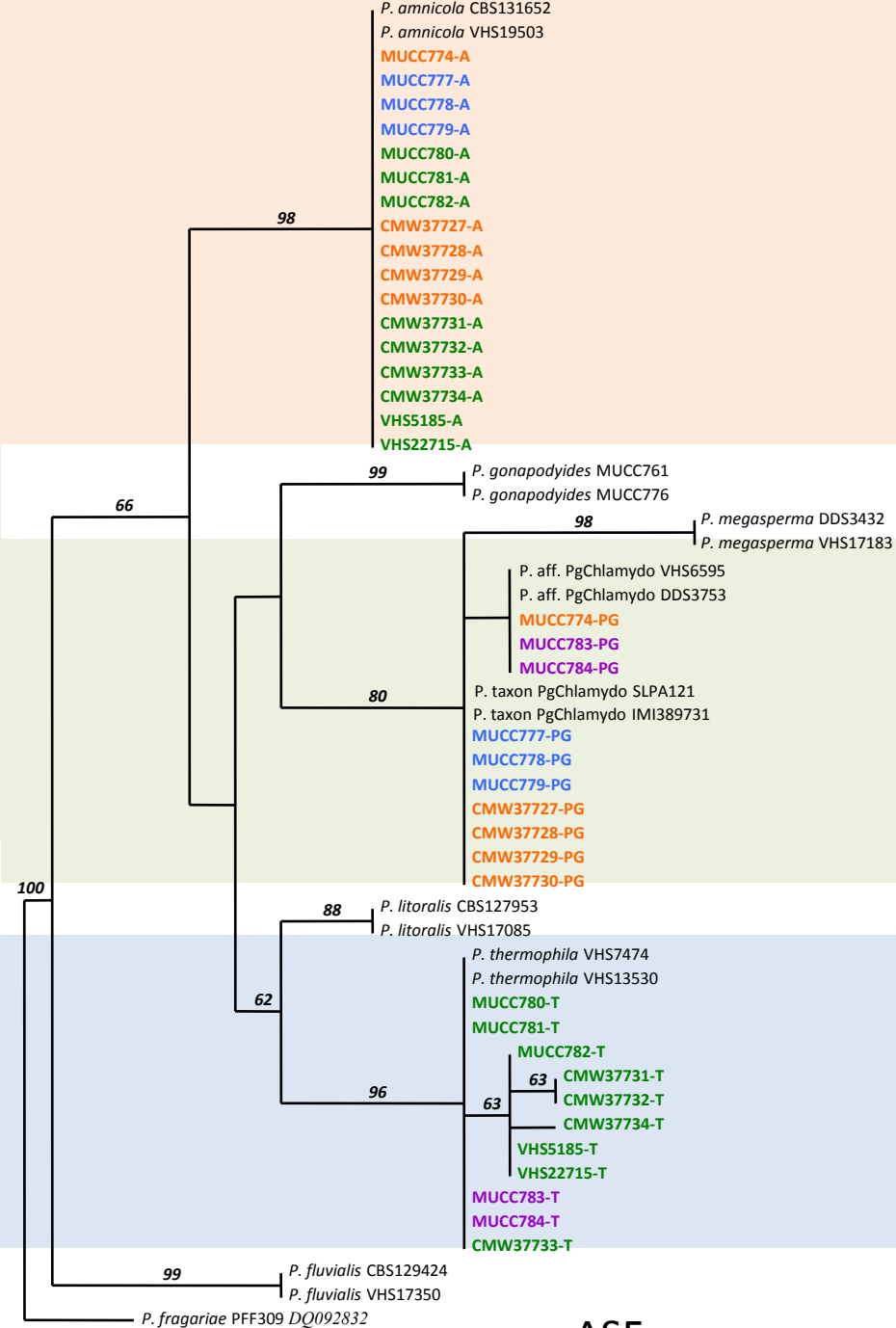
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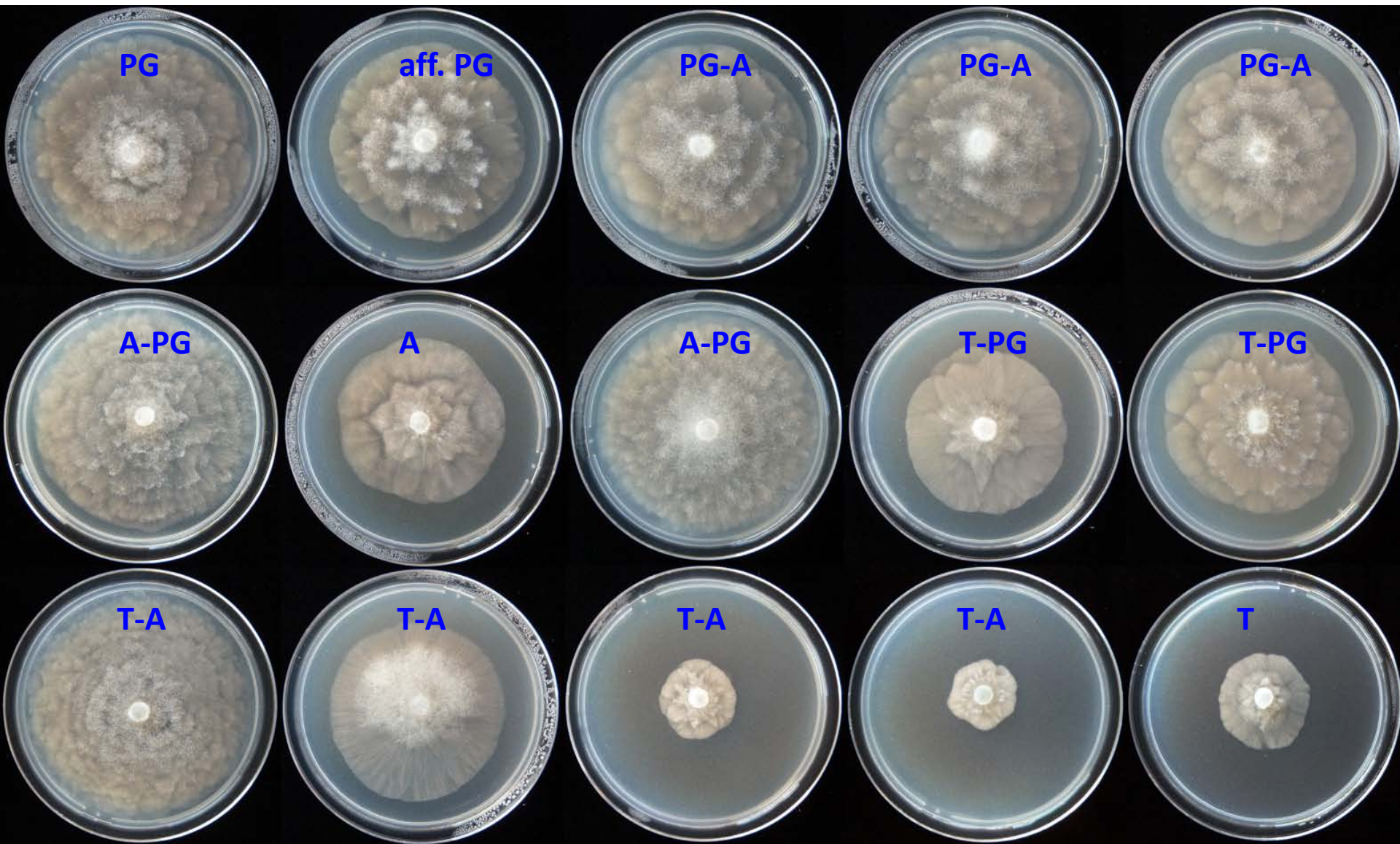
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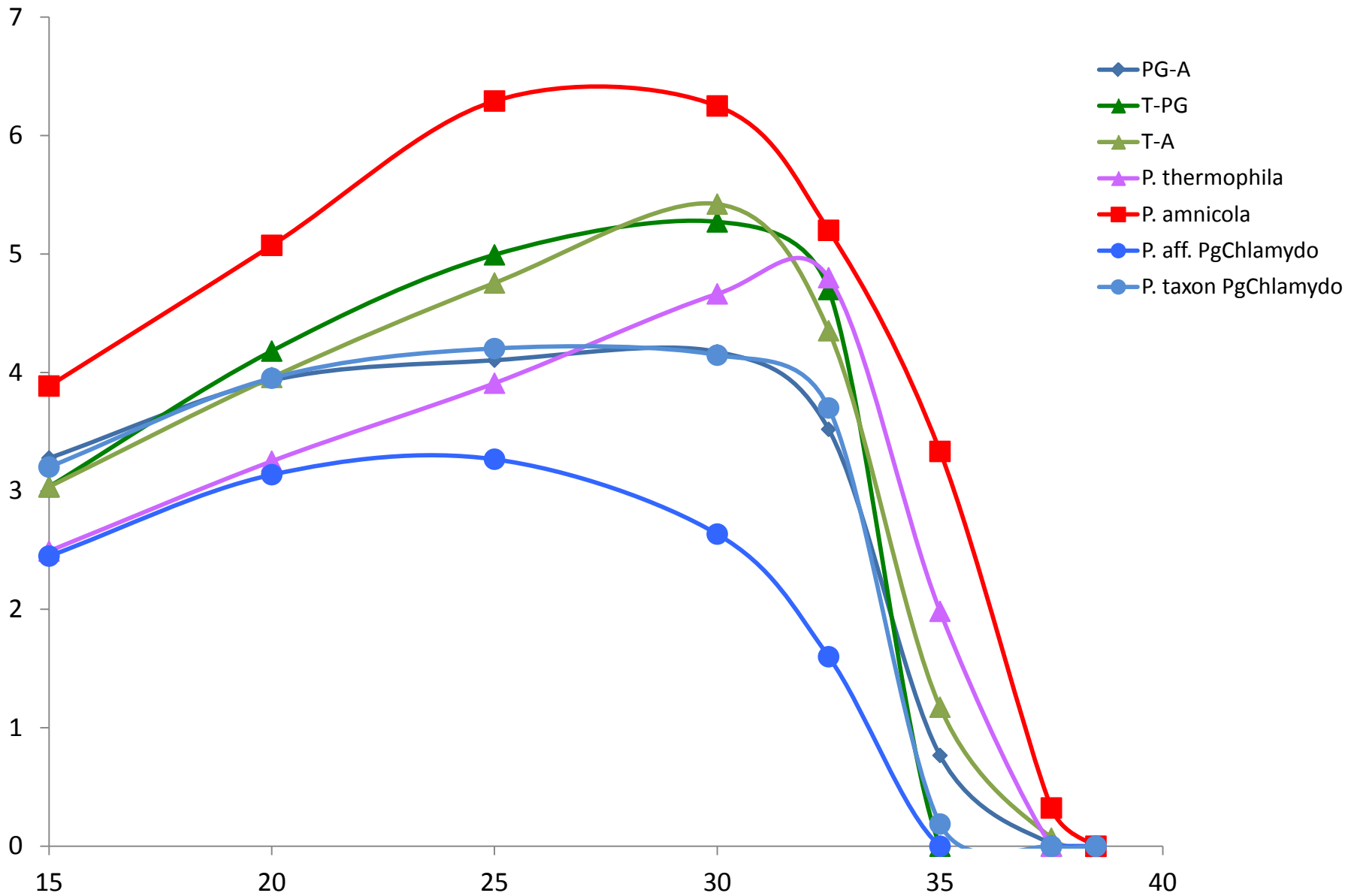
COX



ASF







	<b>P. thermophila</b>	<b>P. amnicola</b>	<b>P. PgChlamydo</b>	<b>PG-A</b>	<b>A-PG</b>	<b>T-A</b>	<b>T-PG</b>
<b>Sporangia shape</b>	ovoid	ovoid	ovoid	ovoid	ovoid	ovoid	limoniform
<b>Sporangia size</b>	45 x25	62 x35	57 x 35	56 x 34	39 x 27	48 x 30	48 x 31
<b>Sexual system</b>	Sterile or silent homothallic	sterile ?	sterile ?	sterile	sterile	sterile	sterile
<b>swellings</b>	Globose or elongated, partly catenulate	Ellipsoid to irregular catenulate hyphal swellings in clusters	globose	globose	No swellings	No swellings	No swellings
<b>chlamydo-spores</b>	Globose, radiating hyphae		Globose, radiating hyphae	globose			
<b>Temp max</b>	35	37.5	35	>35<37.5	>35<37.5	>35<37.5	35

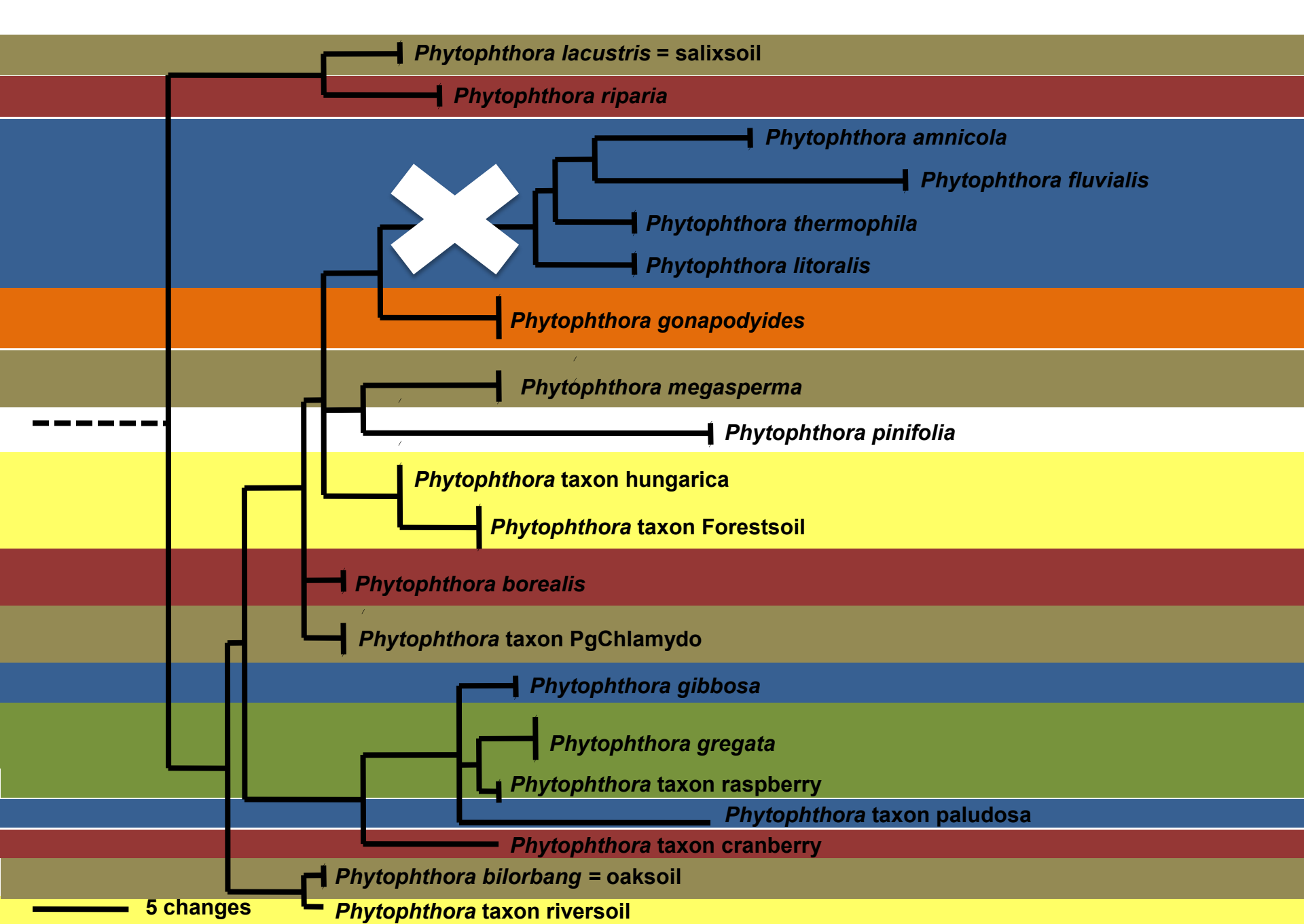


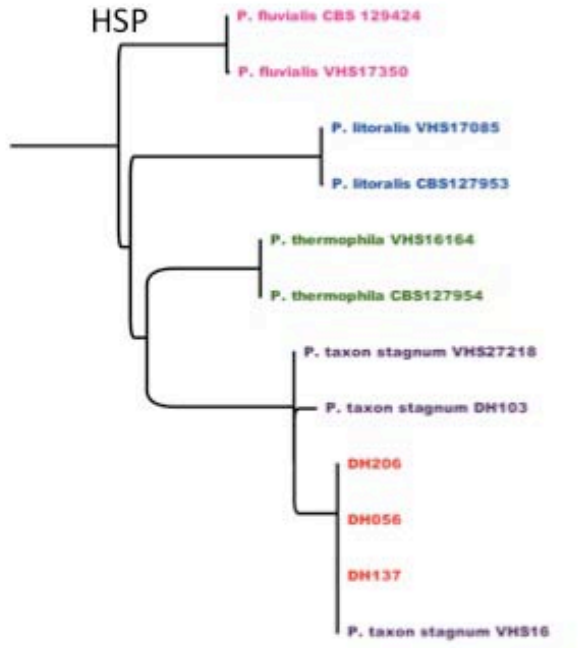
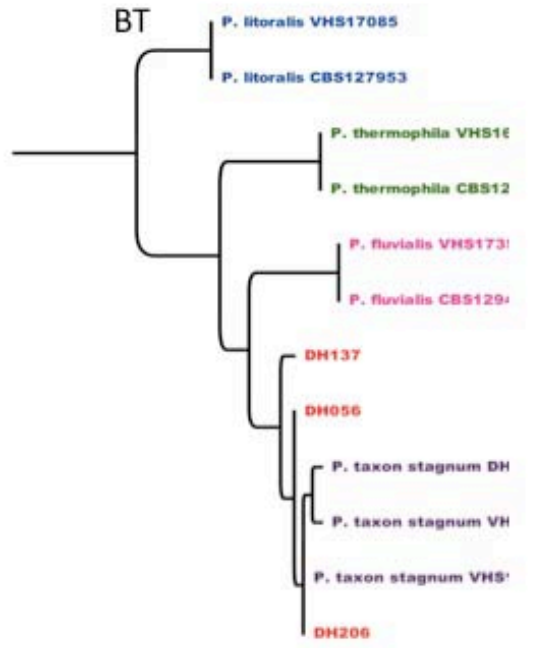
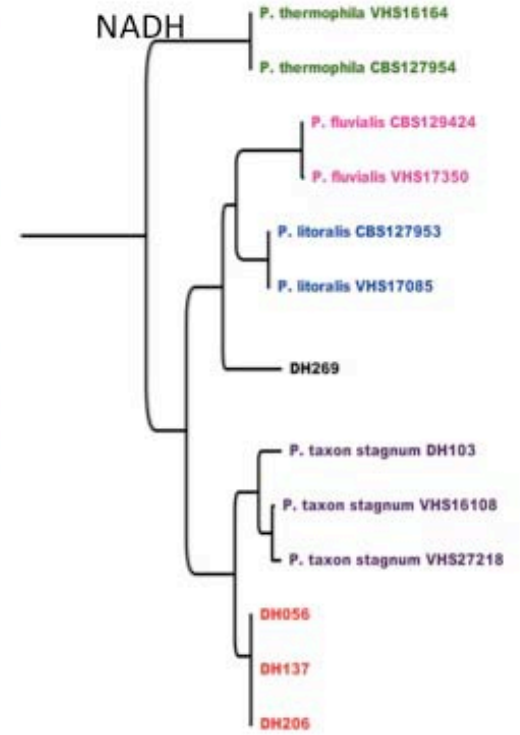
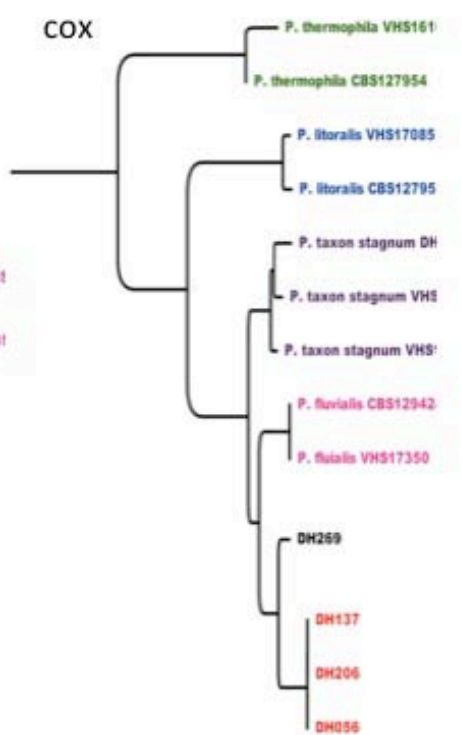
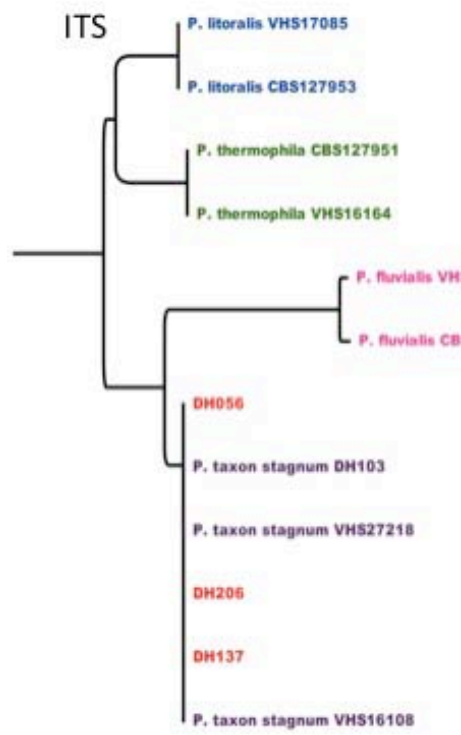
	WA	EA	RSA
<i>P. thermophila</i>	+++	+++	
<i>P. amnicola</i>	+++	?	
P. PgChlamydo		+++	
P. aff. PgChlamydo	+++	?	+++
PG-A		+++ (VIC)	+++
A-aff. PG		+++ (TAS)	
T-A	+++	+++ (TAS)	+++
T-aff. PG		+++ (TAS)	

parental species and all hybrids in Australia

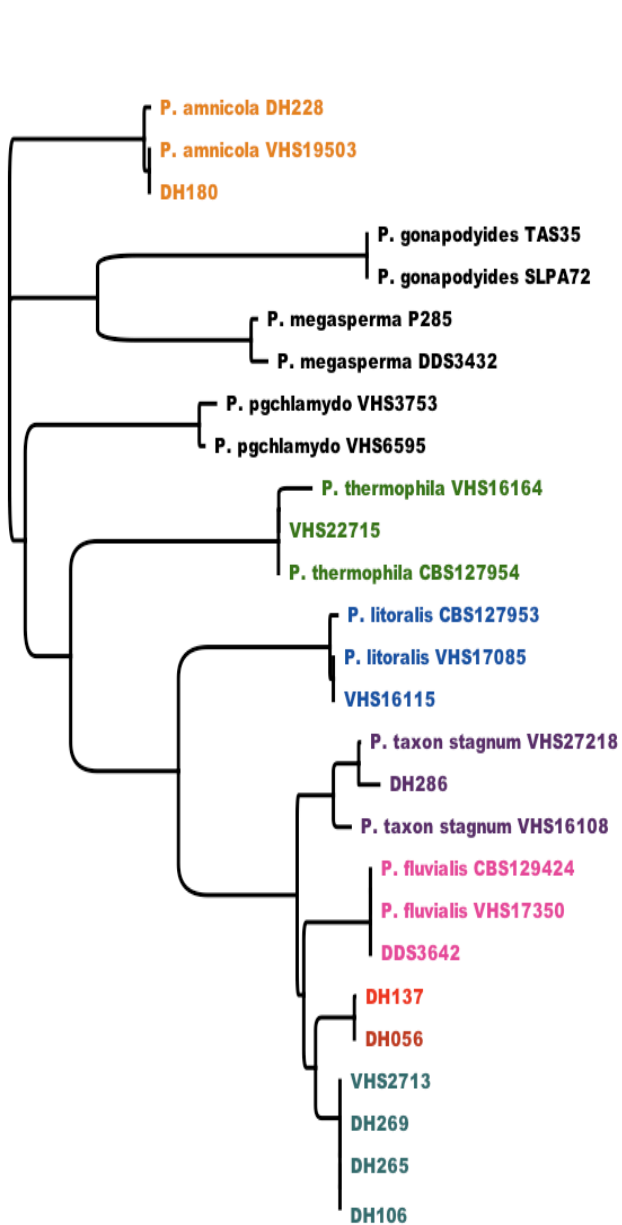
- In general, hybrid formation appears to be through sexual cross between two parental species. How?
- PgChlamydo and *P. amnicola* both appear to be sterile when mated with tester strains, but one isolate *P. thermophila* formed oospores after being left in soil extract. Are we unable to induce the right conditions in laboratory?
- all parental species and hybrids common in waterways.
  1. Is there a hybrid swarm perhaps through somatic hybridisation between sterile hybrids?
  2. can they back cross with parental species?
  3. or are hybrids constantly formed by new hybridisation events between parental species, but die out if conditions are unfavourable for their growth and survival
- how did they get to South Africa? are they found elsewhere in the world?

- these hybrids between amnicola, thermophila and PgChlamydo are widespread in all waterways examined to date in Australia and South Africa (except Western Australia)
- I then excluded these hybrids from the matrix and examined only 'hybrid' isolates from Western Australia (approx 50 isolates-many more available)
- they have been isolated predominantly from waterways BUT they have also been isolated in the rhizosphere soil of dying plants after inundation.
- for this presentation I have selected representative isolates of the different hybrid types

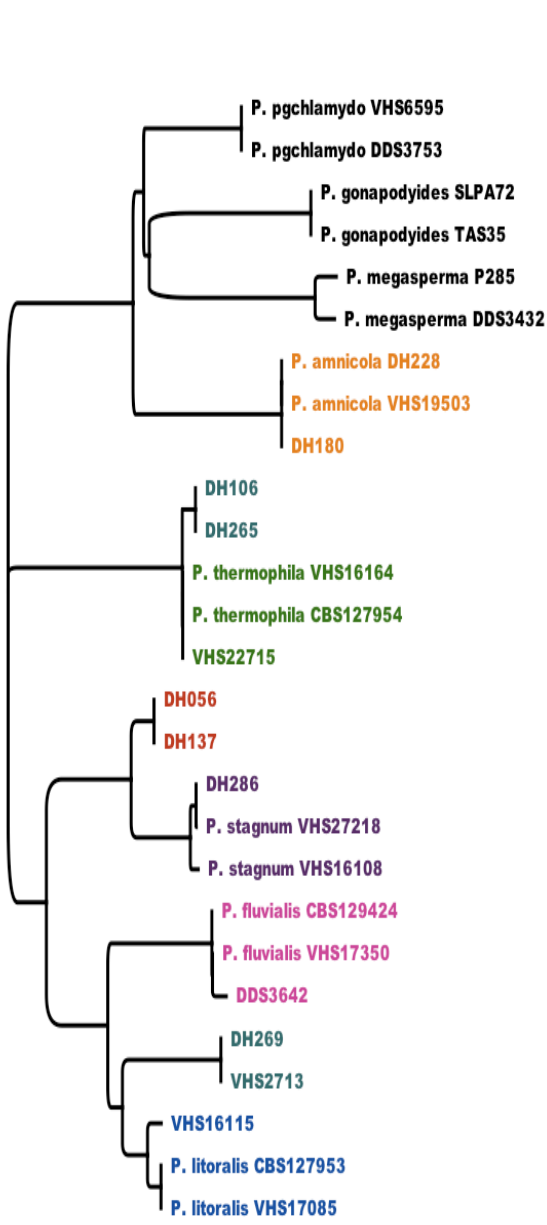






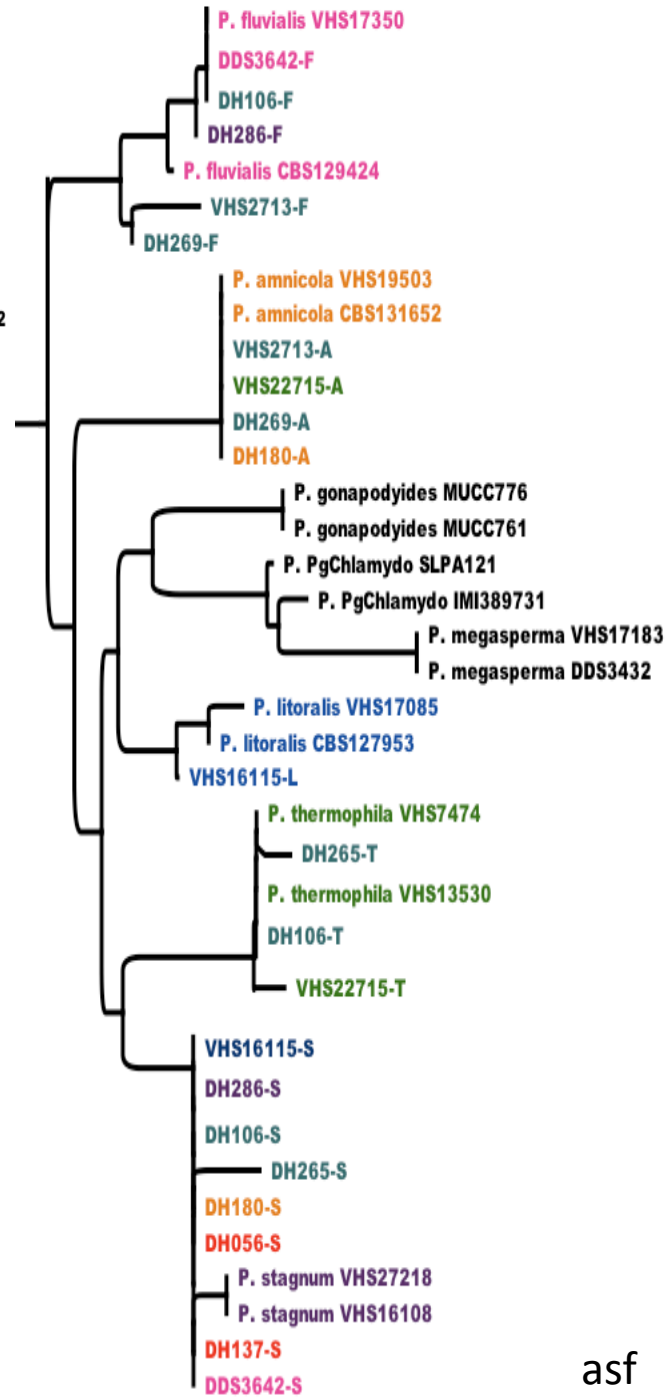


COX



0.0060

nadh



asf

	<b>ITS CLONES</b>	<b>COX ID</b>	<b>NADH</b>	<b>ASF</b>
VHS2713	A-S	2.5	2.5	A-F
269	A-S	2.5	2.5	A-F
VHS22715	T-A	<i>P. thermophila</i>	T	T-A
265	T-S	2.5	T	T-S
106	T-S	2.5	T	F-S-T
180	A-S	<i>P. amnicola</i>	A	A-S
286	S-F	<i>P. taxon stagnum</i>	S	S-F
DDS3642	F-S	<i>P. fluvialis</i>	F	F-S
VHS16115	S-L	<i>P. litoralis</i>	L	S-L
56	S	2.8	2.8	S
137	S	2.8	2.8	S

**images T. Jung**

- In WA we have evidence for hybrids forming with *P. amnicola*, *P. thermophila*, *P. litoralis*, *P. fluvialis* and *P. taxon stagnum* as parental species
- *cox1* and NADH data suggest there may be two additional species not yet identified
- hybrids formed are; A-S, T-A, L-S, S-F, F-S
- based on ASF there is also S-A and S-F, but *cox* types unknown
- based on ASF alleles corresponding to F-S-T, may be some sort of backcrossing (sterile?) or somatic hybridisation
- some isolates have ITS and ASF like *P. taxon stagnum* but unknown *cox* and NADH types
- possibilities to explain observations include non-inheritance of alleles from one parent or mitotically unstable progeny (hybrids)
- next step will be flow cytometry to assess genome size, microsatellites would also be useful but difficult to design over several genera

*P. gonapodyides* and *PgChlamydo* not found in Western Australia